












Review

From ‘Farm to Fork’: Exploring the Potential of Nutrient-Rich and Stress-Resilient Emergent Crops for Sustainable and Healthy Food in the Mediterranean Region in the Face of Climate Change Challenges

Javier Matías ¹, María José Rodríguez ², Antonio Carrillo-Vico ^{3,4}, Joan Casals ⁵, Sara Fondevilla ⁶, Claudia Mónica Haros ⁷, Justo Pedroche ⁸, Nieves Aparicio ⁹, Nieves Fernández-García ¹⁰, Ingrid Aguiló-Aguayo ¹¹, Cristina Soler-Rivas ^{12,13}, Pedro A. Caballero ¹⁴, Asunción Morte ¹⁵, Daniel Rico ¹⁶ and María Reguera ^{17,*} on behalf of the NutriCrop Spanish Consortium/Group



Citation: Matías, J.; Rodríguez, M.J.; Carrillo-Vico, A.; Casals, J.; Fondevilla, S.; Haros, C.M.; Pedroche, J.; Aparicio, N.; Fernández-García, N.; Aguiló-Aguayo, I.; et al. From ‘Farm to Fork’: Exploring the Potential of Nutrient-Rich and Stress-Resilient Emergent Crops for Sustainable and Healthy Food in the Mediterranean Region in the Face of Climate Change Challenges. *Plants* **2024**, *13*, 1914. <https://doi.org/10.3390/plants13141914>

Academic Editor: Oscar Vicente

Received: 10 March 2024

Revised: 8 April 2024

Accepted: 8 July 2024

Published: 11 July 2024



Copyright: © 2024 by the authors. Licensee MDPI, Basel, Switzerland. This article is an open access article distributed under the terms and conditions of the Creative Commons Attribution (CC BY) license (<https://creativecommons.org/licenses/by/4.0/>).

- ¹ Agrarian Research Institute “La Orden-Valdesequera” of Extremadura (CICYTEX), 06187 Guadajira (Badajoz), Spain; javier.matias@juntaex.es
 - ² Technological Institute of Food and Agriculture of Extremadura (INTAEX-CICYTEX), Avda. Adolfo Suárez s/n, 06007 Badajoz, Spain; mariajose.rodriguez@juntaex.es
 - ³ Instituto de Biomedicina de Sevilla, IBiS/Hospital Universitario Virgen del Rocío/CSIC/Universidad de Sevilla, 41013 Seville, Spain; vico@us.es
 - ⁴ Departamento de Bioquímica Médica y Biología Molecular e Inmunología, Facultad de Medicina, Universidad de Sevilla, 41009 Seville, Spain
 - ⁵ Fundació Miquel Agustí/HorPTA, Department of Agri-Food Engineering and Biotechnology, Universitat Politècnica de Catalunya (UPC)-BarcelonaTech, 08860 Castelldefels, Spain; joan.casals-missio@upc.edu
 - ⁶ Institute for Sustainable Agriculture, Consejo Superior de Investigaciones Científicas, Avda. Menéndez Pidal s/n, 14004 Córdoba, Spain; sfondevilla@ias.csic.es
 - ⁷ Cereal Group, Institute of Agrochemistry and Food Technology (IATA-CSIC), Av. Agustín Escardino 7, Parque Científico, 46980 Valencia, Spain; cmharos@iata.csic.es
 - ⁸ Group of Plant Proteins, Instituto de la Grasa, CSIC. Ctra. de Utrera Km. 1, 41013 Seville, Spain; j.pedroche@csic.es
 - ⁹ Agro-Technological Institute of Castilla y León (ITACyL), Ctra. Burgos Km. 119, 47071 Valladolid, Spain; apagutni@itacyl.es
 - ¹⁰ Department of Abiotic Stress and Plant Pathology, Centro de Edafología y Biología Aplicada del Segura (CSIC), Campus Universitario de Espinardo, 30100 Murcia, Spain; nieves@cebas.csic.es
 - ¹¹ Postharvest Programme, Institute of Agrifood Research and Technology (IRTA), Parc Agrobiotech Lleida, Parc de Gardeny, Edifici Fruitcentre, 25003 Lleida, Spain; ingrid.aguiló@irta.cat
 - ¹² Departamento de Producción y Caracterización de Nuevos Alimentos, Institute of Food Science Research-CIAL (UAM+CSIC), Campus de Cantoblanco, Universidad Autónoma de Madrid, C/Nicolas Cabrera 9, 28049 Madrid, Spain; cristina.soler@uam.es
 - ¹³ Sección Departamental de Ciencias de la Alimentación, Facultad de Ciencias, Universidad Autónoma de Madrid, Campus de Cantoblanco, 28049 Madrid, Spain
 - ¹⁴ Food Technology, Department of Agriculture and Forestry Engineering, Universidad de Valladolid, 34004 Palencia, Spain; pedroantonio.caballero@uva.es
 - ¹⁵ Departamento Biología Vegetal, Facultad de Biología, Campus Universitario de Espinardo, Universidad de Murcia, 30100 Murcia, Spain; amorte@um.es
 - ¹⁶ Department of Medicine, Dermatology and Toxicology, Universidad de Valladolid, Av. Ramón y Cajal, 7, 47005 Valladolid, Spain; daniel.rico@uva.es
 - ¹⁷ Departamento de Biología, Campus de Cantoblanco, Universidad Autónoma de Madrid, C/Darwin 2, 28049 Madrid, Spain
- * Correspondence: maria.reguera@uam.es; Tel.: +34-91497-81-89
 † Collaborators of the NutriCrop Spanish Consortium/Group are provided in the Acknowledgments.

Abstract: In the dynamic landscape of agriculture and food science, incorporating emergent crops appears as a pioneering solution for diversifying agriculture, unlocking possibilities for sustainable cultivation and nutritional bolstering food security, and creating economic prospects amid evolving environmental and market conditions with positive impacts on human health. This review explores the potential of utilizing emergent crops in Mediterranean environments under current climate scenarios, emphasizing the manifold benefits of agricultural and food system diversification and

assessing the impact of environmental factors on their quality and consumer health. Through a deep exploration of the resilience, nutritional value, and health impacts of neglected and underutilized species (NUS) such as quinoa, amaranth, chia, moringa, buckwheat, millet, teff, hemp, or desert truffles, their capacity to thrive in the changing Mediterranean climate is highlighted, offering novel opportunities for agriculture and functional food development. By analysing how promoting agricultural diversification can enhance food system adaptability to evolving environmental conditions, fostering sustainability and resilience, we discuss recent findings that underscore the main benefits and limitations of these crops from agricultural, food science, and health perspectives, all crucial for responsible and sustainable adoption. Thus, by using a sustainable and holistic approach, this revision analyses how the integration of NUS crops into Mediterranean agrifood systems can enhance agriculture resilience and food quality addressing environmental, nutritional, biomedical, economic, and cultural dimensions, thereby mitigating the risks associated with monoculture practices and bolstering local economies and livelihoods under new climate scenarios.

Keywords: NUS crops; Mediterranean agrifood systems; abiotic stressors; agricultural diversification; nutritional quality; healthy diets; climate resilience

1. Introduction

Considering the current trends in population growth, achieving global food security remains a future challenge in which agriculture plays a key role [1]. Currently, agriculture is based on the monoculture of a limited number of species [2]. This practice leads to increased fragility of the agricultural system (e.g., in the face of biotic and abiotic stresses), resulting in a depletion of available food ingredients, as our diet relies on a few plant species [3,4]. Furthermore, there is substantial scientific evidence indicating that more diverse agrifood systems are generally safer from both nutritional and agronomical perspectives, showing greater resilience to environmental changes and offering a broader array of nutritionally effective food products [3]. Indeed, growing a variety of crops enhances the nutritional diversity of the local diet. This is particularly important for certain communities that rely heavily on a few staple crops, helping address nutritional deficiencies and improve overall health [4,5]. Introducing novel crops will also offer farmers supplementary income streams, diminishing dependence on a singular crop and alleviating the financial risks associated with market fluctuations or adverse weather conditions [6]. Particularly, certain crops may be more resilient to drought, heat, or other climate-related challenges, providing a buffer against the impacts of climate change [7]. Also, diversifying crops into a rotation system helps break pest, weed infestation, and disease cycles, improves soil fertility, and reduces the risk of soil degradation [8] supporting sustainable agricultural practices. Likewise, the diversification of our agricultural system can contribute to minimizing food dependence that may be subjected, as we have experienced in recent times, to changes linked to international conflicts [9].

Currently, two-thirds of the plant-based foods we consume primarily come from three cereals, wheat, rice, and maize, providing 60% of dietary energy [10]. While the nutritional value of these crops is undeniable, they are deficient in essential nutrients crucial for ensuring a healthy and balanced diet, such as minerals, proteins, essential amino acids, or vitamins, among others [4,11]. Therefore, the diversification of our food crops (for human and animal consumption), by including crops with high nutritional value, is essential for achieving a balanced nutrient intake and promoting greater sustainability and moderation in the consumption of animal-origin foods [12].

The distinct climatic patterns of the Mediterranean region are characterized by mild, wet winters and warm, dry summers [13]. The unique climatic conditions and diverse ecosystems of these areas face increasing challenges derived from climate change which are linked to the escalating occurrence of droughts and elevated temperatures, which directly impact agriculture [14]. Also, pests and diseases may proliferate under altered climate

conditions, posing additional challenges for farmers [15]. Furthermore, rainfed agriculture, which plays a pivotal role in southern and eastern Mediterranean countries, makes a substantial contribution to food security in the region [16]. Therefore, the Mediterranean agrifood system will require the development of adaptation strategies to cope with the aforementioned climatic changing conditions, giving special consideration to rainfed agriculture, particularly in light of its importance in relation to its extent in terms of surface area. This may involve the cultivation of more resilient crop species, which include neglected and underutilized species (NUS) and improved (bred) crop varieties, changes in farming practices, and the implementation of sustainable agricultural techniques to mitigate the impact of climate change, among others [10,17].

NUS, a novel and evolving category in agriculture, holds profound significance for global food security, particularly in addressing nutritional challenges. These crops represent a diverse array of plant species that exhibit resilience and adaptability. Their emergence is a response to the pressing need for sustainable and nutritious food sources, mostly when considering the challenges faced by conventional crops, such as susceptibility to pests, diseases, and climate change, especially in susceptible regions such as the Mediterranean [13–15]. They are promising alternatives for cultivation in diverse ecosystems and their adaptability can contribute to increased Mediterranean agricultural productivity [18].

Overall, it should be considered that the widespread adoption of agricultural diversification practices in the Mediterranean region can contribute to establishing a more sustainable and effective agriculture and food system at both local and global scales [19]. The cultivation of new crops can boost rural economies by creating employment, supporting local businesses, and contributing to overall economic growth. However, it is important to note that the successful introduction of new crops requires careful consideration of factors such as soil suitability, climate conditions, market demand, and farmers' capacity to adapt to new cultivation practices, among others [17]. Additionally, supportive policies, extension services, and research initiatives play a vital role in facilitating the successful integration of new crops [20].

Taking all these aspects into consideration, this review aims to explore the potential of nutrient-rich and stress-resilient emergent crops as a viable solution to foster sustainable and healthy food production in the face of climatic challenges in the Mediterranean region. By examining the adaptability and nutritional value of selected NUSs (including quinoa (*Chenopodium quinoa* Willd.), amaranth (*Amaranthus* spp.), chia (*Salvia hispanica* L.), moringa (*Moringa oleifera* Lam.), buckwheat (*Fagopyrum esculentum* Moench and *Fagopyrum tataricum* (L.) Gaertn.), millet (including *Panicum miliaceum* L. or *Setaria italica* L., among others), teff (*Eragrostis tef* (Zucc.) Trotter), hemp (*Cannabis sativa* L.), or desert truffles (including *Terfezia clavervyi* Chatin)), we aim to shed light on their role in mitigating the adverse effects of climate change on agricultural systems. This work not only delves into the agronomical features of these crops but also considers their nutritional, food technological, and health characteristics, emphasizing the importance of diversifying crops in the Mediterranean region for resilient and nutritionally rich food sources. Through this exploration, we aspire to contribute to the development of strategies that promote food security and sustainability in the Mediterranean region amidst the dynamic landscape of climate change.

2. Agronomical Aspects Related to NUSs for the Mediterranean Region

2.1. Historical Background

Since the advent of agriculture, the Mediterranean region has been a centre of agricultural innovation and crop diversification [21]. Pedoclimatic conditions, characterized by a summer dry season and light soils, facilitated the introduction of Neolithic crop cultures centred around annual plants [22]. Early farmers inherited agricultural practices from the Near East, incorporating domesticated crops such as wheat and barley (*Hordeum vulgare* L.), along with legumes such as peas (*Pisum sativum* L.), lentils (*Lens culinaris* L.), and chickpeas (*Cicer arietinum* L.) [23]. This underscores the significance of these initial introductions over millennia. The prolonged history of co-evolution between these introduced plants, local

agroclimatic conditions, and human preferences has shaped the Mediterranean area into an important secondary diversity centre [24].

Built on this foundation, throughout history, new plant species were introduced, being crucial for colonizing new croplands and enhancing food security. These can be considered the emerging crops of their respective eras. For instance, in early times, rye (*Secale cereale* L.), initially introduced as a weed, became a significant crop for expanding agricultural societies in northern Europe due to its competitive strength in poor soils and unfavourable climates [25]. In the Middle Ages, Arabic settlements introduced crops like rice, playing a vital role in developing agriculture in deltaic areas, and sorghum (*Sorghum bicolor* L. Moench), cultivated in marginal lands with insufficient rainfall, contributing to ensuring food production [26]. In more recent times, exchanges with the Americas introduced key species, notably maize and potato (*Solanum tuberosum* L.), swiftly integrated into agricultural systems, especially in regions facing challenges in producing bread-making cereals.

Several other ‘emerging crops’ were successful in specific areas, highlighting the significance of plant introductions for local agriculture in the past. Many of these are now considered NUS but hold potential as alternative crops to enhance agricultural production in specific environments [27]. Some other examples are common millet (*Panicum miliaceum* L.), foxtail millet (*Setaria italica* L.), buckwheat, or different *Triticum* species such as einkorn (*T. monococcum* L.), polish wheat (*T. polonicum* L.), club wheat (*T. compactum* L.), or spelt (*T. spelta* L.) [28]. These plants played pivotal roles in food security during periods of agricultural crisis, caused by the depletion of soil fertility or climate crisis. A notable example of the role of emerging crops in adapting to climate changes is demonstrated by societies’ response to the Little Ice Age (LIA) cooling (1550–1710) [29]. This cooling period had adverse effects on agricultural production, leading to heightened famine. Societies that diversified agriculture, through the introduction of new crops and adapting cultivation techniques, exhibited increased resilience [30].

