



Universitat de Lleida

Identification of Transcriptional Responses in Root and Leaf of *Prunus persica* Under Conditions of Drought Stress Using RNA Sequencing

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Estación Experimental de Aula Dei (EEAD-CSIC)



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2- Objectives

3- Material and Methods

4- Results and Discussion

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1-Introduction

Family: Rosaceae Subfamily: Prunoidae Subgenus: Amygdalus

Genus: Prunus





Almond *P. dulcis* (Mill) D.A.

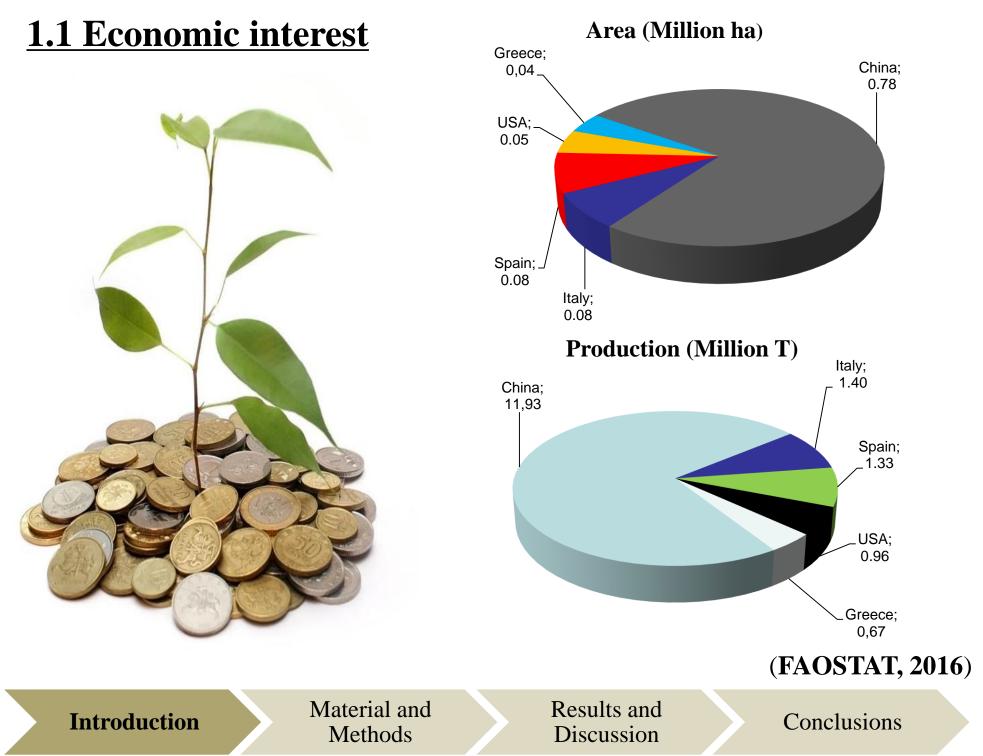
Apricot *P. armeniaca* L.



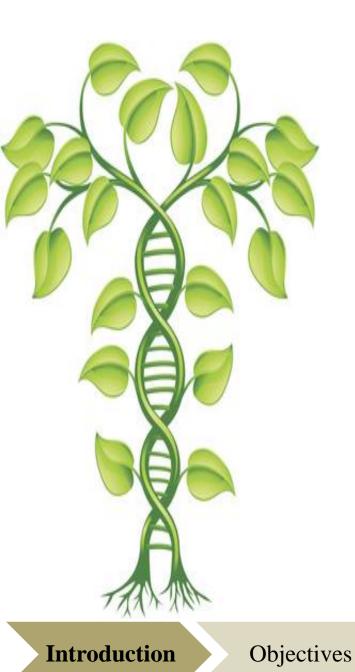
Plum *P. domestica* L.

Introduction

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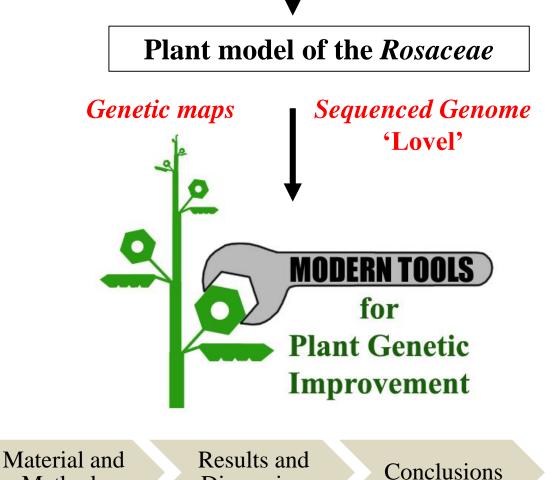


1.2 Genetic interest



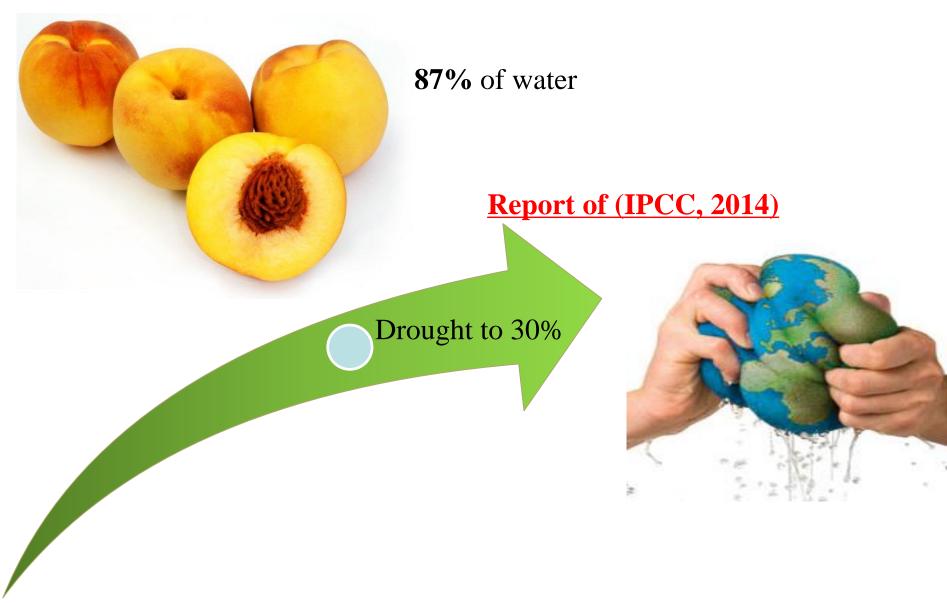
- Diploid species 2x = 2n = 16
- Small genome ≈ 230 Mbp
- Short juvenile phase
- Self-compatibility

Methods



Discussion

1.3 Importance of water for cultivation



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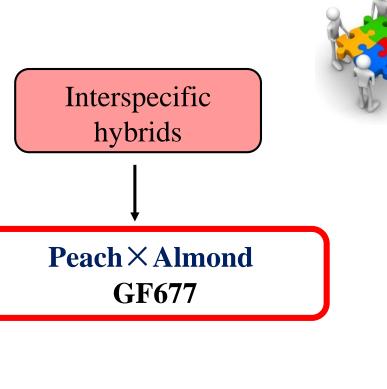
Material and Methods