Apart from plant species, fungi have also been important for human nutrition. For example, fungal species have been harvested from the wild or cultivated to complement diets [31]. A notable example is the desert truffle (*Terfezia clavaryi* Chatin), which has recently been integrated into Mediterranean cropping systems. Historically, desert truffles were gathered from the wild by diverse cultures across the Middle East, Africa, and Australia. Despite geographical differences, these cultures used similar methods for truffle collection, preparation, and use [32]. The cultivation of desert truffles began relatively recently, with the first plantation established in Murcia, Spain, in 1999. This effort led to the production of truffles two years later, marking the start of global desert truffle cultivation, which has since expanded to various regions in Spain and other countries [33,34].

In summary, from the origin of Mediterranean agricultural societies to the present, emerging crops have been fundamental to agricultural development and food security. Adapting agriculture to the current climate crisis should draw from past successful experiences, leveraging the agronomic potential of underutilized crops and the diversity of landraces that have co-evolved with Mediterranean agrosystems.

2.2. Importance and Benefits

Biodiversity holds paramount importance for ensuring food security and nutrition in Mediterranean agricultural ecosystems. The rich diversity within agricultural systems serves as a foundational asset for continually enhancing varieties essential for sustaining agriculture in the context of climate change. In line with this, the crucial role of NUS in sustainable agriculture, ensuring improved environmental protection and a consistent food supply, entails selecting cost-effective varieties adapted to current cultivation areas and of economic and agronomic interest [1]. Despite NUS encompassing crops that lack broad adoption, they offer added value compared to conventional crops, being nutrient rich, adaptable to poor soils and arid areas, requiring minimal water for proper development, and utilizing nearly 100% of the plant (root, leaf, seed, etc.) or truffle.

In the face of climate change and evolving markets, knowledge and innovation become pivotal for sustainable success in agriculture [35]. Enhancing innovation performance becomes crucial for institutions and companies in the agricultural sector to remain competitive. Beyond technology adoption, innovation must anticipate market needs and identify superior products and processes, as exemplified by the introduction of new sustainable crops in Mediterranean areas such as Portugal, Spain, Italy, Greece, Turkey, Egypt, and Morocco [36].

Recognizing the significant negative impact of climate change on food production, the international scientific community acknowledges increased risks such as temperature fluctuations, droughts, and floods. The agricultural sector, a net emitter of greenhouse gases, faces vulnerability to climate change effects. Adapting, reducing vulnerabilities, and increasing resilience to climate change is thus crucial. The introduction of NUS crops emerges as a strategy to adapt and ensure food security and good nutrition in the face of climate change, aligning with the 17 Sustainable Development Goals committed by the global community [37]. However, climate change poses challenges to achieving these goals, hindering livelihood development and food security, especially for vulnerable populations. The incorporation of NUS becomes instrumental in overcoming these challenges and fostering food security and nutrition for humanity in the future [37].

2.3. Classification and Characteristics

NUSs are also referred to as novel or emergent crops, break crops, or alternative crops, among other terms in use. These crops can be classified based on various criteria such as use, climate adaptability, nutritional content, growth requirements, and other specific traits. According to their intended use, emerging crops can be destined for food, e.g., quinoa, or for industrial use, e.g., hemp. The classification can also focus on traits associated with climate adaptability, which make crops well suited for specific conditions (Table 1). Consequently, different categories can be established in this line, such as drought-tolerant crops (e.g., quinoa [38], millet [39], hemp [40], and desert truffles [41]), heat-resistant crops (e.g., millet [42], amaranth [43]), or salinity-tolerant crops (e.g., quinoa [44], sorghum [45]). Furthermore, crop growth requirements can be categorized into three levels: low, medium, and high. In the case of NUS, there is significant importance placed on cultivating crops with relatively low growth requirements. This emphasis is geared towards reducing the environmental impact of agriculture on climate change and enhancing the resilience of agricultural systems. Moreover, emerging crops can also be classified based on their nutritional value (See Section 3), establishing different categories such as high-quality protein crops (e.g., quinoa, amaranth, moringa, desert truffles), nutrient-dense crops (e.g., moringa, quinoa, amaranth), or omega-3-rich crops (e.g., chia, flaxseeds (*Linum usitatissimum* L.)), among others.

Quinoa, amaranth, millet, buckwheat, hemp, teff, moringa, desert truffles, and chia, among others (Figure 1), are considered underutilized climate-resilient crops [46–53] (Table 1). The introduction of these eco-friendly crops into our agriculture has the potential to enhance biodiversity and resilience in agricultural systems. Currently, agricultural production heavily relies on only a few species, and diversifying with these crops can contribute to a more sustainable and robust agricultural ecosystem [51]. These emerging crops showcase agronomic traits well suited to diverse environmental conditions, characteristic of each species. For instance, quinoa shows great potential due to its ability to withstand abiotic stresses growing in harsh environments with low fertility [54]. Its adaptability to diverse environmental conditions is attributed to its extensive genetic diversity [55], exhibiting tolerance to drought, frost, and salinity, while thriving on poor soils [54]. Amaranth, millet, hemp, chia, and desert truffle also demonstrated resilience to drought [39,40,56–58], making them low-water-requirement options for cultivation. Teff is a warm-season cereal crop adapted to a wide range of environments that can be grown under rainfed conditions [59]. Moringa exhibits tolerance to drought conditions. In regions

with a subtropical climate, moringa can resist up to six months of dry seasons, provided the annual rainfall amounts to 500 mm [60].

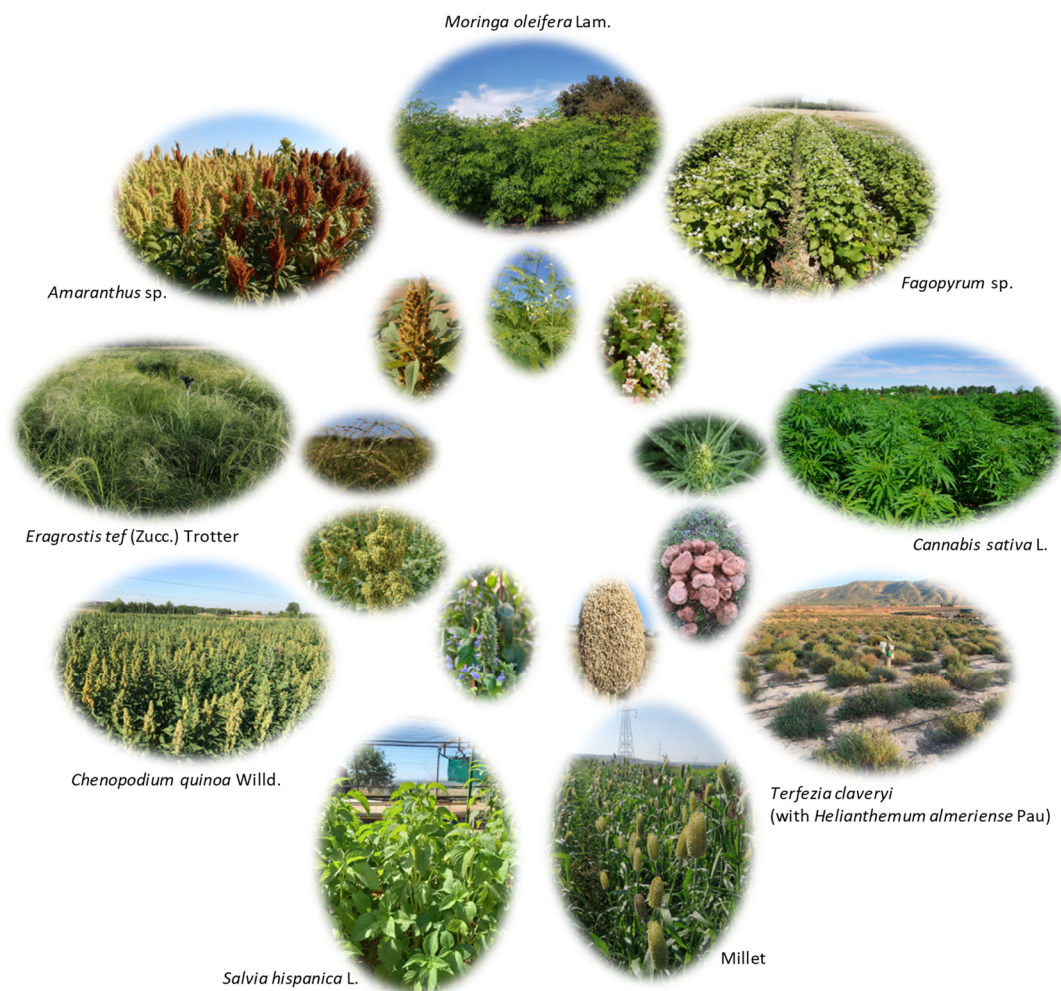


Figure 1. NUS crops with the potential for cultivation in Mediterranean environments. Representative images of different nutrient-rich and stress-resilient emergent crops for sustainable and healthy food in the Mediterranean region, grown under field conditions across different regions of Spain. The images depict the crops in the field (outer images) and the panicle, inflorescence, or truffle for each crop (inner image). The crops included in the figure are *Chenopodium quinoa* Willd., *Amaranthus* sp., *Fagopyrum* sp., *Eragrostis tef* (Zucc.) Trotter, *Salvia hispanica* L., *Moringa oleifera* Lam., *Cannabis sativa* L., and millet (*Pennisetum glaucum* (L.) R.Br. *Terfezia clavaryi* Chatin (with *Helianthemum almeriense* Pau)).

Tolerance to heat has been reported for amaranth, a C4 plant, and millet [42,57], while millet, amaranth, hemp, and quinoa can withstand relatively saline soils [45,61–63]. Buckwheat thrives in cold climates and is well suited for poor soils, with reduced water requirements [64]. By contrast, certain desert truffle species from *Terfezia* sp. and *Tirmania* sp. face population decline due to reduced rainfall and changes in land use, prompting research on domestication for a drought-resistant crop in line with sustainable practices [33]. Overall, these minor crops can thrive in marginal soils.

The primary challenge hindering their expansion and introduction in regions like the Mediterranean basin is the scarcity of commercially available varieties well adapted to the specific environmental conditions of these areas. For example, photoperiod is the primary obstacle to quinoa's adaptability in the northern hemisphere [65]. Quinoa is a facultative short-day and day-neutral plant [66]. Photoperiod is also a major limitation for chia cultivation in Europe because it requires a floral induction photoperiod of around 12 h

of darkness [67]. Furthermore, quinoa and buckwheat growing under high temperatures during flowering can experience dramatic yield losses [50,68]. On the contrary, it has been shown that one of the primary constraints on the growth and productivity of moringa is low temperatures. Moringa thrives in tropical and subtropical climates. Moringa can withstand light frosts down to $-3\text{ }^{\circ}\text{C}$. Additionally, it is a demanding crop in terms of light [60]. In the case of teff, lodging represents the most significant issue limiting yields [69], and drought adaptation in desert truffle mycorrhizal plants is key when aiming at expanding its cultivation and involves enhancing plant nutrition, photosynthesis, water use efficiency, and root colonization morphology [41]. Additionally, “turmiculture,” an agricultural approach with minimal water input and sustainable practices, enables the cultivation of this natural resource in arid and semi-arid areas [34].

Therefore, in the context of climate change, traits that make emerging crops well suited for Mediterranean conditions include tolerance to the harsh environmental conditions expected to intensify in the coming years such as drought, heat, and salinity [70].

Table 1. A list of NUS crops with the potential of cultivation in Mediterranean environments.

Common Name	Scientific Name	Botanical Family	Type	Abiotic Stress Tolerance			
				Cold	Heat	Drought	Salinity
Quinoa	<i>Chenopodium quinoa</i> Willd.	Amaranthaceae	Dicotyledons/Herbaceous/C3	Medium	Medium/low	High	Medium/high
Amaranth	<i>Amaranthus</i> sp. (75 species [71])	Amaranthaceae	Dicotyledons/Herbaceous/C4	Medium/low	High	High	Medium
Buckwheat	<i>Fagopyrum</i> sp. Subfamilies:	Polygonaceae	Dicotyledons/Herbaceous/C3	Medium	Medium/low	High	Medium
Millet/Sorghum	- Chloridoideae	Poaceae	Monocotyledons/Herbaceous/C4	Low	High	High	High
	- Panicoideae (11 species excluding teff) [72]						
Teff	<i>Eragrostis tef</i> (Zucc.) Trotter	Poaceae	Monocotyledons/Herbaceous/C4	Medium	High	Medium	Medium/low
Chia	<i>Salvia hispanica</i> L.	Lamiaceae	Dicotyledons/Herbaceous/C3	Low	Medium	Medium	Medium
Moringa	<i>Moringa oleifera</i> Lam.	Moringaceae	Dicotyledons/Woody/C3	Very low	High	High	Medium/high
Hemp	<i>Cannabis sativa</i> L.	Cannabaceae	Dicotyledons/Herbaceous/C3	Low	Medium/high	High	High
Desert truffles	<i>Terfezia clavayii</i> Chatin (with <i>Helianthemum almeriense</i> Pau)	Pezizaceae (Cistaceae)	Ascomycete (Dicotyledons/woody/C3)	Medium/high	Medium	Medium	Medium/low

2.4. Challenges and Limitations

The cultivation of NUSs poses various challenges across different facets of the agricultural and supply chain landscape. Ongoing threats to agricultural environments stem from a growing world population, limited arable land, and swift climate changes. Projections suggest that the global population will surpass 9 billion by 2050 [73]. The challenge of feeding this expanding population is a global concern. To secure food and ecosystem stability, it is essential to develop NUSs as crops for sustainable agriculture, emphasizing increased net production while minimizing adverse environmental impacts. Nevertheless, incorporating NUSs into existing rotations introduces complexities, necessitating careful consideration of interactions with other crops to prevent adverse impacts on soil health and pest dynamics. The utilization of diversified crop rotations is seen as pivotal in addressing sustainability challenges in modern agriculture. However, designing rotations that strike compromises across multiple sustainability domains remains a persistent challenge [74]. Pests and diseases pose another set of challenges, as resistant varieties for NUSs may be limited, rendering them susceptible to various threats. In this context, monitoring and managing pests and diseases become challenging due to the relative novelty of NUS in certain regions, where existing control measures may not be well established. Advancements in documenting pathogens and pests affecting NUSs, along with the development of corresponding management strategies, are, therefore, essential [75].