Results and Discussion

1.4 Fruit rootstocks (Prunus)

Seedling rootstock





Introduction

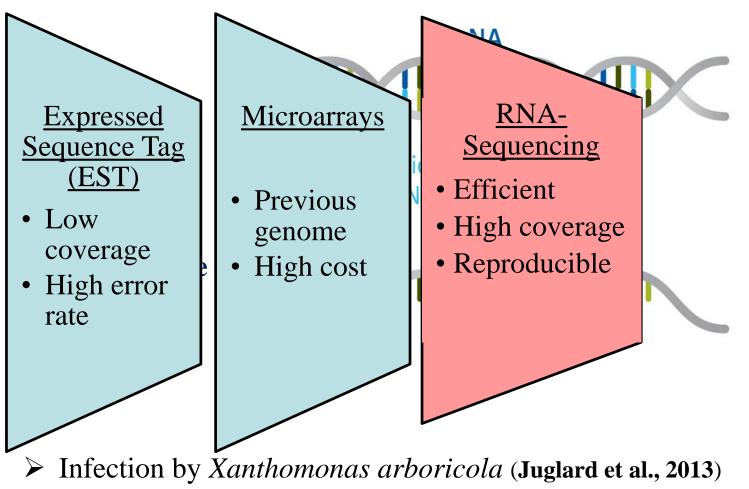
Objectives

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1.5 Influence of the rootstock on the variety

Vigour of the rootstock Root system well developed > The accumulation of osmolytes **Biochemical** Molecular Morphological Material and Results and Introduction Objectives Conclusions Methods Discussion

<u>1.6 Transcriptome analysis</u>



- Infection by Plum pox Virus en P. domestica (Rubio et al., 2014)
- Hypoxia in *P. persica* (Arismendi et al., 2015)

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2- Objectives

General objective

Investigate the molecular mechanisms involved in the response of *P*. *persica* to drought and identifying the associated candidate genes. **Specific objectives**

- Exploration of the transcriptomic variations between control and drought-affected plants using RNA-seq.
- ➤ Identification of the differentially expressed genes (DEGs).
- Annotation and functional classification of DEGs, then investigation of the metabolic pathways involved.
- > Validation of the reliability of the RNA-seq analysis by RT-qPCR.

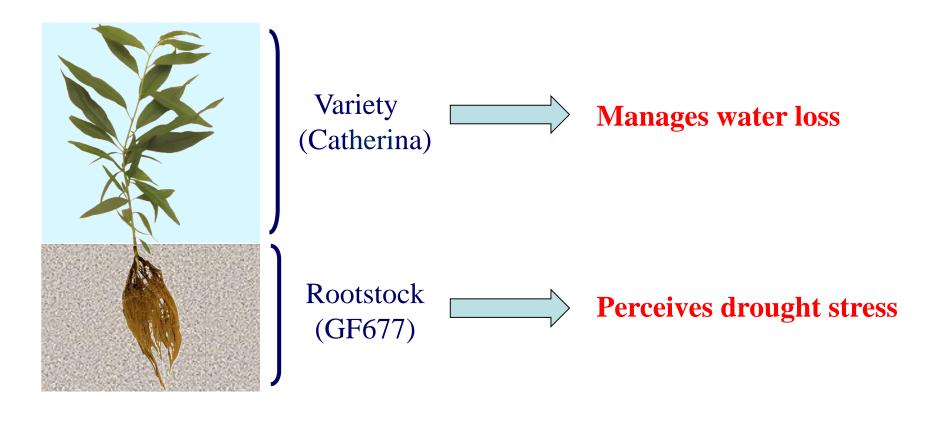
Introduction

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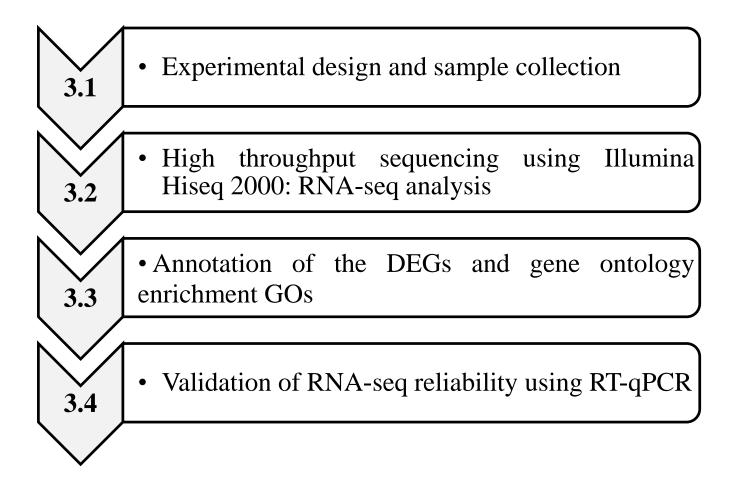
2.1 Originality of the research in peach

Characterization of transcriptional response to drought stress in root and leaf



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3- Methodology



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3.1 Plant material: rootstock GF677 and variety Catherina



Micropropagation

2 weeks pots 300 cm³ (peat)



Micrografted with 'Catherina'

- Temperature 23°C day / 18°C night
- 14 h light / 10 h dark



3 weeks 30 plants

pots 15 L peat : sand (1:1)

(Jiménez et al., 2013)

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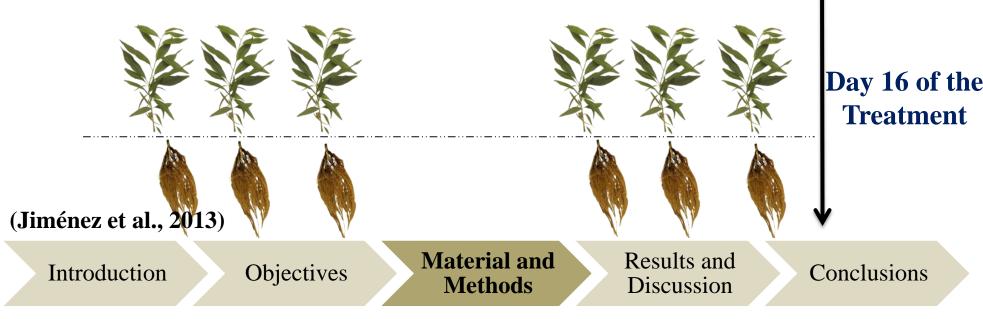
3.2 Experimental design and sample collection



Drought (80% of evapotranspiration)

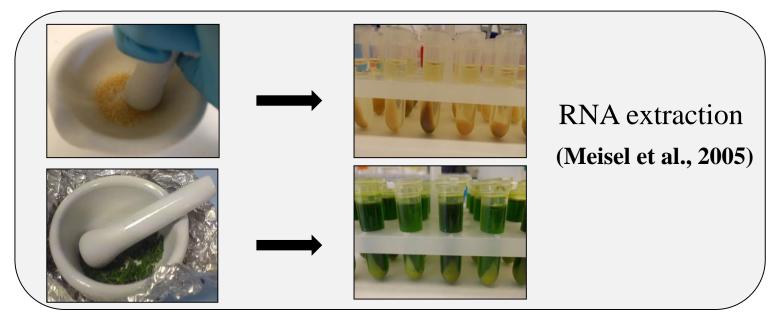
Control

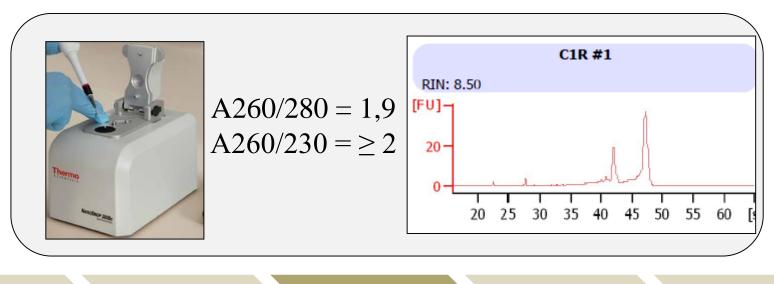
Measurements of physiological parameters Days (0, 7, 12, 16) (Jiménez et al., 2013)