NUSs face an additional challenge due to the growing interest of consumers worldwide in aligning their food consumption with sustainability [76] and the increasing demand for healthier food choices [77]. These are the factors driving the increasing interest in nutrient-rich and stress-resilient NUS crops. Furthermore, processing emerging crops faces

infrastructure and technological challenges. It is crucial to adapt existing facilities to the processing requirements of NUSs and concurrently establish quality control measures to meet market standards. Despite the great interest in NUSs, most are not yet well adapted and highly marketable. Developing efficient supply chains, encompassing logistics, distribution, transportation, storage, and distribution networks, is crucial [78]. Limited awareness among consumers and retailers necessitates marketing and educational initiatives. The specific niche markets associated with NUSs create challenges in establishing demand, requiring targeted marketing strategies. Socio-economic factors further complicate the adoption of NUSs. Convincing farmers to embrace change and cultivate NUS crops, especially when traditional crops have established markets and proven agronomic practices, is a hurdle. The economic feasibility of novel crops' cultivation compared to traditional crops plays a pivotal role, requiring assurances of long-term financial viability. Additionally, the lack of supportive policies, such as research incentives or financial aid, can impede widespread adoption.

Thus, addressing all the above-mentioned challenges is essential for the successful integration of NUS crops into agricultural practices, ensuring sustainable cultivation, economic viability, and widespread adoption. Collaboration among farmers, researchers, policymakers, and industry stakeholders is crucial to developing sustainable cultivation practices, improving processing technologies, and creating effective market strategies. Additionally, investments in research and development, extension services, and capacity-building programs can contribute to the successful integration of emerging crops into agricultural systems and markets.

2.5. Genetic and Breeding Advances

Genetic improvement in NUS crops has been considerably lower compared to major crops. Consequently, there exists significant potential for enhancement in various agronomical and nutritional traits. Additionally, many of these crops originate from regions with agroclimatic conditions vastly different from those in the Mediterranean. Hence, ongoing breeding efforts are essential to develop varieties better adapted to Mediterranean agroclimatic conditions. As previously mentioned, photoperiod sensitivity poses a challenge for cultivating some NUS crops in the Mediterranean basin. Numerous accessions of quinoa, amaranth, chia, buckwheat, and hemp require short days for optimal production. For instance, quinoa exhibits two germplasm pools: the primary Andean highland quinoa and the secondary central and southern Chilean quinoa [79]. Fortunately, certain quinoa accessions showing day-length insensitivity have been identified in the coastal conditions of southern Chile. These have been utilized to develop varieties capable of thriving in European long-day environments. European varieties, developed in Denmark and the Netherlands, are currently available, and a breeding program is underway in Spain to create new varieties specifically adapted to Mediterranean field conditions [80]. Similarly, while many buckwheat varieties are sensitive to photoperiod, Europe now cultivates varieties that are not influenced by day length [81]. Regarding chia, there are a few patented varieties, such as the 'Ouro' variety from France, exhibiting long-day flowering characteristics, obtained through selective breeding [82]. Hemp, being a short-day plant with flowering highly dependent on photoperiod and temperature, also includes varieties like 'Finola' that can initiate flowering independently of photoperiod [83].

One of the key constraints for crop cultivation in the Mediterranean basin is the occurrence of episodes of high temperatures during the crop growing cycle. Numerous studies have demonstrated that elevated temperatures at flowering time can significantly diminish yield in quinoa and buckwheat [50,68]. Consequently, enhancing heat tolerance has become a crucial breeding target to increase the yield of quinoa and buckwheat in the Mediterranean basin.

The success of developing new crop varieties that thrive in different environments and exhibit enhanced agronomic and nutritional traits depends on genetic variability, understanding the genetics of target traits, and having effective improvement methods and

tools. Quinoa germplasm exhibits high diversity, allowing its adaptation to various environments. However, accessing South American germplasm presents limitations, requiring negotiations with national governments and often necessitating facilitation by international organizations like FAO [84]. Despite being domesticated 7000 years ago by ancient Andean civilizations, quinoa breeding programs only commenced in the 1960s in the Andes and later in other regions from the 1970s onwards [85]. Several quinoa breeding programs, including hybridization, are ongoing in different countries. However, scientific knowledge about critical aspects of quinoa breeding, such as the inheritance of desirable agronomic and quality traits or the identification of molecular markers linked to them for marker-assisted selection, remains limited. The sequencing of the quinoa genome by Jarvis et al. (2017) [86] has provided a powerful tool to expedite quinoa breeding. For instance, this genome sequence facilitated the identification of a gene controlling saponin content [86]. Moreover, the availability of the quinoa genome sequence has enabled the identification, through genome-wide association studies (GWAS), of candidate genes for photoperiodic flowering regulation, thousand seed weight, and other agronomic traits [87,88]. Additionally, extreme gradient boosting has been employed to identify candidate genes potentially controlling betalain content in quinoa seeds [89]. These studies open the possibility of utilizing marker-assisted selection in quinoa breeding.

In contrast, a notable scarcity of genetic diversity has been observed within domesticated chia accessions, with slightly greater variability present in wild accessions [82]. Furthermore, gene banks hold limited chia germplasm [90]. Additional constraints for chia breeding include the presence of small and fragile corollas, fused flower parts, and a propensity for self-pollination, factors that have impeded the establishment of breeding programs centred on hybrids [91]. Chia has not been the focus of extensive modern plant breeding efforts, and improved cultivars have primarily arisen through the selection of lines from mixed germplasm sources, typically landraces [91]. Thus, limited knowledge exists regarding the genetics of pertinent traits in chia, although some progress has been achieved. Studies on the inheritance of photoperiod response, seed colour, lodging resistance, and shattering resistance have been conducted [91]. Also, transcriptomic studies have identified genes involved in secondary metabolites and oil biosynthesis [92]. Furthermore, chia genome sequencing has facilitated the characterization of genes associated with polyunsaturated fatty acids (PUFAs), mucilage biosynthesis, and those coding for oleosin, caleosin, and steroleosin [93].

Amaranth is considered an orphan crop due to the limited efforts directed towards its genetic improvement [94]. Nevertheless, some progress has been achieved through hybridization (including hybrids between different species), selection, and mutagenesis methods [79]. While most varieties have been developed using conventional breeding, gamma irradiation-induced mutation has also been employed in amaranth for the development of new cultivars [95]. Nonetheless, genetic studies in amaranth are scarce, with markers identified for plant growth, morphology, seed characteristics, and flowering behaviour. Inheritance studies have been conducted for nutritional factors, including starch characteristics, the perisperm layer of grain, seed protein content, and male sterility [79]. A Quantitative Trait Loci (QTL) for flower colour was identified in *A. hypochondriacus* L. [96]. Additionally, a recent GWAS in *Amaranthus tricolor* L. identified marker-trait associations associated with branching index, inflorescence colour, petiole pigmentation, and terminal inflorescence shape and attitude [97]. Noteworthy, genomic resources for amaranths include the genomes of several *Amaranth* spp. (the genomes of *A. hypochondriacus* L., *A. tricolor* L., *A. palmeri* S. Wats., and *A. cruentus* L. are available in the NCBI dataset), genome-wide SNPs and SSRs for *A. caudatus* L. and *A. hypochondriacus* L. and ‘popAmaranth,’ a population genetic genome browser for grain *Amaranthus* and their wild relatives [95].

Among all buckwheat species, only two are cultivated for grain food production: common buckwheat (*Fagopyrum esculentum* Moench) and tartary buckwheat (*F. tataricum* (L.) Gaertn). Common buckwheat has a wide distribution in Asia, Europe, America, and Austria, while tartary buckwheat is primarily grown in Asia [98]. Various breeding programs

are dedicated to improving buckwheat varieties in Europe, where the cultivated types are short, early maturing, and not sensitive to photoperiod [81]. A crucial aspect of buckwheat breeding is that common buckwheat is highly allogamous due to the presence of unique dimorphic and sporophytic types of self-incompatibility, making it a heteromorphic strictly self-incompatible crop. The heteromorphic self-incompatibility in common buckwheat is controlled by a single gene locus [99]. A draft assembly of the common buckwheat genome, generated by Zhang et al. [100], successfully identified novel candidate genes controlling this trait. Additionally, a high-quality, chromosome-scale tartary buckwheat genome was released [101]. Progress has also been made in understanding the genetics of various traits in buckwheat. This includes studies on the genetic background of buckwheat flavonoid synthesis [102], QTLs for easy grain dehulling [103], and the identification of a gene involved in bitterness [104] which have contributed to advancing buckwheat breeding. Other authors have identified, by GWAS, candidate genes correlated with flavonoid accumulation and grain weight [100], as well as various morphological and yield-related traits [105] and rutin hydrolysis [106].

Millets, a group of small-grained, self-pollinated cereal grasses, include several species such as pearl millet (*Pennisetum glaucum* (L.) R.Br.), finger millet (*Eleusine coracana* Gaertn.), proso millet (*Panicum miliaceum* L.), foxtail millet (*Setaria italica* (L.) P. Beauv.), little millet (*Panicum sumatrense* Roth ex Roem. & Schult.), and kodo millet (*Paspalum scrobiculatum* L.) or sorghum (*Sorghum bicolor* (L.) Moench) [107]. While millets naturally exhibit high genetic diversity, several hybridization methods have been developed to further enhance diversity [108]. The current discussion will focus on two millet species grown in Europe: foxtail millet and proso millet.

Foxtail millet, recognized for its high diversity, is emerging as a model organism for grasses due to its relatively small genome [107]. Molecular analyses suggest that China is the centre of foxtail millet diversity, with local landrace groups differentiating after domestication. China, Europe, and Afghanistan–Lebanon are proposed centres of domestication for this crop [107]. Molecular tools available for foxtail millet breeding include markers such as SNPs, ILP, and SSRs, used for genetic diversity studies and the mapping of various traits, including domestication and latitudinal adaptation traits, panicle morphology, inflorescence structure, branching and height, and agronomic traits [109]. Molecular facilities encompass databases for molecular markers, transcription factors, miRNA, and transposable element-based markers. Reference genomes and a pan-genome are also available [110,111]. GWASs have been conducted for agronomic and quality traits, as well as water absorption in seeds. In addition, transcriptomic studies have explored traits such as salt tolerance, drought, branching, freezing, and response to rust. Additionally, the expression of some NBLRR genes in response to *Magnaporthe oryzae* (T.T. Hebert) M.E. Barr has been analysed [112].

Despite global breeding efforts and limited genetic characterization, proso millet remains an under-researched and underutilized crop, with only partial success achieved so far. Breeding methods for proso millet include pure line selection, pedigree selection, and backcrossing. The self-pollinating nature of proso millet has posed challenges in hybridization, resulting in only a few registered and released cultivars [107]. Genomic tools for proso millet are scarce, with RAPD, AFLP, SSR, and SNP markers, along with GBS and SPET sequencing, used for diversity studies revealing high diversity [113–115]. Also, a pan-genome is available [98], and a GWAS analysis for agronomic and seed traits has been conducted [116]. Gene expression studies include the identification of selenate and selenite transporters and their expression under salt stress and selenium, as well as a transcriptomic study investigating pathways associated with starch synthesis and metabolism to understand the effect and molecular mechanisms of nitrogen fertilization in starch synthesis in waxy and non-waxy proso millet varieties [117,118].

Teff has been cultivated in the Horn of Africa for millennia. Despite its high germplasm diversity, including wild relatives, landraces, and farmer varieties, this variability has not been efficiently exploited in breeding programs. However, Ethiopia has derived several

varieties from farmer cultivars through mass selection or hybridization programs, resulting in increased yields. Chemical mutagenesis and CRISPR/Cas9 have been attempted to obtain varieties showing resistance to lodging, drought, increased seed size, and tolerance to herbicides, soil acidity, and salt [119,120]. Advances in marker-assisted selection have also been made with the sequencing of the teff genome [121]. QTLs have been identified for morphology and yield traits [122,123], and a GWAS approach used to study agronomic performance identified candidate genes for climatic adaptation and farmer appreciation [120]. Metabolomic studies for salt and drought tolerance, along with a transcriptomic analysis for calcium stress, have also been reported [124–126].

Hemp breeding has been intensive in Europe, resulting in a multitude of available varieties. Initially, breeders employed mass selection to enhance local landraces, followed by the introduction of foreign genetic material through crossing to augment fibre and seed yield [83]. In its natural state, *Cannabis sativa* L. is an annual dioecious plant, with a minor proportion of monoecious plants [127]. Dioecy is governed by two specific genes at linked loci, and the identification of female plants in early development is facilitated by the use of Y-specific DNA markers [128]. A pivotal moment in hemp breeding occurred with the discovery that fibre content is inherited from both the maternal and paternal sides in dioecious hemp. Another significant breakthrough was the identification of hermaphroditic monoecious plants capable of self-pollination, enabling the development of more uniform new varieties. Subsequently, the methods of self-pollination and crossbreeding of monoecious and dioecious plants became more widespread, contributing to the evolution of modern hemp varieties [83]. The identification of plants exhibiting photoperiod insensitivity marked another milestone. The molecular studies in hemp have primarily focused on the synthesis of secondary metabolites and disease resistance. Chemotype inheritance has been investigated [129], and genes involved in cannabinoid synthase have been mapped, with the identification of a gene responsible for cannabichromenic acid (CBCA) biosynthesis [130]. SNPs associated with genes in the cannabinoid pathway have been reported [131], along with QTLs linked to cannabinoid quantity and their ratios [129]. Genes related to resistance against powdery mildew have also been identified [132]. Also, a reference genome is available [133], which has facilitated the identification of genes involved in fatty acid and vitamin E biosynthesis.

Moringa is a cross-pollinated crop indigenous to India that exhibits both geitonogamy and xenogamy. The cultivation of this crop has expanded globally. Moringa varieties are classified into perennial and annual types. Perennial varieties are typically propagated from cuttings, while annuals are propagated from seeds. Both annual and perennial varieties are extensively utilized in breeding programs with a focus on various desirable traits, including dwarf stature suitable for leaf production, high-yielding types, elevated seed and oil content, and resistance to pests and diseases. Diversity genetic studies in Moringa have employed various molecular markers such as RAPD, SSR, CytP450, ISSR, SRAP, and SCoT [134]. Multiple reference genomes are available, with the most recent reported by [135]. These genomes have been instrumental in identifying genes responsible for high protein content, fast growth, and heat tolerance, along with heat shock transcription factors playing a significant role in defence against drought stress [136]. A transcriptome developed by [137] has further contributed to identifying genes responsible for the biosynthesis of secondary metabolites (such as quercetin, kaempferol, and benzylamine), vitamins, and iron transporters. Proteins involved in heat stress have also been reported [138].

3. Nutritional and Technological Aspects Related to NUSs for the Mediterranean Region

3.1. Nutritional Composition

Currently, diets largely depend on starch-rich staple crops such as maize, wheat, and rice [139]. Although these foods cover a large part of the daily caloric needs, they do not provide adequate amounts of other necessary nutrients and micronutrients [140]. Most agriculture-based solutions have not focused on improving nutritional quality but on food production [141]. This has resulted in neglecting the nutritional quality of crops and

biodiversity, which negatively impacts human health. Therefore, there is a need to explore new or emerging crops with high nutritional quality (Table 2).

NUSs are often part of ancestral cultural traditions and have emerging value due to their unrecognized traits [142]. Recently, many minor crops have been marketed as health foods or “superfoods”, as was the case with quinoa (*Chenopodium quinoa*). The growing market interest in foods of high nutritional quality could represent the transition from a marginal crop to one of greater interest [143].

The growing interest in pseudocereals (such as quinoa, amaranth, and buckwheat) arises from their heightened protein content, improved amino acid composition, and balance when compared to traditional cereals, coupled with the presence of biologically active molecules [144]. Quinoa and amaranth species are esteemed for their richness in proteins [145], featuring a well-balanced essential amino acid profile [146], high levels of essential fatty acids such as linoleic (C18:2, ω -6) and α -linolenic (C18:3, ω -3) acids [147], and significant amounts of dietary fibre and minerals [148]. Similarly, buckwheat seeds stand out as a valuable source of protein and essential amino acids [149]. Chia, also recognized as an underutilized pseudocereal [150], boasts a nutritional profile encompassing essential nutrients such as proteins, omega-3 fatty acids, carbohydrates, dietary fibre, vitamins, and minerals [151]. Teff is nutritionally dense, featuring elevated levels of protein, carbohydrates, minerals, and dietary fibres [152]. Sorghum, on the other hand, provides slow-digesting starch for improved satiety and glycaemic response, along with a unique array of polyphenols with potential applications in reducing caloric impact and acting as a naturally stable food colouring source [153]. Conversely, leaves from minor crops such as moringa and hemp, distinguished by their rich nutritional constituents, including high protein, calcium, fibre, and vitamins, as well as low fat content, have garnered attention primarily for their non-nutritive phytochemicals [154]. Regarding desert truffles, although the protein levels are typically lower than their carbohydrate content, they are considered a healthy and hypocaloric product. This is not only due to their low fat content but also because most of their carbohydrates are classified as dietary fibres with structures different from those found in cereals and pseudocereals [155].

Table 2. Nutritional composition of selected NUSs with potential of cultivation in Mediterranean areas. Proximal compositional analysis has included protein, fat, ash, dietary fibre, and carbohydrate contents.

Nutritional Component	Protein g/100 g	Fat g/100 g	Ash g/100 g	Dietary Fibre g/100 g	Carbohydrates g/100 g	References
Quinoa	13.1–18.7	1.9–9.5	2.9–3.8	9.4–22.7	41.5–77.0	[156–158]
Amaranth	13.0–17.6	4.2–8.5	2.4–4	10.0–25.0	48.0–69.0	[156,159,160]
Buckwheat	8.5–18.8	1.5–6.5	1.7–2.7	20.0–26.0	54.5–57.4	[161,162]
Chía	15–25	25–40	4–4.8	18.0–35.5	26.0–42.0	[156,163]
Sorghum	4.4–21.1	1.5–8.9	1.1–2.2	10.2–19.0	63.7–80.0	[153,164]
Teff	9.1–12.6	2.3–3.3	2.4–2.5	2.4–9.8	70.8–73.8	[165]
Moringa	18.6–39.4	1.4–5.2	7.9–8.2	2.0–10.2	29.8–51.6	[166,167]
Hemp	21–32	25.4–35.9	3.7–6.3	28.8–38.8	32.5–38.1	[168]
Desert truffle	8–29	3–7	2.6–15	1.4–13.2	46.0–83.0	[169]

In addition, all these crops are gluten-free and therefore suitable for celiac disease patients [170]. It should be considered that the nutritional composition of these crops depends on different factors such as climatic conditions, geographical location, soil attributes, and cultivation year. Therefore, the data provided in this review are ranges or average values found in the literature.

3.1.1. Protein and Amino Acids

Plant proteins, particularly those derived from pseudocereals such as quinoa, amaranth, and buckwheat, have emerged as vital vegetable protein sources that not only impart health benefits but also contribute to environmental sustainability and enhance global food security [145,171]. The proteins in these pseudocereals exhibit high quality, characterized by a balanced essential amino acid composition [172]. Quinoa, with a protein content ranging from 13.1 to 18.7%, stands out for providing all nine essential amino acids (Table 2). Similarly, amaranth seeds boast a protein content ranging from 13.0 to 17.6%, closely rivalling wheat [173]. Buckwheat presents a protein content ranging from 8.5 to 18.8% (Table 2); however, the presence of antinutritional factors like tannins and proteases impacts protein digestibility [174]. Despite this, buckwheat proteins demonstrate a well-balanced composition of amino acids, with an abundance of lysine, typically limited in most cereals [175]. Chia seeds exhibit a protein content of 15 to 25% (Table 2), surpassing many traditional cereals in protein content [150]. These proteins are particularly rich in essential amino acids such as arginine, leucine, valine, lysine, and phenylalanine.

Cereals gaining recognition for their nutritional value include sorghum and teff. Sorghum samples display protein content ranging from 4.4 to 21.1% (Table 2). However, the presence of the hard-to-digest protein kafirin and a low lysine content reduces sorghum's nutritional value for both humans and animals [176]. Teff seeds present a protein content range of 9.1 to 12.6% (Table 2) and exhibit a balanced essential and non-essential amino acid composition, with notable quantities of arginine, asparagine, and lysine, and lower amounts of leucine, phenylalanine, glutamic acid/glutamine, and proline [177].

In the case of *Moringa oleifera* Lam. leaves, consumed as highly nutritious vegetables, it stands out as a valuable source of plant protein, ranging from 18.6 to 39.4% (Table 2). These leaves contain all essential amino acids, positioning moringa leaf protein as a high-quality plant protein [166]. Similarly, *Terfezia* sp., *Tirmania* sp., or *Picoa* sp. truffles showed protein levels ranging from approx. 29 to 8% including sulphur-containing amino acids (cysteine and methionine) that generally remain limited in other plant-derived foods [178].

3.1.2. Oils and Fatty Acids

Due to the rich quality and quantity of the lipid fraction in quinoa, amaranth, buckwheat, and chia, they are referred to as alternative oilseed crops. The oil content in quinoa varies from 1.9 to 9.5% (Table 2), with a rich source of essential fatty acids such as linoleic and α -linolenic acids. Oil content in quinoa seeds consists of approximately 88–90% unsaturated fatty acids (UFAs), out of which 66–68% are polyunsaturated fatty acids (PUFAs). Saturated fatty acids (SFAs) account for around 10–11% of the total fatty acids in the seed [171,179]. Linoleic acid is the main fatty acid present in quinoa seeds (58.5–61.4%) and has been proven to provide various health benefits such as preventing cardiovascular diseases [180]. The other major fatty acids in quinoa seeds are oleic acid (18:1) (17.5–20.5%) > palmitic acid (C16:0) (9.0–10.4%) > α -linolenic acid (5.1–7.2%) [171,181]. Furthermore, the ω -6: ω -3 ratio in lipid consumption is acknowledged as pivotal, as it is linked to various chronic diseases resulting from specific dietary patterns [147]. Low ω -6: ω -3 ratios are related to a low incidence of these diseases [182]. In quinoa, the ω 6: ω 3 ratio is found to be around 6–9:1 [171,179,183]. While this ratio is higher than the ideal one (4/1), it is much better than the typical ratio found in Western diets (15–16.7/1) [184]. On the other, amaranth seeds are also a source of high-quality fat (4.2–8.5%) (Table 2) [171]. Nevertheless, the composition of fatty acids varied compared to quinoa seeds, with lower linoleic acid content (35.2–37.3%). On the contrary, oleic acid (35.4–36.7%) was higher than that in quinoa. Important differences were also found between both species for palmitic acid (21% in amaranth seeds). The differences in the fatty acid profile extend also to the ω 6: ω 3 ratio, higher in amaranth (27.5–36.1/1) than in quinoa seeds [171].

Buckwheat's lipid content is low (1.5–6.5%) (Table 2) but of great importance due to its composition. Buckwheat is a rich source of unsaturated fatty acids (74.5–79.3%) [185], primarily oleic and linoleic acids. These, along with palmitic acid, the predominant satu-

rated fatty acid, constitute the most abundant components of the lipid fraction [186]. The α -linolenic acid is also present in smaller amounts.

Chia seeds are characterized by a high fat content (25–40%) (Table 2). They are particularly rich in polyunsaturated fatty acids (~80%), including α -linolenic acid (55.2–65.9%) and linoleic acid (17.8–22.1%), with a very low $\omega 6:\omega 3$ ratio (approximately 0.3–0.4) [163,187], making them a significant source of omega fatty acids. Palmitic acid is the principal saturated fatty acid, comprising 6.9–7.8% of the total lipids [163,188].

Sorghum seeds are claimed to have lower fat content when compared to other grains [189]. However, sorghum grains contain nearly 1.5 to 8.9% lipids (Table 2), with a composition based on linoleic acid (37.8–54.4%), oleic acid (25.8–43.5%), palmitic acid (14.1–20.8%), linolenic acid (0.98–2.3%), and stearic acid (C18:0) (0.99–2.2%) [190]. In contrast, polyunsaturated fatty acids are more susceptible to oxidation, reducing the shelf life and storage stability of fats and oils. In teff grains, the total fat content has been found within the range of 2.3–3.3 (Table 2), which was higher than the content in wheat (2.2–2.3%) and rice (2.9–2.9%) [191] but slightly lower than sorghum (3.6–3.8%), oat (3.7–6.4%) [191], and millet (3.4–6.5%) [192]. The predominant fatty acids are linoleic acid (17.3–39.9%), palmitic acid (17.6–43.4%), stearic acid (17.2–22.0%), oleic acid (9.98–17.4%), and α -linolenic acid (5.9–16.5%) [193].

In the case of moringa leaves and desert truffles carpophores, they show a low oil content (of 1.4–5.2% and 3–7%, respectively, Table 2); however, their lipid composition is outstanding. Moringa oils contain approximately 80% unsaturated and approximately 20% saturated fatty acids. Oleic acid is the most abundant (66–75%, approx.) [194], making moringa oil comparable to olive oil in terms of oleic acid. In truffles, oleic (32%) and linoleic (29%) acids are major fatty acids, as the latter is the precursor of the 1-octen-3-ol compound responsible for the characteristic ‘mushroom’ aroma [195]. Among the different truffle species, *Picoa* sp. contains less palmitic and stearic acids but more oleic acid than *Terfezia* sp. [196]. Normally, vegetable oils from corn, rice bran, palm, peanut, safflower, and soybean contain less than 40% oleic acid. The high proportion of oleic acid in moringa oil renders it well suited for nutritional purposes and as a stable medium for cooking and frying [197]. Additionally, the presence of palmitic (~6%), stearic (~5–6%), and behenic (~4.4%) fatty acids has been confirmed in previous studies [198,199].

3.1.3. Ash and Mineral Content

The ash content in a food item represents the total amount of minerals present, holding significant importance for various biological activities that benefit the human body. However, in vegetable and truffle crops, the concentrations can vary widely depending on the type of soil where the plant or fungus is grown and the fertilization rates applied [148].

Among the pseudocereals, quinoa, and amaranth seeds are good sources of minerals [148]. In general, the ash content of quinoa seeds (2.9–3.8%) (Table 2) is greater than that of rice, wheat, and other grains [157,200]. In the same way, the ash content in amaranth seeds ranged from 2.4 to 4% (Table 2). The main minerals found in these pseudocereal seeds are potassium (K), phosphorous (P), calcium (Ca), and magnesium (Mg), with amaranth being the plant species that typically shows the highest amount of minerals followed by quinoa [200]. In general, higher contents of P, Ca, and Mg and lower K contents are found in amaranth compared to quinoa [171]. K, the major mineral in both types of seeds, is an important chemical element that helps to prevent muscle weakness, respiratory insufficiency, and hypotension in humans [201]. On the other hand, Fe and Ca are crucial minerals in the diet [202]. Ca contents were up to three times higher in amaranth than in quinoa [203] and higher compared to other vegetable food products considered rich in Ca, such as almonds or legumes [204]. However, quinoa seeds contain more Fe than traditional grains, although saponins and phytic acids present in the seeds could affect their bioavailability to some extent [157].

Buckwheat grains showed a total mineral proportion between 1.7 and 2.7% (Table 2), slightly lower than the other pseudocereals, such as quinoa and amaranth. In general,

P, K, Mg, and Ca are found at reasonable levels, while Fe, manganese (Mn), and Zinc (Zn) have lower concentrations in buckwheat [205]. Other authors found high levels of Mg, Ca, Fe, Zn, Mn, and Cu, while the contents of selenium (Se), molybdenum (Mo), and cadmium (Cd) are comparatively low [206]. Additionally, it has been reported that buckwheat could be used as a dietary nutrient source for Mg, Zn, Cu, Mn, and Se [206,207]. On the other hand, chia seeds as well as desert truffles are also a good source of minerals, nearly 4.0–4.8%, out of which K, Mg, Ca, and P are present in the highest amounts. In chia seeds, Fe is also present in substantial quantities, whereas in truffles its concentration varies significantly [163,178,188,195]. Ca is the most abundant microelement found in chia seeds, followed by P, Mg, and Na, according to the results obtained by [163]. The concentration of Ca in chia seeds is higher than that found in milk, and Fe concentration is higher than that found in lentils, spinach, and liver [208,209].