Start of Treatment

3.2 Experimental design and sample collection (II)





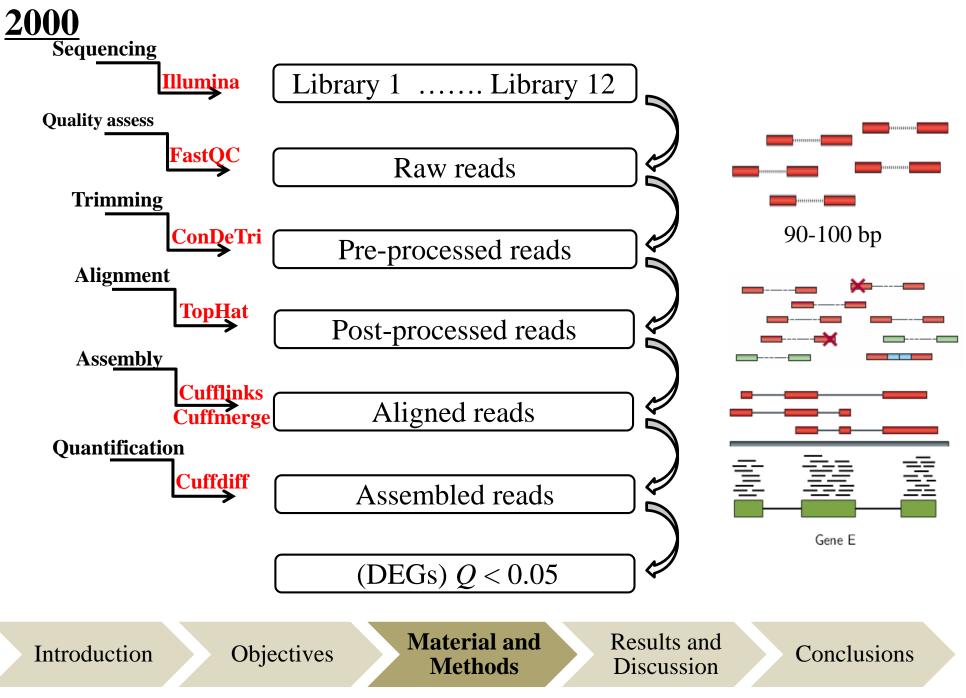
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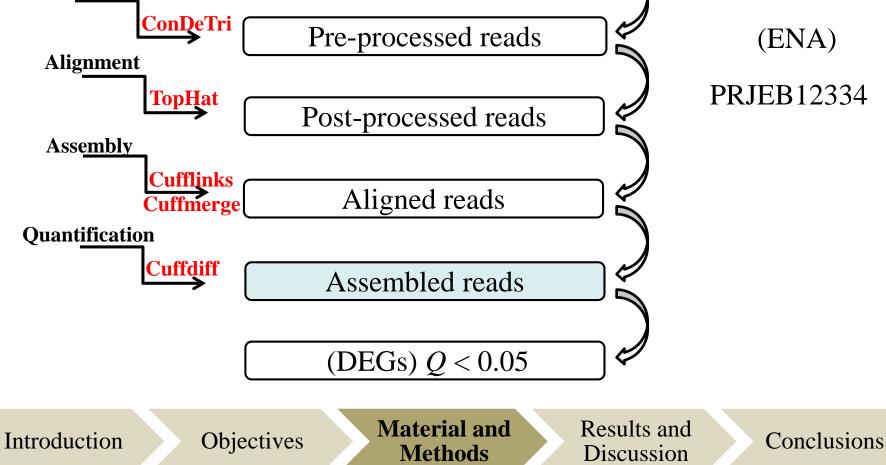
Material and Methods

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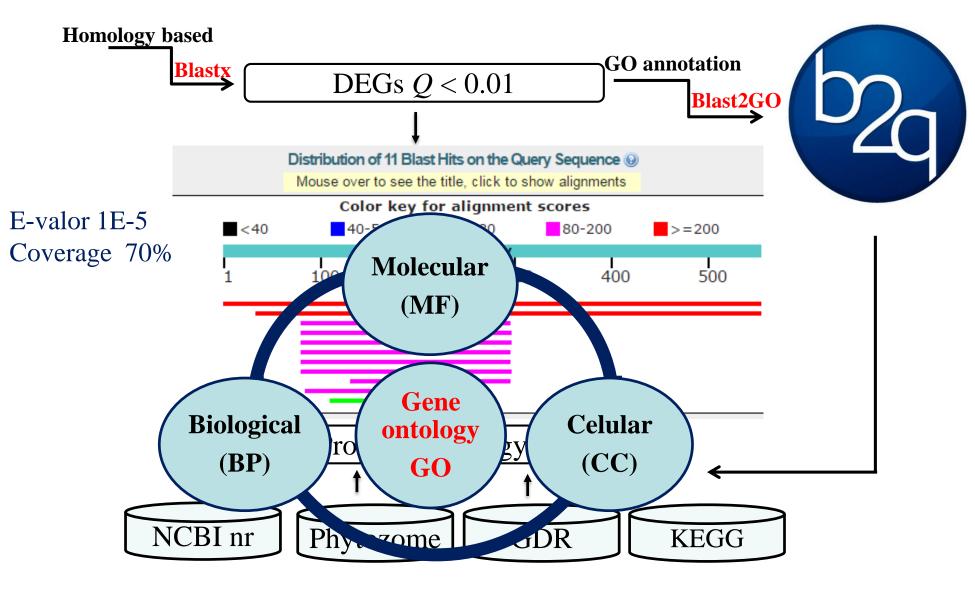
3.2 High throughput sequencing using Illumina Hiseq-



3.2 High throughput sequencing using Illumina Hiseq 2000 Sequencing Illumina Library 1 Library 12 Quality assess FastQC Raw reads Trimming ConDeTri Pre-processed reads (ENA)



3.3 Annotation of the DEGs and GO enrichment

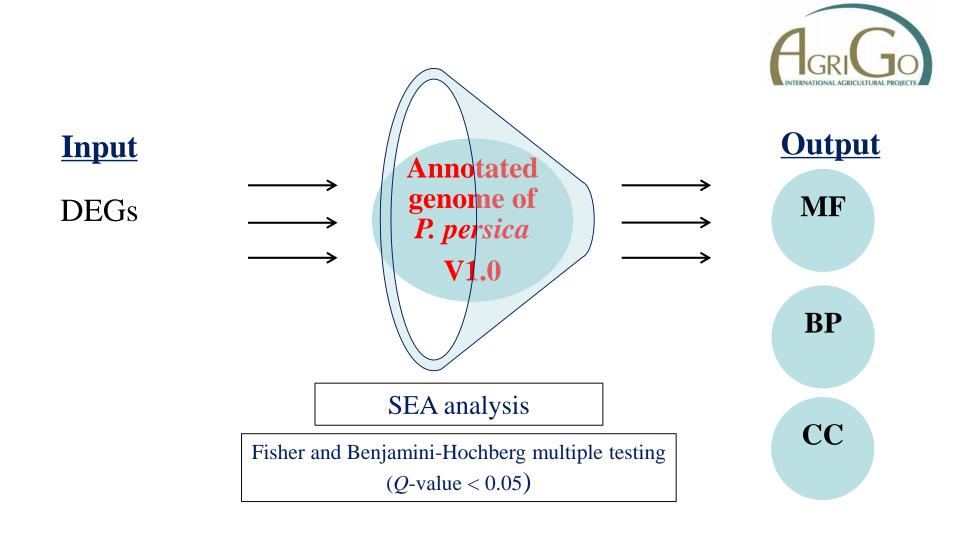


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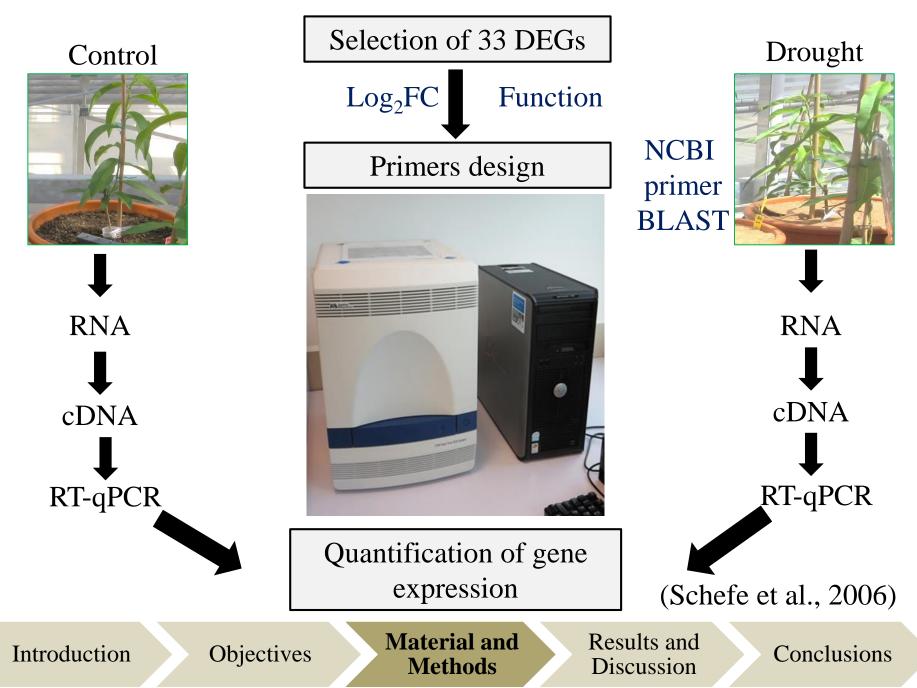
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3.3 Annotation of the DEGs and GO enrichment (II)





3.4 Validation of RNA-seq reliability using RT-qPCR



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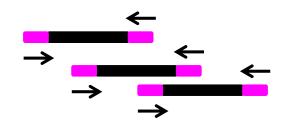
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4.1 Physiological responses to drought stress

		16 days of drought			 > Wilting > Defoliat (Jiménez et al.) 	tion
Treatments	SWC %	Ψs MDa	g_s	<i>Ci</i>	A_N	-
	70	MPa	mol H ₂ O m ⁻² s ⁻¹	µmol CO ₂ mol ⁻¹	μmol CO ₂ m ⁻² s ⁻¹	-'
Well-	26.63 ± 0.18^{b}	-0.73 ± 0.05^{b}	0.56 ± 0.03^{b}	277.30 ± 2.17^{b}	$20.82 \pm 0.54^{\rm b}$	
watered	20.03 ± 0.10	-0.75 ± 0.05	0.50 ± 0.05	211.30 ± 2.11	20.02 ± 0.04	
Water- deprived	10.69 ± 0.28^{a}	-1.08 ± 0.02^{a}	0.37 ± 0.03^{a}	261.27 ± 4.27^{a}	18.35 ± 0.42^{a}	Ļ
(Data are	e mean \pm SE o	f n=6 replicate	es)	(Rahm	ati et al., 2015))

Sequencing (Paired-end)

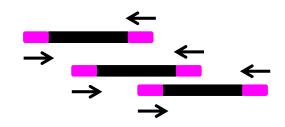


	Libraries	Raw reads Clean reads (M)			M)	Mapped reads (M)			
	Libraries	(M)	Paired	Unpaired	Total	Paired	Unpaired	Total	
ſ	RC 1	61.30	28.54	12.31	40.85	10.71	4.16	14.78	
	RC 2	25.52	10.10	5.15	15.25	7.83	4.07	11.90	
Root -	RC 3	22.69	8.48	4.75	13.23	7.03	3.95	10.98	
	RD 1	24.97	9.37	5.04	14.41	7.49	4.05	11.54	
	RD 2	24.49	7.76	5.47	13.23	5.89	4.29	10.18	
	RD 3	20.81	8.14	4.79	12.93	6.51	3.81	10.32	
ř	LC 1	23.12	8.72	5.44	14.16	7.14	4.57	11.71	
	LC 2	23.02	9.05	5.30	14.35	7.48	4.51	11.99	
	LC 3	22.30	8.01	4.76	12.77	6.63	4.07	10.70	
Leaf –	LD 1	22.00	8.33	4.42	12.75	6.77	3.75	10.84	
	LD 2	22.86	6.20	5.51	11.71	5.37	4.93	10.30	
	LD 3	22.00	8.63	4.33	12.96	7.20	3.78	10.38	
L	Total	315.08	121.33	67.27	188.60	86.05	49.94	135.99	

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Material and Methods Results and Discussion

Sequencing (Paired-end)



Results and

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	Libraria	Libraries Raw reads		ean reads (M)	Mapped reads (M)			
	Libraries	(M)	Paired	Unpaired	Total	Paired	Unpaired	Total	
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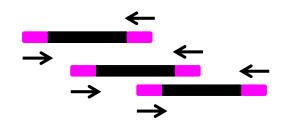
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Sequencing (Paired-end)

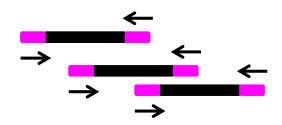


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Sequencing (Paired-end)