Sorghum and teff grains contain a relatively low ash content (1.2 and 2.5%, respectively) (Table 2). Sorghum contains a substantial amount of various minerals such as K, Mg, P, and Ca [210,211], with a bioavailability of 90% for Na and K. Thus, it provides a significant concentration of these elements for the human diet. Regarding teff grains, they present higher contents of Fe, Ca, and Cu than other cereals which is probably due to the small plant size and increased contact with soil [212]. Other authors also found higher concentrations of K followed by Mg, Ca, Na, and Fe [213]. The elevated levels of these minerals in teff grains may result from the ready availability of these elements in the soil, facilitating their uptake by the plant and subsequent translocation to various plant parts [214]. In addition, teff is noted for its high Fe bioavailability [215], which is associated with a lower prevalence of Fe-deficiency anaemia in populations where teff is consumed regularly.

Moringa leaves showed ash contents of around 8% (Table 2), including K, Na, Mg, and Fe as the predominant minerals, in decreasing amounts. The high value of ash indicated that moringa leaves are a good source of mineral elements. Thus, the contents of these minerals in moringa leaves could satisfy the recommended daily intake (RDI) [216]. Furthermore, it was reported that the mineral content in moringa leaves is significantly positively correlated with the elemental mineral concentration of the soil, and differences in the stage of growth of the moringa leaves could also contribute to the variations of the elemental mineral concentrations [217].

3.1.4. Fibre and Carbohydrates

The pseudocereals in whole grain form are considered a rich source of dietary fibre [218]. The total dietary fibre (TDF) content in different varieties of quinoa, amaranth, and buckwheat ranged from 9.4 to 22.7%, 10 to 25%, and 20 to 26%, respectively (Table 2). In the case of buckwheat grains, they show more fibre than other pseudocereals [219], and these values surpass those obtained for other grains such as wheat, rice, and corn [173].

Chia seeds contain 18 to 35.5% dietary fibre (Table 2), placing them at the top of the list in providing more fibre. The TDF content of sorghum and teff grains differs, with contents between 10.2 and 19% and 2.4 and 3.9%, respectively (Table 2), found in the literature. However, higher values (8.6 and 9.8%) [220] were also reported for teff grains, likely associated with different methodologies or varieties used. Additionally, the fibre content in moringa leaves and desert truffles was relatively lower compared to most forage plants being 2–10.2% for moringa, 1.4–12.6% for *Terfezia* and 6.5–13.2% for *Tirmania* species. The dietary fibres in truffles differ from those in other plant crops, primarily comprising chitins and β -glucans with distinct branching boundaries compared to cereals (Table 2) [58,155,221].

On the other hand, carbohydrates are essential for the proper functioning of the body and should make up 50% of our daily calorie intake [221]. In quinoa, amaranth, buckwheat, and desert truffles, carbohydrates were the major macronutrients. The literature consulted found values between 41.5 and 77.0%, 48 and 69%, 54.5 and 57.4%, and 46 and 83%, respectively (Table 2), equal to or slightly lower than other cereals such as wheat, rice, and corn grains (71.7, 81.7, and 74.3 g 100 g⁻¹, respectively) [173,178,222]. However, chia

seeds showed a lower carbohydrate content (26–42%) (Table 2) compared to the other pseudocereals. It should be noted that a lower average carbohydrate content in chia seeds could be due to the high values obtained for all other nutritional parameters. Thus, since carbohydrate contents of plant food are calculated by difference, an increase in protein, fat, and moisture contents will ultimately affect the carbohydrate contents and contribute to observed increases. Sorghum and teff grains and moringa leaves are also rich sources of carbohydrates, with contents found in the literature ranging from 63.7 to 80%, 70.8 to 73.8%, and 29.8 to 51.6%, respectively (Table 2). Importantly, the main carbohydrate component of all crops studied is starch, constituting around 50–70% of it [157,223].

3.2. Bioactive Compounds

The widespread utilization of pseudocereals is attributed to their favourable nutritional value and the presence of bioactive compounds. These compounds, notably flavonoids, phenolic compounds, and peptides, exhibit numerous beneficial effects on human health [224]. However, the extraction conditions significantly impact the quantity of bioactive compounds extracted (solvent, extraction time, and temperature), leading to substantial variations in results reported in the literature. Consequently, this literature review refrains from providing specific values for each of the compounds found in different crops.

3.2.1. Phenolics

Phenolic compounds, characterized by hydroxyl groups on aromatic hydrocarbon rings, constitute secondary plant metabolites. This compound family includes phenolic acids, flavonoids, phenols, quinines, coumarins, lignans, phenylpropanoids, and xanthones [225], existing in both free and bound forms. These compounds, particularly flavonoids, phenolic compounds, and peptides, confer various health benefits through antioxidant capacity, scavenging free radicals, and preventing oxidative stress [150,172].

Pseudocereals, like quinoa, amaranth, and buckwheat, exhibit rich concentrations of bioactive phytochemicals. Quinoa, in particular, boasts flavonoids such as quercetin and kaempferol, contributing to antioxidant properties and health benefits [200,226]. Buckwheat stands out as the richest source of phenolic acids, predominantly found in free forms, influencing antioxidant activity [227]. Chia seeds contain significant phenolic compounds, including various flavonoids, but also diterpenoid phenolics such as rosmarinic acid, both contributing to the antioxidant properties of these seeds [224,228]. Sorghum, uniquely rich in bioactive constituents, presents high concentrations of phenolic compounds such as ferulic and protocatechuic acids, as well as flavonoids like quercetin and kaempferol [229,230]. Teff grains prominently feature ferulic acid and flavone derivatives, with additional phenolic compounds such as protocatechuic and cinnamic acids [231]. Simple phenolic compounds and flavonoids have been extensively documented in desert truffles, as well as in edible mushrooms. However, there are conflicting reports regarding the ability of these fungi to either absorb or synthesize these compounds, especially flavonoids [195,232].

In summary, high-value seed flours, including those from buckwheat, quinoa, sorghum, and teff, exhibit elevated levels of phenolic substances and flavonoids, surpassing whole-grain cereal flours in antioxidant and anti-inflammatory effects [233,234]. Moringa leaves are rich in phytochemicals, with phenolic acids, flavonoids, and anthocyanins contributing to their nutritional profile [235,236].

3.2.2. Phytosterols

Phytosterols, lipophilic compounds structurally akin to cholesterol, play a vital role in cholesterol metabolism and exhibit potential health benefits against cancer, diabetes, hepatoprotective effects, and cardiovascular diseases through their cholesterol-lowering characteristics [150]. In quinoa seeds, major phytosterols include β -sitosterol, campesterol, and stigmasterol, with higher quantities than those found in barley, rye, millet, and corn [180,237]. Amaranth seeds also contain these phytosterols, with β -sitosterol as the

predominant compound [238]. Interestingly, according to [239], β -sitosterol, a significant phytosterol, may possess anti-diabetic properties.

Buckwheat grains feature β -sitosterol as the primary phytosterol, along with others like campesterol, avenasterol, and D-7-stigmasterol. Additionally, the infrequently observed cycloartenol was identified in buckwheat samples [240]. Chia seeds similarly contain phytosterols such as campesterol, stigmastanol, stigmasterol, D5-avenasterol, and β -sitosterol [241]. Sorghum grains encompass common sterols, including β -sitosterol, campesterol, and stigmasterol, positioning sorghum as a rich source of phytosterols compared to fruits, vegetables, and other cereals [230].

While there is limited evidence in the literature for the presence of phytosterols in teff grain, one study reported the existence of β -sitosterol and β -sitosterol-3-O- β -D-glucoside in teff grains [242]. Also, moringa leaves have emerged as a rich source of phytosterols, with β -sitosterol being the major compound, followed by stigmasterol, campesterol, and Δ 5-avenasterol [154].

Fungal sterols in desert truffles remain inadequately investigated. Some reports suggest that the *Terfezia* genus predominantly contains brassicasterol (98%), with ergosterol constituting less than 2%. However, ergosterol is a major constituent in fungal membranes [243].

3.2.3. Betalains

Betalains, nitrogen-compound plant pigments, have garnered attention in functional foods due to their bioactive properties, including antioxidant and anti-inflammatory effects [244,245]. Betalains demonstrate superior antioxidant activity compared to polyphenols and are exclusive to plants of the order Caryophyllales, including certain cultivars of the family Amaranthaceae [246].

Quinoa seeds derive their yellow, black, and red colours from betalains, making quinoa a potential crop for betalain extraction, although some varieties may lack measurable amounts of betalains [244]. Betalains are further categorized into red-violet betacyanins and yellow-orange betaxanthins [247]. The incorporation of these pigments in food products is officially certified by the European Union (additive E-162) and the US FDA [248]. Amaranthine and isoamaranthine are two betacyanins found in amaranth, while red and black quinoa seeds contain betacyanins such as betanin and isobetanin [249]. Some studies report low amounts of betalains in specific amaranth varieties [250].

3.3. Antinutritional Factors

Certain phytochemical compounds found in seeds, such as phytates, saponins, lectins, and protease inhibitors, can exert both positive and negative effects on human health. At low concentrations, these compounds may contribute to well-being by exhibiting antioxidant properties, anti-inflammatory effects, and anti-cancer or immune-enhancing effects. However, when present in higher concentrations, these phytochemicals can interfere with the absorption of essential minerals, induce haemolysis, disrupt protein digestion and absorption, or harm the gut wall, potentially leading to increased intestinal permeability. While a moderate intake of these compounds offers health benefits, excessive consumption should be avoided to maintain a balanced diet and ensure optimal nutrient absorption. Additionally, oxalates and nitrates, naturally occurring compounds in various plant foods, are generally considered harmless when consumed in moderation. Overall, various antinutritional factors are present in the edible parts of NUS crops, and their influence should be considered during the assessment of nutritional characteristics. In this regard, it is crucial to highlight the very limited knowledge about the presence of antinutrients in the majority of the NUS crops analysed in this work. This underscores the necessity of conducting additional research on this topic. Nonetheless, a concise analysis is presented herein, and the findings are summarized in Table 3. Furthermore, to mitigate the presence of antinutrient compounds, various actions or strategies can be employed during food processing, as outlined in Table 3.

In the case of phytic acid, also known as myo-inositol hexakisphosphate, it is commonly found in grains and seeds as phytates. While phytic acid intake has reported benefits such as antioxidant function and the prevention of heart diseases and cancer, it is notorious for inhibiting the absorption of minerals, proteins, and trace elements [251,252]. The formation of insoluble complexes in the gastrointestinal tract renders them non-absorbable, while enzymatic degradation of phytic acid by phytases improves the nutritional value of cereal, pseudocereal, legume, and oilseed products [253–255].

Saponins, characterized by a linear arrangement of hexose or pentose glycoside units linked to sapogenin aglycones, form complexes with proteins, lipids, zinc, and iron, exhibiting a haemolytic effect, albeit absorbed in limited quantities; they can enhance membrane permeability, potentially aiding in intestinal food intake and drug assimilation [256,257]. However, their bitter taste poses a challenge to the widespread utilization of quinoa or amaranth seeds [258].

Another antinutrient compound family, protease inhibitors, abundant in legumes, cereals, pseudocereals, and oilseeds, forms stable complexes with proteolytic enzymes, influencing human physiology in both beneficial and detrimental ways [259,260]. While they are considered antinutritional factors due to their inhibitory effects on digestive enzymes, quinoa and amaranth seeds contain minimal amounts, unlike commonly consumed grains [256]. Conversely, the limited digestibility of buckwheat protein is attributed to various factors, including protease inhibitors, tannins, and α -amylase inhibitors [162]. Oxalates, found in various plant tissues including leaves, stems, hypocotyl-root, and seeds, hinder the absorption of calcium and magnesium, contributing to kidney stone formation. Amaranth and quinoa are considered high oxalate sources, with the highest concentrations observed in leaves and stems. However, the majority of oxalates are found in an insoluble form, and due to their high concentrations of calcium and magnesium, this may result in low oxalate absorbability [261]. Nonetheless, to mitigate the risk of stone formation, limiting oxalate consumption to below 100 mg per day is advisable, with individuals prone to kidney stones potentially requiring further reduction to less than 50 mg per day [262]. Cooking grains before consumption is essential to reduce soluble oxalate intake, as soluble sources are more readily absorbed, posing a risk for kidney stone development [263]. In the case of lectins, whose toxicity is contingent upon the quantity consumed and the frequency of ingestion, they are proteins that can bind to cells, leading to structural and physiological damage in tissues upon consumption, resulting in potentially adverse health effects [264]. They have been found in quinoa, amaranth, or desert truffle [265–267].

Nitrates, while not inherently harmful, can convert to nitrites during digestion, potentially leading to the formation of nitrosamines in the stomach [268]. The World Health Organization (WHO) has not specified a recommended daily allowance (RDA) for nitrates, but the European Food Safety Authority (EFSA) has set an acceptable daily intake (ADI) at 3.7 mg/kg of body weight/day, approximately 260 mg/day for a 70 kg adult. Generally, grains or seeds from selected NUS crops fall below this threshold. However, given the lack of information on nitrate content in the green parts of these crops, it is advisable to conduct nitrate content analyses when consumed as vegetables, as vegetables are a primary source of nitrates in plant-based foods.

Table 3. Mainly antinutritional factors in selected crops and the treatments to inhibit their action.