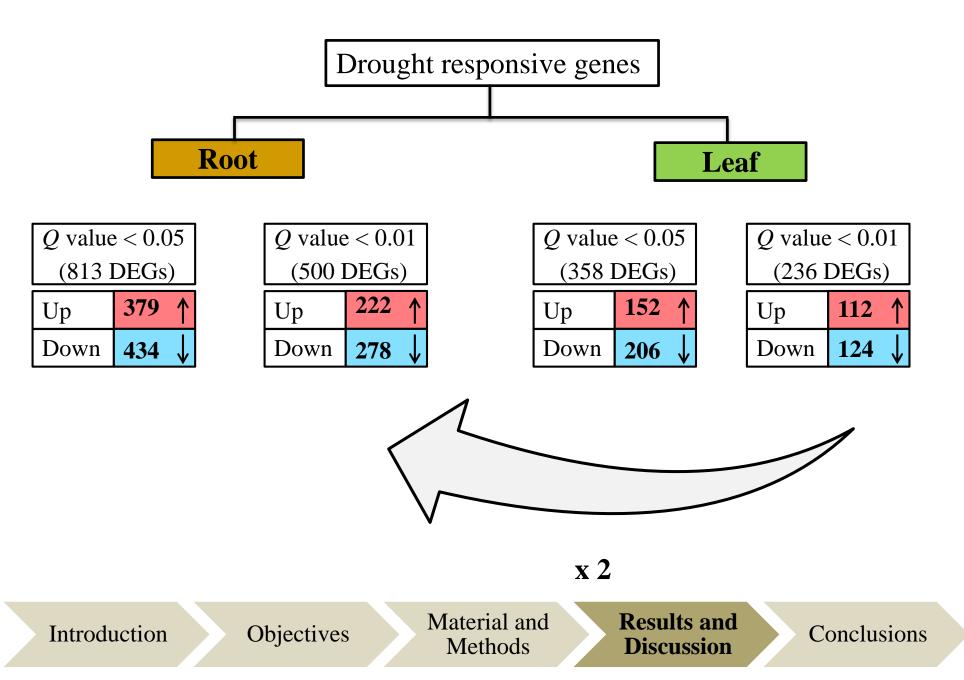
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l	Total	315.08	121.33	67.27	188.60	86.05	49.94	135.99

72.34%

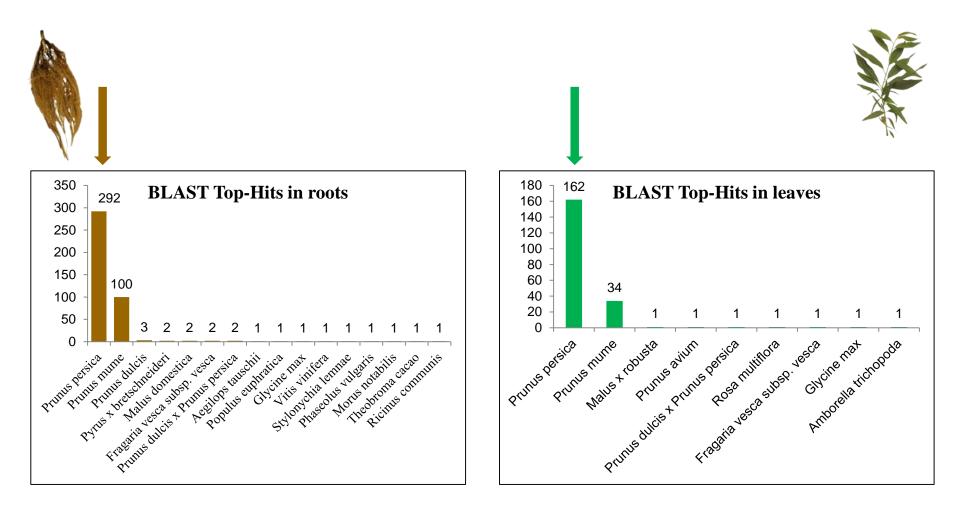
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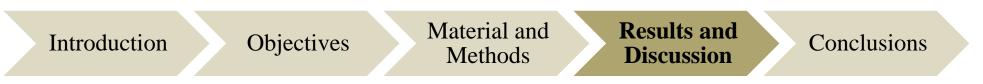
Material and Methods Results and Discussion

4.3 Analysis of differentially expressed genes (DEGs)

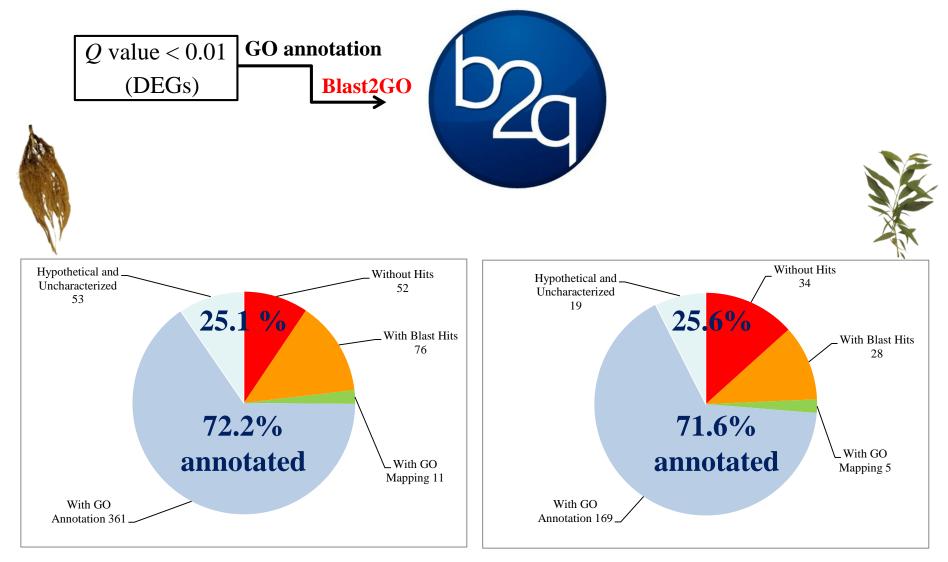


4.4 Sequence homology searches



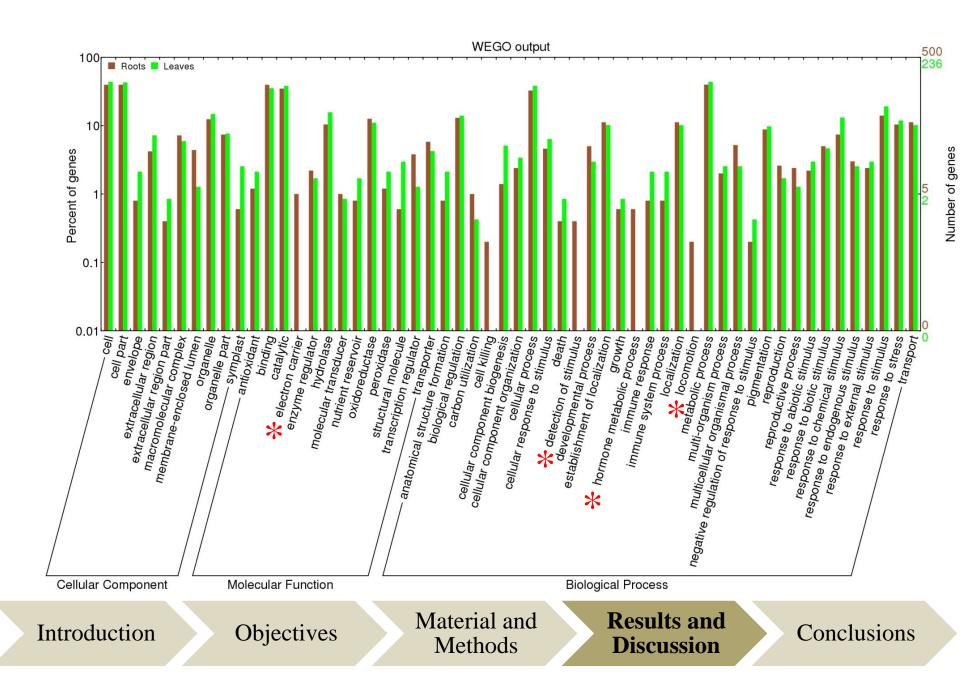


4.5 Functional annotation of DEGs



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4.5 Functional annotation of DEGs (56 GOs)



4.6 Gene ontology enrichment GOs



3 GO terms (data not shown)



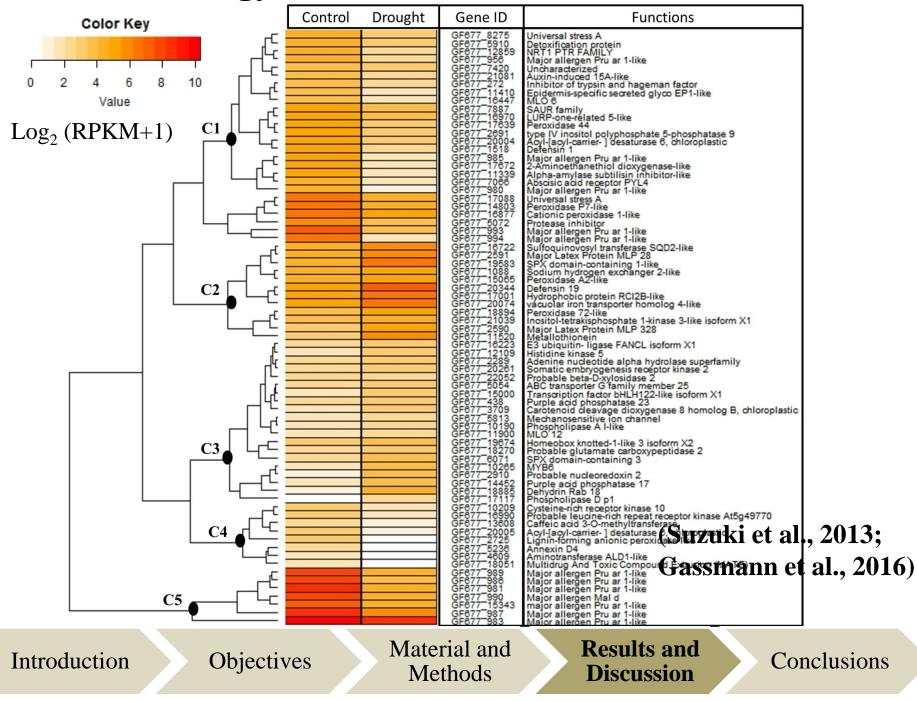
26 GO terms (Biological process and molecular function)

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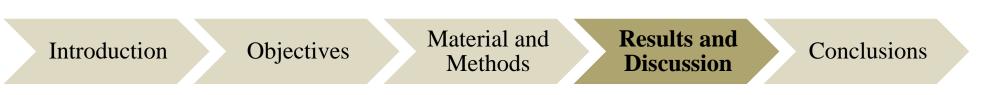
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4.6 Gene ontology enrichment GOs (II)

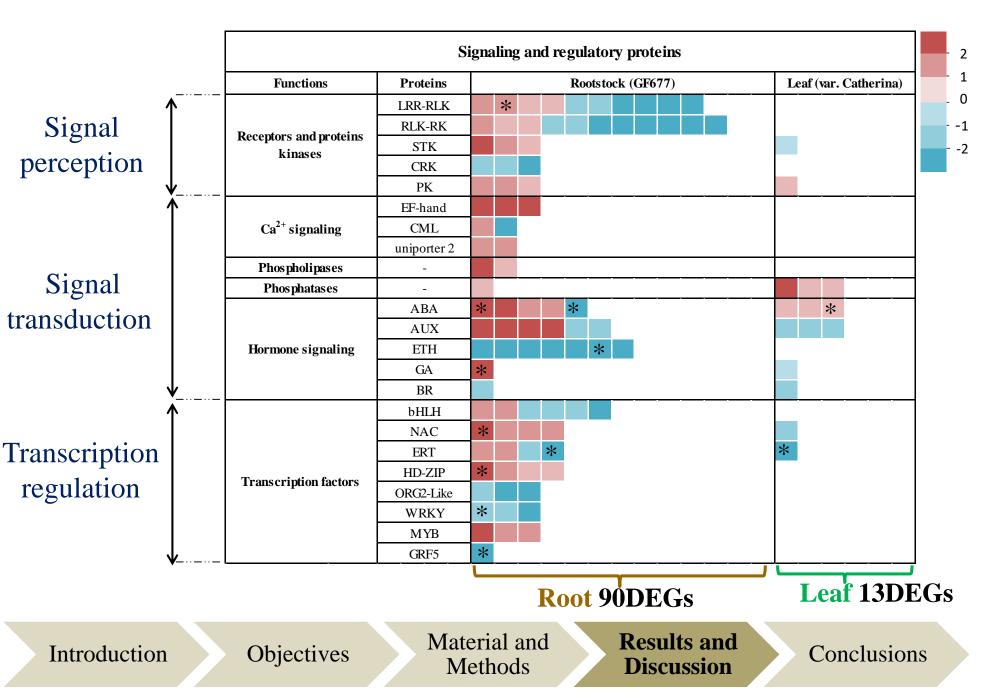


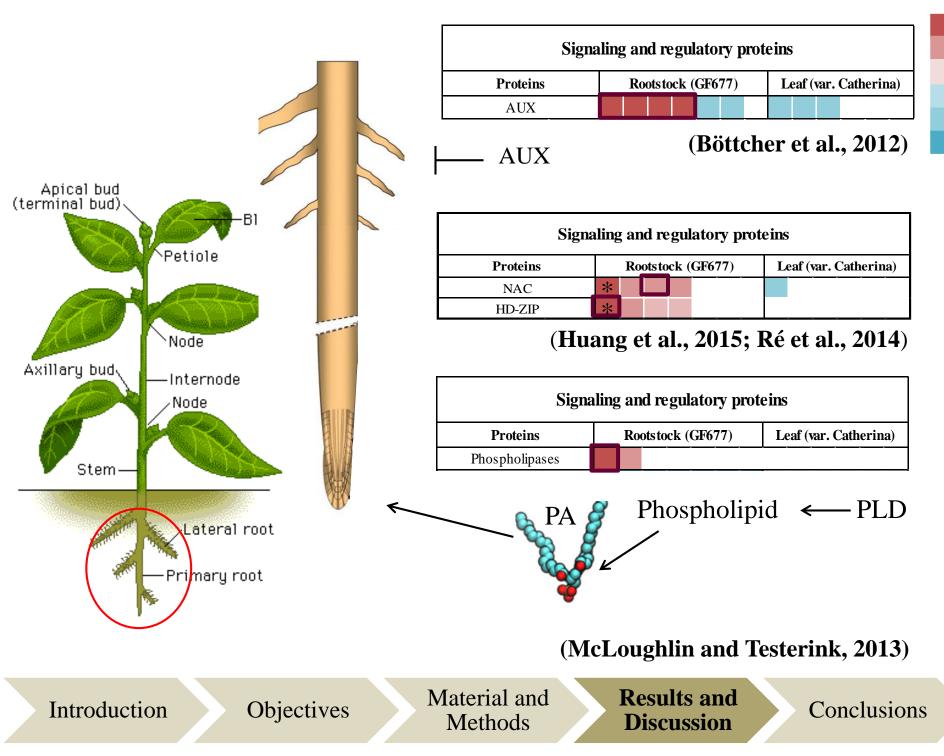
4.7 Functional characterization of DEGs

Regulatory	• Signaling and transcriptional regulation
Functional	• Cell protection and damage repair