Antinutrient Factor	Phytic Acid	Saponins	Protease Inhibitors	Lectins	Oxalates	Nitrates	References
Crop and Usual Treatment	g/100 g	mg/100 g	mg/100 g	g/100 g	mg/100 g	mg/100 g	
Quinoa	0.50–1.39	1.90–6.24; 199.07 629.7–1633.3 **	2.4 * g/kg	5.2 HU [†] ; 9.3%	4.31; 6.17–9.45	21.83	[158,258,266,269]
Amaranth	0.61–1.34	5.33–10.66 HU/mg 79–186	0.519–1.016 TIU/mg	0.5–7.3; 11.2–130.6 HAU/mg	178–278; 46–152	48–84	[258,261,267,270, 271]
Buckwheat	0.63; 1.7	0.058	55 α U/mg protein; 94 α U/mg protein	NYR (+)	93–155	NYR (–)	[161,272–274]
Millet	0.12–0.92; 0.336–0.649; 0.519–0.784; 0.64–0.81	0.039–0.044	0.385–0.985 TIU/mg flour	NYR (–)	11.3–20.0	NYR (–)	[164,272,275–277]
Teff	0.682–1.374; 1.35	722	NYR	NYR (–)	3.50–5.27 ***	NYR (–)	[278–281]
Chia	1.5–2.6	616	445 TIU/mg protein	NYR	NYR (–)	NYR (–)	[163,282]
Moringa (seeds)	0.115 δ ; 2.54	1080 δ	15.4; 120 δ	NYR (+)	2.21 δ	NYR	[283,284]
Hemp	2.66–3.08	123.54 α	3.6% β	NYR	NYR	NYR	[285–287]
Dessert truffles	0.581	12.38	NYR	18.11 HU	NYR	NYR	[265,288]
Usual Treatment to diminish the antinutrient factor concentration or deactivation. In general, antinutrient levels can be controlled by cultivar selection and breeding, field management, and processing	Soaking, Steeping, Malting, Fermentation, Germination, Extrusion, Popped grains, Phytase activation by sprouting, Germination, or lowering the pH Inclusion of exogenous phytases Agronomic conditions Genetic engineering	Germination Popped seeds Extrusion Sweet varieties (<110 mg/100 g) Agronomic conditions	Cooking around 100 °C or at higher temperatures Popped grains Extrusion	Cooking around 100 °C or more Soaking Extrusion Popped seeds Germination	Water cooking-soluble oxalate, Soaking/Steeping Control of fertilization/soil	Water cooking Soaking/Steeping Control of fertilization/soil	

* g/kg: the amount of enzyme inhibited per kg seed meal; HU[†]: Haemagglutinating Units $\times 10^{-6}$ per kg of seed meal, treated rat blood cells; TIU/mg, HAU/mg, and HU/mg: Trypsin Inhibitory Units, Haemolytic Activity Units, and Haemolytic Units per mg of the sample on a dry weight basis. ** expressed in sapogenins (the hydrolysed product of saponins); *** the amount in straw; δ defatted seed flour; α in the extract; β trypsin inhibitor; NYR not yet reported; presence: (+); apparently at a lower amount than other grains/seeds: (–).

3.4. Nutritional Challenges and Considerations

Currently, the global community confronts a spectrum of nutritional challenges encompassing malnutrition and hunger, juxtaposed with concerns regarding obesity and diet-related ailments such as diabetes and cardiovascular diseases. Concurrently, the exigencies of climate change and water scarcity necessitate attention, alongside underutilized food wastes, byproducts, and the inadequacy in meeting the protein demand of the world population [289]. In this regard, it becomes necessary, and almost mandatory, to leverage and capitalize on the scientific and cultural resources and knowledge at our disposal. Hence, aligning with the central theme of this review, the possibilities presented by the utilization and revaluation of NUSs are well recognized. These possibilities contribute to enhanced biodiversity and sustainable development, not only at the agronomic level but also within the industrial context. The ultimate goal is to achieve improved production, enhanced nutrition, a healthier environment, and an overall better quality of life for all, leaving no one behind [290].

Beyond popular labels such as ‘superfoods’ or ‘miracle seeds or trees’, the crops discussed in this work exhibit a chemical composition in macro- and micronutrients that are either similar to or superior to the four main plant crops dominating the global agrifood system—namely, rice, wheat, soybeans, and corn. Notably, the production of these major crops is concentrated in a few countries globally (USA, China, India, Argentina, Brazil, and Russia). A distinctive advantage of many emerging crops highlighted in this study is their superior adaptability to abiotic stress conditions, as detailed in Section 2 on agronomical

aspects related to NUSs for the Mediterranean region [10,291]. Hence, it is imperative to undertake more comprehensive studies on the introduction and advancement of these emerging crops, involving genetic improvement, hybridization, artificial intelligence, and enhanced control over abiotic stresses and pests. The objective is to diversify both the plant raw materials dedicated to food and the national and local production levels. This strategic approach aims to mitigate recent food crises, such as those experienced during the COVID-19 pandemic or conflicts in regions like Ukraine and Gaza. The acquisition of greater agronomic knowledge will empower governments to implement agricultural programs and policies that positively impact nutrition.

Conducting research and disseminating findings on the nutritional aspects of emerging crops, including food composition, dietary assessments, and human nutritional requirements, is essential for integrating these crops into society and highlighting their health benefits. This effort addresses the crucial nutritional challenge of meeting the rising global protein demand through sustainable production, diversified protein sources, and efficient resource use.

As stated in Table 2, the seed protein content shown by some can exceed 20% (e.g., chia [150] or hemp [292]). Furthermore, all exhibit higher protein contents than commonly consumed cereals [145,171,176]. In this context, an underutilized and valuable resource to be revalued and exploited is the protein contribution from other parts of the plant besides the seeds, notably the leaves. Crops like moringa, for instance, present protein values ranging between 16 and 40% in this regard [166] or desert truffles, with values ranging between 14 and 29% [178,195]. The protein content can be further enhanced through the extraction and commercialization of other components applicable in the food industry, such as oil, starch, or fibre fraction. This process results in protein concentrates or isolates [293], which can serve as plant-based ingredients in products with high protein content catering to vulnerable populations such as the elderly, malnourished children, or pregnant women.

Recent advances in food and health emphasize the potential of peptides, mono- and polyunsaturated fatty acids, pigments, and low glycaemic index starches from these crops, suggesting promising research directions. This progress requires enhancing and updating extraction technologies, such as microwaves, ultrasound, high pressures, fermentation, tribo-electrostatics, and supercritical fluids. Additionally, there is ongoing exploration and utilization of extraction methodologies, such as those assisted by enzymes or eutectic solvents [294–299].

Another significant nutritional challenge associated with these crops is the utilization or revaluation of antinutritional compounds, particularly found in the seeds. Examples, as described earlier in this review, include saponins in quinoa [300], tannins in sorghum [301], or phytic acid in amaranth, chia, and millet [302]. Initially utilized by the plant as a defence or survival mechanism against pests, predators, or adverse weather conditions, these components were traditionally deemed exclusively antinutritional for decades. However, recent scientific studies have shifted the focus, attributing certain health benefits at low concentrations, such as antioxidant or anti-inflammatory properties [303–305]. The development of a more friendly and profitable technology with these raw materials at the industrial level will increase this revaluation and help to enhance the benefits of these crops at the food level.

3.5. Technological Aspects

The surge in quinoa demand is attributed to its distinctive nutritional and techno-functional characteristics. Formerly stigmatized as a symbol of rural poverty and indigeneity, quinoa has gained superfood status [306]. Notably, quinoa starch, with its high peak viscosity, holds promise for enhancing techno-functionality in gluten-free dough formulations and reinforcing edible biodegradable films. Research by Cordoba-Cerón et al. [307] showcased improved techno-functional properties in protein-rich pasta when incorporating defatted high-protein quinoa flour. Fungal fermentation has emerged as a viable method to enhance the bioactivity of flours, as demonstrated in quinoa, leading to increased levels of

resistant starch and fibre [308]. Amaranth protein surpasses cereals in biological value and exhibits favourable techno-functional properties, including high solubility and foaming capacity. Studies by Kierulf et al. [309] demonstrated that amaranth and quinoa flours, retaining a portion of their protein content, possess excellent techno-functional properties, qualifying them as natural alternative surfactants for stabilizing food emulsions [309].

Chia seeds, endowed with intrinsic technological characteristics such as high water-holding capacity and robust gelling properties, are suitable as fat replacers in baked and processed meat products [310,311]. While other NUSs hold nutritional interest in food formulation, their technological applications present challenges. Hemp seed protein (of approximately 20–25%), characterized by low allergenicity and a complete amino acid profile, faces limitations in techno-functional properties for successful formulation in food products [312]. Extrusion technology has proven successful in overcoming these limitations, particularly for products mimicking meat [313]. Moringa encounters obstacles in food formulation due to organoleptic characteristics such as an astringent bitter taste and unpleasant smell. Encapsulation technology has been proposed to address these challenges while enhancing thermal stability [314]. Alternatively, the use of moringa flower buds in powdered format has been suggested for direct incorporation into pasta products, enhancing nutritional profiles and antioxidant activity [167].

Desert truffle (*Terfezia arenaria*), although not listed as an edible food in the European Union, presents an intriguing opportunity for sustainable plant-based meat analogues due to its unique aroma and composition analogous to meat [315]. Nevertheless, desert truffles have been successfully employed to enhance the nutritional composition and antioxidant content of biscuits [169].

3.5.1. Culinary and Processing Aspects

The comprehensive examination of the culinary and processing characteristics of nutrient-rich and stress-resilient emergent crops provides insights into their potential applications in traditional cuisines and their nutritional benefits for promoting their consumption in conventional culinary practices and improving nutraceuticals for high-quality functional foods [316]. Quinoa, for example, is extensively utilized as a grain or flour in the preparation of bread, soups, and cooked products, either independently or in conjunction with other flours [54]. Given its compatibility with traditional cuisine culture, quinoa seamlessly integrates into South American dishes, lending its nutty flavour and fluffy texture to traditional Peruvian quinoa stews and contemporary salads, showcasing adaptability across diverse global cuisines [317]. Moreover, expanded or puffed quinoa introduces another variation of traditional quinoa products. This involves subjecting the grain to a high temperature and pressure, resulting in a quinoa pop. High temperatures and long processing times, common in energy bars and instant cereal production, can cause a decrease in protein and linoleic acids, impacting the nutritional quality of the product [318].

Chia seeds stand out for their flavour and impressive water-absorbing properties, offering a versatile, gel-like consistency that makes them a favoured ingredient in beverages, puddings, smoothies, and various sweet and savoury recipes for a nutrient-rich boost [319]. Their combined nutritional and technological attributes are optimal for use in baked goods as an egg replacement ingredient, or in the production of gluten-free bread or pasta, owing to their hydrocolloid and thickening properties [320]. Numerous studies have explored the incorporation of chia seeds and chia mucilage gel as substitutes for eggs, oil, or fat in diverse food products resulting in more nutritious items with acceptable sensory qualities, especially evident in pound cakes, reduced-fat bread baking, and improved quality in sweet wheat doughs [321–323]. Additionally, plant-based meat analogues offer an innovative strategy for emulating the sensory and nutritional attributes of traditional meat products. With an increasing demand for these alternatives, there is a growing exploration for raw materials capable of replicating meat products. Chia seeds emerge as a promising candidate. Defatted chia flour, known for its high protein content, is valuable, especially due to the chia mucilage fraction. This can serve as an effective ingredient contributing to enhancing

the textural properties of plant-based meat alternatives [324]. Quinoa and chia, as viable options for reintegrating into the food value chain, offer coproducts that, when utilized in meat production innovation, especially in controlling lipid oxidation, can play a crucial role in enhancing shelf life [325,326].

Amaranth, with its distinctive earthy and peppery flavour profile, has become a culinary cornerstone in Central and South America, India, and Africa. In Mexico, it is favoured for crafting delectable, sweet treats, while in India, it features a myriad of both savoury and sweet dishes. Its minuscule grains contribute a unique texture and nutritional richness to a diverse range of culinary creations, including soups, stews, and baked goods. Recently, there has been a resurgence in the popularity of amaranth, marking a revival five centuries after it served as a staple food for ancient Mesoamericans [327]. Today, amaranth is easily accessible in a variety of commercial food products worldwide, showcasing its enduring popularity and recognized nutritional benefits [328]. Recent developments involve integrating grain amaranth into energy foods, including breakfast items, bread, crackers, and pancake mixes. Additionally, it can be enjoyed popped as a wholesome snack option [327].

The scientific literature robustly supports the culinary utilization of *Moringa oleifera*, highlighting its potential as a valuable source of phytochemicals with diverse applications in functional and medicinal foods [329–332]. The incorporation of moringa into bakery products has shown significant nutritional benefits, such as improved digestibility, enhanced dough stability, increased antioxidant capacity, and extended preservation [333].

Additionally, moringa's antioxidant and antimicrobial properties make it suitable for reducing lipid oxidation and inhibiting bacterial growth, thereby enhancing the nutritional content of food items like fish or chicken burgers, cream cheese, or mutton patties [334–336]. Protein extracts derived from fractions of moringa seeds have been reported as a potential ingredient to uphold texture and reduce syneresis in yoghurt production [337]. As the trend in food fortification is increasing, moringa has been explored as a functional ingredient in fortification in protein and fibre with acceptable sensory quality in products such as pasta, cookies, bread, or tempeh-like fermented products [338–340]. Incorporating moringa into the food industry offers a significant opportunity to tackle nutritional deficiencies, particularly in regions with extreme poverty or food scarcity [341]. By harnessing moringa's nutritional richness, food manufacturers can develop products that provide nourishment and enhance the well-being of communities in need [342].

Also, buckwheat, millet, and teff are noteworthy emerging crops, particularly as gluten-free grains, making them suitable for food industrial applications that prioritize nutritious and gluten-free food products [343]. Buckwheat is experiencing increased popularity in Eastern Europe, featuring in dishes like blini, and Asian cuisine through soba noodles. Its nutritional value and presence of bioactive compounds like peptides, flavonoids, and phenolics position it as a promising crop for developing gluten-free products such as cookies, bread, or focaccia to cater to individuals with celiac disease [344,345]. In Central Asia, where buckwheat is a dietary staple, a strategy to decrease meat consumption involves introducing 3D food printing in the formulation of gluten-free, plant-based meat alternatives creating and customizing balanced foods [346]. Aligned with buckwheat, millet, a dietary staple in many African and Asian nations, is utilized in porridges, flatbreads, and pilafs, and its gluten-free nature also makes it a suitable alternative for individuals with gluten sensitivities [345,347]. Also, the increasing global demand for fermented foods has opened the possibility of utilizing multigrain millet, given its substantial probiotic properties [348]. For instance, fermenting whey cereal beverages with millet and moth bean produced an innovative dairy alternative with desirable functional and sensory traits, maintaining acceptability for 12 days [349]. Originating from Ethiopia, teff flour enhances the technological process and quality of wheat bread, boosting its nutritional value and specific qualities by improving structural and mechanical parameters [350]. Additionally, teff serves as a substitute for breadcrumbs and soybean meals in conventional meat products, enhancing moisture retention and sensory characteristics, making it suitable for

individuals with celiac disease and soy protein sensitivity [351]. Furthermore, extrusion cooking has been investigated to create expanded teff-based products mixed with chickpea flour, showing promising physical, functional, and sensory attributes, indicating potential applications in the food industry [352].