4.7.1 Regulatory role of DEGs

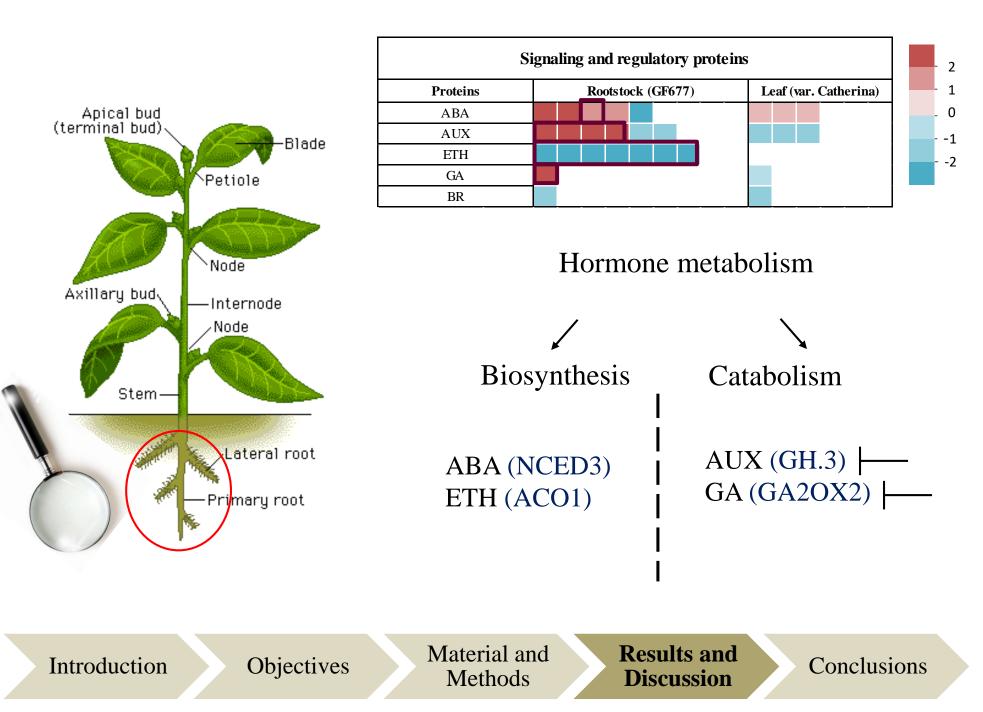




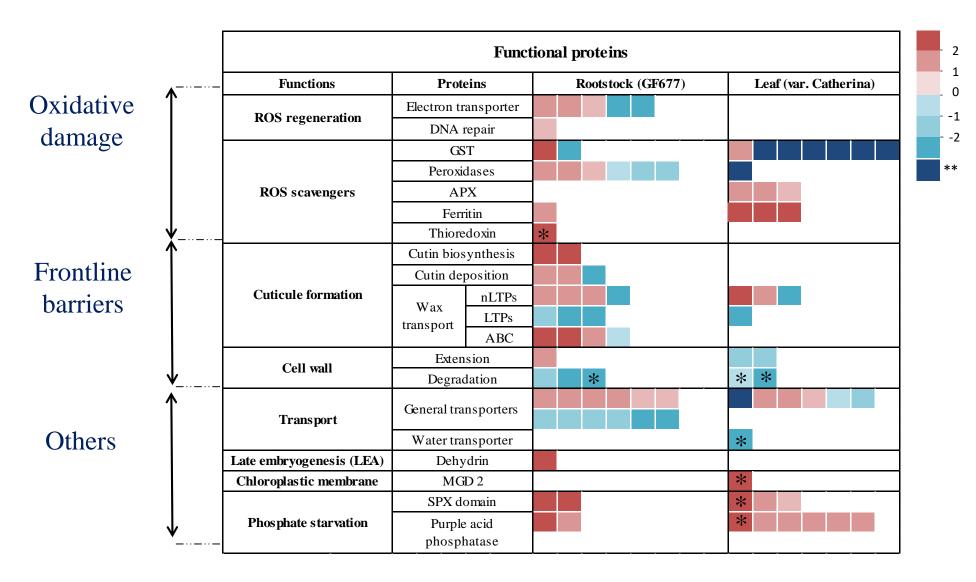
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4.7.2 Funcional role of DEGs



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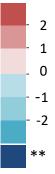
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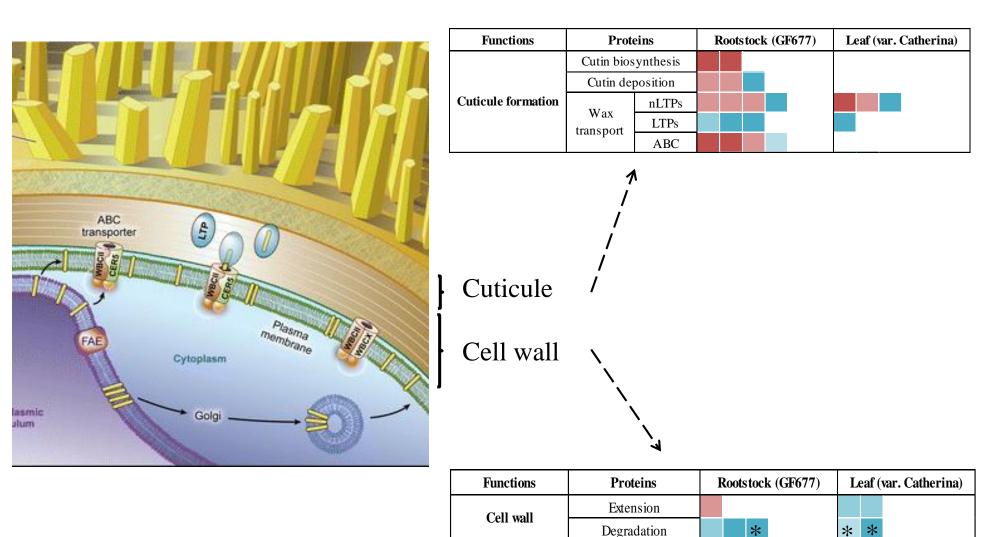
4.7.2 Funcional role of DEGs

Functional proteins				
Functions	Proteins	Rootstock (GF677)	Leaf (var. Catherina)	
ROS regeneration	Electron transporter			
	DNA repair			



Functions	Proteins	Rootstock (GF677)	Leaf (var. Catherina)	
ROS scavengers	GST			
	Peroxidases			
	APX			
	Ferritin			
	Thioredoxin	*		