Hemp seeds, known for their nutty flavour, play a prominent role in global cuisine, whether sprinkled over salads, blended into smoothies, or incorporated into baked goods contributing to a boost in protein and omega-3 fatty acids. Various processes for producing value-added ingredients from hemp seeds, including oil and protein, are well suited for targeted food applications, presenting market-led opportunities. Examples include hemp flour and derived oil for fortified food products, as well as the extraction of protein isolates and concentrates for plant-based meat and dairy alternatives [353]. For instance, incorporating hemp flour in cookies, which imparts high antioxidant activity, or substituting starch with 20% hemp protein concentrate to enhance the quality of gluten-free bread [354,355] or fermenting hemp seeds to produce fermented drinks with prebiotic activity, has resulted in an alternative dairy product [356]. Furthermore, both dry and high-moisture extrusion techniques have resulted in hemp-based products, including energy bars and meat analogue formulations, exhibiting high sensory acceptability [313,357].

Finally, in Middle Eastern cuisine, desert truffles are revered as a delicacy, imparting a distinctive earthy and umami flavour to dishes. They are savoured through various preparations, including frying, boiling, or as a meat replacement in cooked dishes [155]. From rice-based delicacies to sauces and desserts, the adaptability of desert truffles showcases their enduring appeal in both traditional and contemporary culinary settings [315]. Due to the nutritional profile of this crop, which encompasses a wide range of bioactive chemical constituents, it has been utilized as a therapeutic source [358].

3.5.2. Technological Treatments

Despite their increasing use in the food industry, minor cereals and pseudocereals often have poor functional properties for certain food manufacturing processes. To overcome these shortcomings, flours or grains of these species have been subjected to various technological treatments that induce structural modifications of their main biopolymers (starch and proteins), leading to significant improvements in their techno-functional properties. Among the technological treatments applied, the physical modification of starches, flours, and grains has become increasingly important, as it allows the functional improvement of these matrices while maintaining the status of food ingredients so that they can be incorporated into the formulation of clean label foods.

Heat Moisture Treatment (HMT) is one of the most commonly used and is carried out at temperatures above the starch gelatinization temperature while maintaining a moisture content of less than 35%. HMT has been applied to buckwheat starch and flour to improve their thermal stability and nutritional properties, making them more suitable for pasta and baked products [359,360]. The treatment of quinoa with HMT has also been proposed to modify the gelatinization behaviour, relative crystallinity, and Fourier Transform Infrared (FTIR) spectra of starch, improving its range of applications as a food thickener [361], and to increase starch/protein interaction in flours, thereby reducing the estimated glycaemic index [362]. Microwave-assisted hydrothermal treatment (MWT) has emerged as a promising alternative, as it allows for faster treatment and reduces the amount of energy required in comparison to conventional thermal heating. Studies on buckwheat grains have shown that the moisture content during MWT modulates the final properties of the flour, thus widening the range of applications for buckwheat in the food industry, such as soups and sauces, cakes, frozen desserts, and bakery products [363]. The use of MWT-treated buckwheat flour in the production of gluten-free products resulted in improved dough viscoelasticity and bread-specific volume, delayed staling, and modulated glucose release kinetics [364]. In the same way, MWT is effective in modifying the functional properties of quinoa flours by altering their protein and starch components, thus influencing the rheological behaviour of bread doughs, which affects breadmaking performance [365].

Annealing (ANN) is an alternative hydrothermal treatment in which starchy materials are treated in excess water between the glass transition and gelatinization temperatures. This technology has been successfully used to improve the thermal stability of buckwheat starch while modifying its *in vitro* digestibility, increasing slow digestible starch and resistant starch [366]. Previous studies on the ANN treatment of amaranth starch have shown that the formation of cross-links between starch chains (amylose–amylopectin) alters the physicochemical properties of amaranth starch with improved thermal stability, increasing its potential use as an ingredient in the food industry [367].

High-intensity ultrasound (US) has also been used for the treatment of starches and flours, which are generally suspended in an aqueous medium in which they undergo physical modification by the phenomenon of cavitation. This technology has been applied to minor cereals such as canary seeds to improve the functional properties of the flours [368]. Sonicated defatted canary seed flours proved to be versatile ingredients with desirable surfactant properties for a wide range of applications such as cakes, muffins, and bread where texture enhancement, stability, and consistency are critical.

High hydrostatic pressure (HHP) is a technology generally used in the treatment and preservation of foodstuffs, which has recently been applied to the treatment of cereals and pseudocereals due to its ability to exert significant effects on starch and protein polymers. The ability to modify the hydration and pasting properties of quinoa and buckwheat starches, as well as the texture and rheology of the gels formed from them, has recently been investigated [369–371]. HHP treatments have also been applied to teff and buckwheat flours [372] and whole buckwheat grains [373], improving the phenolic content of the resulting flours and their structuring capacity in gluten-free systems, depending on the treatment variables.

4. The Health Benefits of NUSs for the Mediterranean Region

Globally, developed countries are experiencing an increase in the incidence of major non-communicable diseases such as cardiovascular disease, diabetes, and obesity. This increase has led to a significant rise in morbi-mortality and healthcare expenditure [374]. The search for novel foods that not only provide balanced nutrition but also have multiple beneficial health effects has made the study of functional foods an incipient field. In this context, the health-promoting aspects of bioactive compounds found in NUS, such as quinoa, chia, amaranth, hemp, moringa, sorghum, buckwheat, teff, or dessert truffles, have been elucidated. Quinoa and amaranth, in particular, not only have commendable nutritional profiles but are also significantly richer in both micro- and phytonutrients than conventional grains, which has led to a growing interest in these crops for potential integration into functional foods and nutraceuticals [375]. Recent studies have focused on quinoa and amaranth intending to identify numerous bioactive compounds, including phenolic acids, flavonoids, betalains, carotenoids, and tocopherols. These compounds have been found to have significant antioxidant, -inflammatory, -hypertensive, and -hyperglycaemic and lipid-lowering properties, and are the main mechanistic basis for their beneficial effects against cancer, neurodegeneration, and metabolic disorders such as obesity and diabetes, which are major risk factors for cardiovascular disease [375]. By and large, quinoa and amaranth are generally considered safe for consumption and are not commonly associated with allergies, although isolated cases have been reported [376]. However, more research is needed to fully understand the prevalence and mechanisms of amaranth and quinoa allergy.

Chia seeds also have excellent nutritional properties [150]. In addition to their high fibre content and balanced range of macro- and micro-nutrients, chia seeds are known for their high concentration of α -linolenic acid, which serves as a precursor for the synthesis of ω -3 fatty acids with long chains such as DHA and EPA. It is now well established that omega-3 polyunsaturated fatty acids (PUFAs) confer several health benefits. In particular, the consumption of omega-3 fatty acids has been associated with a decrease in triglyceride and non-high-density lipoprotein cholesterol levels, supporting the protective role of these

fatty acids against cardiovascular events [377]. In addition to reducing dyslipidaemia, omega-3 fatty acids also exert protective benefits against atherosclerosis, inflammation, diabetes, and cancer, although some research findings are controversial and highlight the need for more clinical studies to clarify the beneficial effects of ω -3 PUFAs [378]. Chia seeds provide health benefits due to bioactive compounds such as phenolic molecules, tocopherols and carotenoids. These compounds have antioxidant properties that contribute to health benefits, particularly in the fight against diseases such as diabetes and cancer [224]. Chia seeds also contain phytosterols, which are mainly associated with reduced cholesterol and hepatoprotective effects. They also play a beneficial role in diabetes and cardiovascular diseases [241].

In recent years, the distinctive components and biological activities of moringa have been extensively studied and documented, highlighting its potential health benefits and therapeutic applications [379]. The whole *M. oleifera* plant, including leaves, fruits, pods, flowers, seeds, and roots, exhibits a range of biological activities. Leaves are rich in active constituents such as flavonoids, polyphenols, terpenoids, phenylpropanoids, fatty acids, sterols, and alkanes, as well as vitamins and minerals [380–383]. Among these, flavonoids and polyphenols are the primary bioactive molecules, showing antioxidant, -cancer, -sepsis, -inflammatory, -hypertensive, and -diabetic and hepatic lipid-lowering properties and beneficial effects on obesity-related reproductive diseases. In addition to fatty acids, essential amino acids, and flavonoids, the seeds also contain isothiocyanates, which play an important role as anti-inflammatory, -oxidant, -bacterial, and -cancer agents [384]. Furthermore, newly discovered components extracted from the stem of the plant, such as tricosanoic acid, cholest-5-en-3-ol, stigmasterol, and gamma-sitosterol, have shown potent antifungal activity [379]. Potent antioxidants such as tocopherols, ascorbic acid, flavonoids, and carotenoids have also been extracted from *M. oleifera* flowers [385].

Hemp seed, derived from the plant *Cannabis sativa* L., has a rich history of use in Asia dating back to prehistoric times. Recently, countries such as the United States, Canada, and Australia have now legalized the farming and use of hemp seed, provided that its tetrahydrocannabinol content remains below 0.3%. This legalization has increased interest in hempseed due to its recognized nutritional benefits and perceived pharmaceutical applications [386,387]. Historically, the economic importance of hemp has been in the use of the fibre-rich stem, valued to produce textiles, clothing, and paper goods, while its seeds have remained largely underutilized. However, in recent years there has been a surge of interest in exploring the nutritional and pharmaceutical properties of hemp. In particular, hempseed oil has the highest concentration of PUFAs of any vegetable oil. It is widely accepted that an increased intake of PUFAs is associated with a reduced risk of several health conditions, including cardiovascular disease, cancer, rheumatoid arthritis, hypertension, inflammatory disorders, and autoimmune diseases. Of note is the highly desirable ω -6: ω -3 ratio found in hempseed, which typically ranges from 2.5 to 3.5/1, in contrast to the ω -6: ω -3 ratio found in typical Western diets, which normally ranges from 15 to 17/1. The importance of the ω -6/ ω -3 ratio in human health is well documented, with a high ratio (>10:1) in the human diet being associated with an increased risk of cancer, inflammation, and cardiovascular disease. On the contrary, a ratio lower than 2.5:1 has been associated with a reduced risk of chronic diseases, the suppression of cancer cells, and lower mortality [184]. In addition to its oil content, cannabis seeds contain two different phenolic compounds: lignanamides and hydroxycinnamic acids. In vitro studies have extensively documented the antioxidant properties of hempseed lignanamides, including compounds such as N-trans-caffeoyltyramine and cannabisin B. Additionally, hempseed is rich in flavonoids, which contribute significantly to its antioxidant capabilities, as demonstrated by cell-free system assays such as ORAC and FRAP. Furthermore, the different fractions obtained from hempseed processing show different levels of antioxidant activity, with the coarsest fraction, dominated by hulls, showing the highest potency in this regard [388]. Hemp seed also contains a higher concentration of tocopherols than most vegetables. These tocopherols have antioxidant properties that effectively inhibit the oxidation of unsaturated

fatty acids [389]. Furthermore, the potential health benefits of cannabinoids, including CBD, have been explored in relation to various conditions such as Alzheimer's disease, cancer, epilepsy, inflammatory diseases, Parkinson's disease, and amyotrophic lateral sclerosis, as observed in studies conducted in mice [390]. In general, preclinical studies indicate the beneficial effects of hempseed supplementation on improving the blood lipid profiles and levels of linoleic acid, α -linolenic acid, and γ -linolenic acid. These enhancements have been associated with improvements in metabolic syndrome and neurodegenerative diseases [391]. In human trials, while several studies have described the beneficial effects of hempseed oil on skin health, mental well-being, and neurological disorders, the effects of hempseed on cardiovascular disease require further verification. Existing studies have produced conflicting results, indicating the need for further research (revised by [386]).

The health benefits of foods made from sorghum grain lie in the fact that sorghum is a rich source of antioxidant compounds such as vitamins, as well as macro- and micro-nutrients, including phenolic acids, flavonoids, and sterols [392]. The high antioxidant activity of bioactive compounds in sorghum grain is mainly due to its polyphenolic composition. In particular, white sorghum flour contains almost twice as many polyphenols as red sorghum flour. Numerous studies have demonstrated the anti-obesogenic effects of sorghum. Extracts of this grain have been shown to significantly inhibit the differentiation and accumulation of triglycerides. In addition, sorghum-supplied diets have been shown to reduce body weight in rats [393]. Human clinical trials have been conducted with sorghum to investigate obesity-related parameters. Participants who consumed sorghum biscuits reported greater satiety than those who consumed wheat-based products [394]. Additionally, sorghum starch undergoes slower digestion compared to starch from other cereals because of the hard outer layer of the endosperm and the existence of tannins. As a result, findings from these investigations indicate the possible incorporation of sorghum into diets for weight control.

Buckwheat is rich in several bioactive compounds that, along with essential nutrients, contribute to positive health outcomes [51]. More than a hundred bioactive compounds have been identified in buckwheat [144], including high levels of flavonoids such as rutin (quercetin-3-d-rutinoside), epicatechin, and quercetin, which vary depending on the type and origin of the buckwheat [395–397]. Moreover, the level of phenolic compounds is highly dependent on the roasting process, which can affect the total content and lead to significant losses [398]. Although the available literature indicates that buckwheat supplementation may provide some benefit in lowering TC and glucose in the context of dyslipidaemia and type 2 diabetes, a recent systematic review and meta-analysis have shown a small association between buckwheat supplementation intervention and cardiovascular risk factors [399]. Therefore, future human trials are needed to unveil the beneficial effects of the bioactive compounds in buckwheat.