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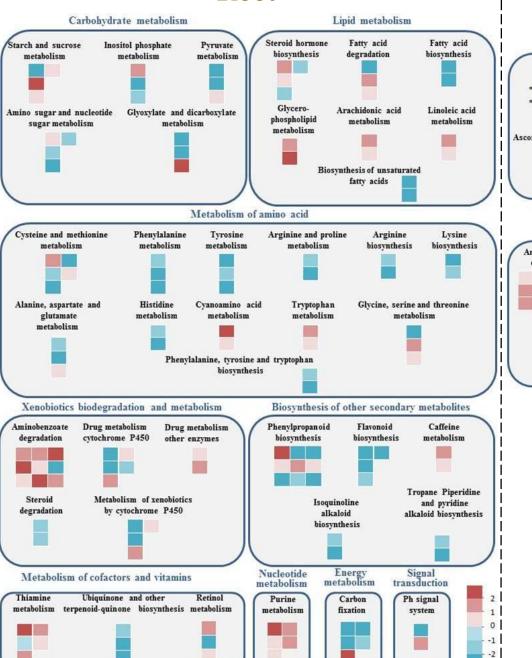
Degradation

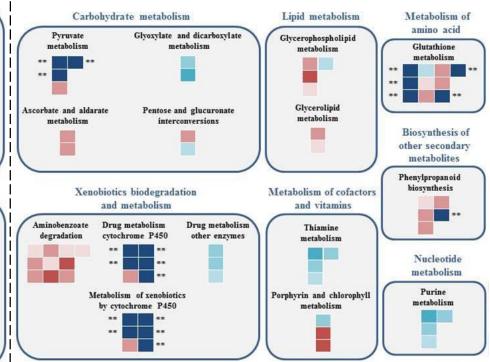
Conclusions

4.8 Metabolic pathways

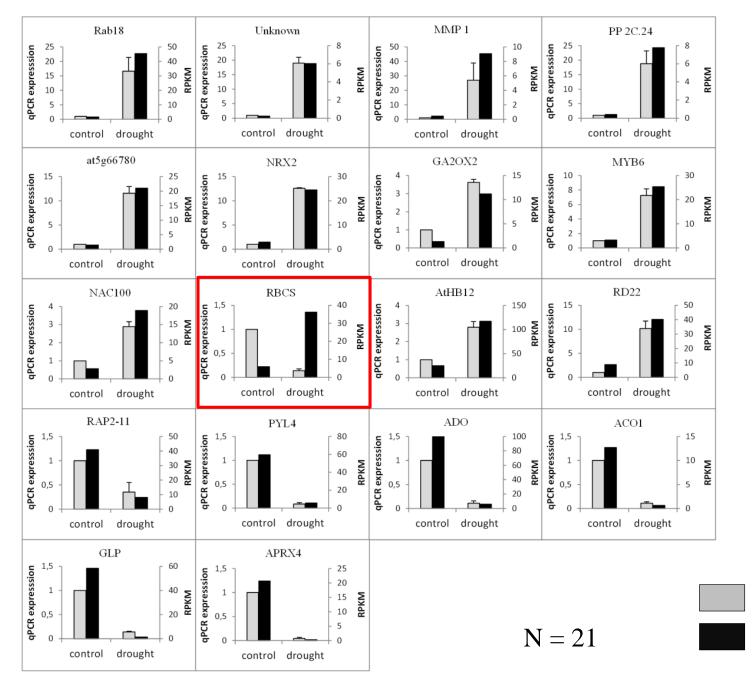
Root







4.9 Validation of RNA-seq reliability using RT-qPCR



RT-qPCR

RPKM

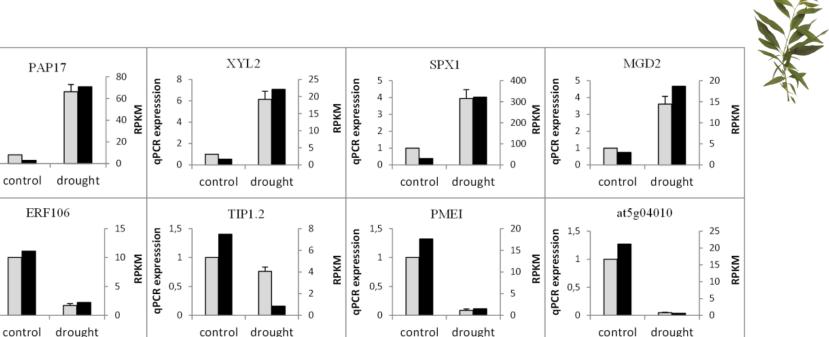
4.9 Validation of RNA-seq reliability using RT-qPCR

1,5

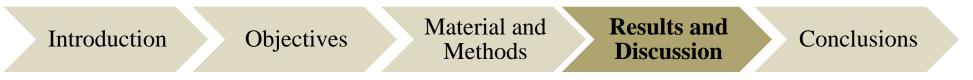
0,5

qPCR expression

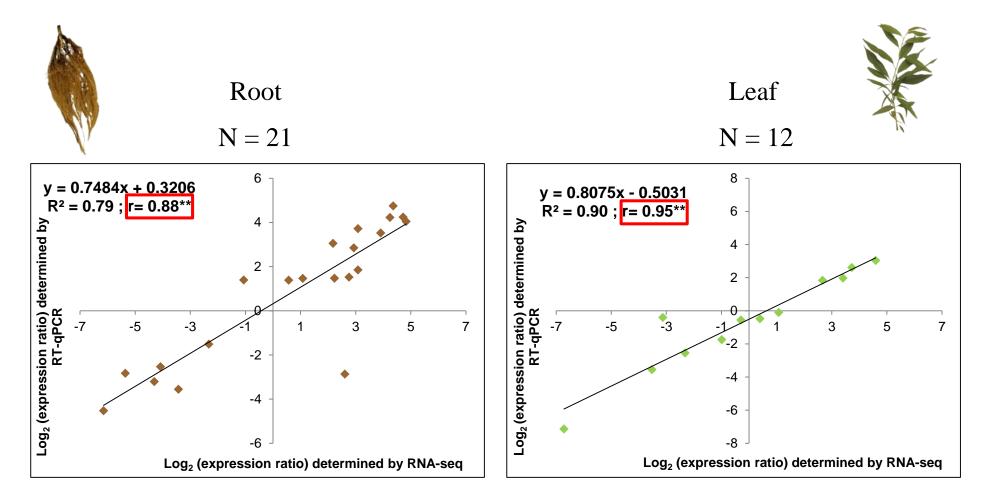
qPCR expression







4.9 Validation of RNA-seq reliability using RT-qPCR (II)



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5- Conclusions

1. RNA-seq is a powerful tool for exploring the diversity of differentially expressed genes expressed in different tissues, however, the analysis of these results is complex and should be validated through other approaches such as RT-qPCR.

2. This study is the first that simultaneously evaluates the effect of drought on roots and on leaves of a grafted variety of peach, observing changes at the physiological and transcriptional level.

3. The differentially expressed genes (DEGs) in roots are approximately twice the number of DEGs found in leaves, suggesting thereby that roots undergo more complex gene regulation, highlighting the role of rootstock as stress buffer during water deprivation.

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5- Conclusions (II)

4. Functional annotation allows dissecting molecular aspects of drought responses, selection of candidate genes, and the formulation of hypotheses that can be validated experimentally.

5. Abscisic acid (ABA) is the only inducible hormone under drought stress conditions, while other hormones, and their associated metabolic pathways, are repressed.

6. GRF5 and AtHB12 are candidate genes potentially involved in drought stress adaptation providing thus a good starting point for investigation in *P. persica*.





Transcriptional Responses in root and leaf of Prunus persica Under Drought Stress Using RNA Sequencing

Ksouri et al., 2016

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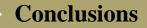
Revised on: 24 Oct 2016

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Thank you for your attention