Teff, another gluten-free pseudo-cereal like quinoa and buckwheat, has attracted considerable research interest due to its many potential health benefits. The use of teff is an opportunity to address the nutritional shortcomings of current commercial gluten-free products. Rich in insoluble polysaccharides, as well as macro- and micro-nutrients, teff is being explored as a viable option for people with type 2 diabetes due to its low glycaemic index. Teff is also naturally high in iron, making it suitable for people with anaemia [400]. Teff also contains higher levels of minerals such as zinc, iron, and calcium than other grains [401]. Furthermore, ferulic acid, a phenolic acid compound known for its potent antioxidant properties, is a major component of teff, along with other bioactives such as flavones like apigenin-6,8-c-diglucoside and apigenin-8-c-glucosyl-7-O-glucosides [400].

Furthermore, many of these NUSs contain antinutritional compounds such as phytic acid and saponins, which have the potential to bind nutrients and reduce absorption in our bodies. However, the use of various processing techniques such as fermentation, sprouting, extrusion, and cooking can increase their bioavailability [375].

Finally, it is important to emphasize the importance of bioactive peptides derived from the NUSs. These peptides, typically 2 to 50 amino acids in length, remain inert within

the intact protein. However, when released by proteases during digestion or through external hydrolysis, they are absorbed by the intestinal tract, reach various organs, and modulate important biological activities. These activities include immunomodulatory, antioxidant, antihypertensive, hypolipidaemic, hypoglycaemic, and hypotensive effects. Such effects play a key role in metabolic diseases such as obesity, metabolic syndrome, or metabolic-associated fatty liver disease [402–405]

5. Future Prospects

In the context of global economic and climatic change, a deeper understanding of agricultural production intensification is required, and new opportunities and challenges must be identified to progress [406]. Following high-input resource-intensive farming systems, an innovative approach is needed to protect and enhance natural resources while increasing productivity. Emerging crops appear capable of ensuring future food security and sovereignty [407]. They contribute to climate change mitigation and adaptation, demonstrating resilience and adaptation to abiotic stress [54,408–410]. For instance, salinization and drought are two major factors altering conditions for crop growth. Quinoa, a facultative halophyte, can withstand high salinity levels and water scarcity [411], and amaranth enriches its nutritional content and antioxidant activity under stress conditions [412]. Furthermore, chia has proven to be a viable option in dry, sandy soil and high-temperature environments [412], growing successfully in water-limiting situations [413], as does desert truffle [41]. Additionally, these crops require lower inputs for productivity, resulting in the limited use of water, fertilizers, and pesticides [33,414,415]. This makes NUSs pivotal in reducing greenhouse gas emissions [416] and promoting the sustainable development and efficient management of natural resources such as water, soil, and air. Moreover, NUSs promote soil protection, aligning with the objectives of the EU policy (CAP 2023–27), aiming for a 20% reduction in fertilizer use by 2030 and the potential to preserve and improve soil health [412].

NUSs also hold the potential for phytoremediation, offering a feasible option to mitigate the effects of heavy metal stress. Different species exhibit varying degrees of heavy metal accumulation and tolerance mechanisms. Using these species as a phytoremediation approach is seen as an optimal solution for reclaiming soil contaminated with heavy metals, ensuring the safety of harvested crops for human consumption. Quinoa, for instance, has shown promising bioremediation potential for soil contaminated with heavy metals [417], while amaranth has been studied extensively for remediating cadmium (Cd)-contaminated agricultural soils [418].

Therefore, meeting the needs of a growing world population by 2050 will require more resources from intensive food production. Monocultures may be vulnerable to unpredictable climate change, potentially leading to severe productivity losses. As previously mentioned, crop diversification, especially through the rotation of underutilized crops with existing systems, can improve food security, disrupt disease and pest cycles, replenish soil nutrients, and diversify pollinator presence [407]. Additionally, the increasing demand for ingredients used in animal feed creates pressure on major crops. In line with this, NUSs can serve as alternative sources of energy and protein, reducing dependence on major crops and competition with human food purposes. These crops could play a crucial role in animal growth and productivity due to their nutritional qualities. For example, Amaranth leaves and grains have shown varied nutritional qualities depending on consumption by monogastric or ruminants [419–421]. Quinoa, with its protein-rich fodder, offers an opportunity to diversify its use beyond animal feed.

Thus, NUSs have the potential to play a crucial role in addressing the world's current food challenges. They offer sustainability, adaptability, and nutritional value that can contribute to food security, conserve agrobiodiversity, and make agricultural and agrifood value chains more resilient. Despite limited progress in the development of superior underutilized crops, greater efforts will be needed to integrate these crops into mainstream agriculture.

6. Conclusions

In conclusion, the exploration of emergent crops in the dynamic context of agriculture and food science presents a groundbreaking avenue towards sustainable cultivation and nutritional innovation, with profound implications for human health. This review has delved into the potential of utilizing NUSs in Mediterranean environments, shedding light on the manifold benefits of diversifying agricultural and food systems. The resilient nature, nutritional richness, and positive health impacts of NUS such as quinoa, amaranth, chia, moringa, buckwheat, millet, teff, hemp, and desert truffles have been thoroughly examined, emphasizing their adaptability to the changing climate. By embracing these crops, we unlocked novel opportunities for agriculture and functional food development in important susceptible agricultural areas such as the Mediterranean region, paving the way for a more resilient and adaptable food system. The analysis made extends beyond the agronomic and nutritional aspects, delving into the broader dimensions of environmental, biomedical, economic, and cultural significance. Promoting agricultural diversification emerges as a key strategy to enhance food system adaptability to evolving environmental conditions, fostering sustainability and resilience. Therefore, by integrating these crops into our agrifood systems, we not only bolster agriculture resilience but also enhance food quality, addressing a spectrum of dimensions crucial for sustainable development. In essence, this revision calls for a paradigm shift towards embracing emergent crops, highlighting their potential to redefine our approach to agriculture, food science, and health. The integration of neglected and underutilized species into our food systems represents a forward-thinking solution, steering us towards a more sustainable and resilient future.

Author Contributions: All authors have contributed to writing—original draft preparation. All authors have read and agreed to the published version of the manuscript.

Funding: The authors gratefully acknowledge the financial support received from the Agencia Estatal de Investigación—Ministerio de Ciencia e Innovación (MICINN, Spain) (RED2022-134382-T).

Acknowledgments: This paper is dedicated to the memory of Maria del Carmen Blazquez Blazquez, for her support and inspiration. All listed authors in the authorship are on behalf of NutriCrop Spanish Consortium/Group integrated by María Reguera Blázquez (coordinator) (maria.reguera@uam.es); Isaac Maestro Gaitán (isaac.maestro@uam.es); Ingrid Aguiló Aguayo (ingrid.aguiló@irta.cat); Gemma Echeverría (gemma.echeverría@irta.cat); Maribel Abadías (isabel.abadías@irta.cat); Nieves Aparicio Gutiérrez (apagutni@itacyl.es); Ana Belen Martín Diana (mardiaan@itacyl.es); Daniel Rico Bargues (daniel.rico@uva.es); José Moisés Laparra Llopis (moises.laparra@imdea.org); Antonio Carrillo Vico (vico@us.es); Patricia J Lardone (plardone@us.es); Ignacio Bejarano Hernando (ibejarano@us.es); Eduardo Ponce España (eponce-ibis@us.es); Cristina Soler Rivas (cristina.soler@uam.es); Francisco R. Marín Martín (francisco.marin@uam.es); Joan Casals Missio (joan.casals-missio@upc.edu); Ana Rivera Pinzano (ana.rivera@upc.edu); Sònia Campo Sanchez (soniacampo@fundaciomiquelagusti.cat); Justo Javier Pedroche Jiménez (j.pedroche@csic.es); Claudia Mónica Haros (cmharos@iata.csic.es); Javier Matías Prieto (javier.matias@juntaex.es); María José Rodríguez Gómez (mariajose.rodriguez@juntaex.es); Verónica Cruz Sobrado (veronica.cruz@juntaex.es); Patricia Calvo Magro (patricia.calvo@juntaex.es); Nieves Fernández García (nieves@cebas.csic.es); Enrique Olmos (eolmos@cebas.csic.es); Sara Fondevilla Aparicio (sfondevilla@ias.csic.es); Asunción Morte Gómez (amorte@um.es); Alfonso Navarro Ródenas (anr@um.es); José Eduardo Marqués Gálvez (joseduardo.marques@um.es); Felicidad Ronda Balbás (mfronda@uva.es); and Pedro Antonio Caballero Calvo (pedroantonio.caballero@uva.es).

Conflicts of Interest: The authors declare no conflicts of interest.

References

1. Ulian, T.; Diazgranados, M.; Pironon, S.; Padulosi, S.; Liu, U.; Davies, L.; Howes, M.J.R.; Borrell, J.S.; Ondo, I.; Pérez-Escobar, O.A.; et al. Unlocking plant resources to support food security and promote sustainable agriculture. *Plants People Planet* **2020**, *2*, 421–445. [[CrossRef](#)]
2. Kopittke, P.M.; Menzies, N.W.; Wang, P.; McKenna, B.A.; Lombi, E. Soil and the intensification of agriculture for global food security. *Environ. Int.* **2019**, *132*, 105078. [[CrossRef](#)] [[PubMed](#)]

3. Mustafa, M.A.; Mayes, S.; Massawe, F. Crop diversification through a wider use of underutilised crops: A strategy to ensure food and nutrition security in the face of climate change. In *Sustainable Solutions for Food Security: Combating Climate Change by Adaptation*; Springer: Berlin/Heidelberg, Germany, 2019; pp. 125–149. [\[CrossRef\]](#)
4. Pingali, P. Agricultural policy and nutrition outcomes—Getting beyond the preoccupation with staple grains. *Food Secur.* **2015**, *7*, 583–591. [\[CrossRef\]](#)
5. Sibhatu, K.T.; Krishna, V.V.; Qaim, M. Production diversity and dietary diversity in smallholder farm households. *Proc. Natl. Acad. Sci. USA* **2015**, *112*, 10657–10662. [\[CrossRef\]](#) [\[PubMed\]](#)
6. Harkness, C.; Areal, F.J.; Semenov, M.A.; Senapati, N.; Shield, I.F.; Bishop, J. Stability of farm income: The role of agricultural diversity and agri-environment scheme payments. *Agric. Syst.* **2021**, *187*, 103009. [\[CrossRef\]](#)
7. Ouda, S.; Zohry, A.E.-H. Climate Extremes and Crops. In *Climate-Smart Agriculture*; Springer: Berlin/Heidelberg, Germany, 2022; pp. 93–114. [\[CrossRef\]](#)
8. Liu, C.; Plaza-Bonilla, D.; Coulter, J.A.; Kutcher, H.R.; Beckie, H.J.; Wang, L.; Floc'h, J.-B.; Hamel, C.; Siddique, K.H.M.; Li, L.; et al. Diversifying crop rotations enhances agroecosystem services and resilience. *Adv. Agron.* **2022**, *173*, 299–335. [\[CrossRef\]](#)
9. Hellegers, P. Food security vulnerability due to trade dependencies on Russia and Ukraine. *Food Secur.* **2022**, *14*, 1503–1510. [\[CrossRef\]](#)
10. Kumar, L.; Chhogyel, N.; Gopalakrishnan, T.; Hasan, M.K.; Jayasinghe, S.L.; Kariyawasam, C.S.; Kogo, B.; Ratnayake, S. Climate change and future of agri-food production. In *Future Foods: Global Trends, Opportunities, and Sustainability Challenges*; Academic Press: Cambridge, MA, USA, 2022; pp. 49–79. [\[CrossRef\]](#)
11. Sands, D.C.; Morris, C.E.; Dratz, E.A.; Pilgeram, A. Elevating optimal human nutrition to a central goal of plant breeding and production of plant-based foods. *Plant Sci.* **2009**, *177*, 377–389. [\[CrossRef\]](#)
12. Mayes, S.; Massawe, F.J.; Alderson, P.G.; Roberts, J.A.; Azam-Ali, S.N.; Hermann, M. The potential for underutilized crops to improve security of food production. *J. Exp. Bot.* **2012**, *63*, 1075–1079. [\[CrossRef\]](#)
13. Lionello, P.; Malanotte-Rizzoli, P.; Boscolo, R.; Alpert, P.; Artale, V.; Li, L.; Luterbacher, J.; May, W.; Trigo, R.; Tsimplis, M.; et al. The Mediterranean climate: An overview of the main characteristics and issues. *Dev. Earth Environ. Sci.* **2006**, *4*, 1–26. [\[CrossRef\]](#)
14. Del Pozo, A.; Brunel-Saldias, N.; Engler, A.; Ortega-Farías, S.; Acevedo-Opazo, C.; Lobos, G.A.; Jara-Rojas, R.; Molina-Montenegro, M.A. Climate Change Impacts and Adaptation Strategies of Agriculture in Mediterranean-Climate Regions (MCRs). *Sustainability* **2019**, *11*, 2769. [\[CrossRef\]](#)
15. Subedi, B.; Poudel, A.; Aryal, S. The impact of climate change on insect pest biology and ecology: Implications for pest management strategies, crop production, and food security. *J. Agric. Food Res.* **2023**, *14*, 100733. [\[CrossRef\]](#)
16. Cantero-Martínez, C.; Gabiña, D. Evaluation of agricultural practices to improve efficiency and environment conservation in Mediterranean arid and semi-arid production systems: MEDRATE project. In *Options Méditerranéennes: Série A Séminaires Méditerranéens*; CIHEAM: Zaragoza, Spain, 2004; pp. 21–31.
17. Grigorieva, E.; Livenets, A.; Stelmakh, E. Adaptation of Agriculture to Climate Change: A Scoping Review. *Climate* **2023**, *11*, 202. [\[CrossRef\]](#)
18. Sood, S.; Malhotra, N.; Tripathi, K.; Laibach, N.; Rosero, A. Editorial: Food of the future: Underutilized foods. *Front. Nutr.* **2023**, *10*, 1307856. [\[CrossRef\]](#) [\[PubMed\]](#)
19. Tamburini, G.; Bommarco, R.; Wanger, T.C.; Kremen, C.; van der Heijden, M.G.A.; Liebman, M.; Hallin, S. Agricultural diversification promotes multiple ecosystem services without compromising yield. *Sci. Adv.* **2020**, *6*, eaba1715. [\[CrossRef\]](#) [\[PubMed\]](#)
20. Raneri, J.E.; Padulosi, S.; Meldrum, G.; King, O.I. Promoting Neglected and Underutilized Species to Boost Nutrition in LMICs. 2019. Available online: <https://cgspace.cgiar.org/server/api/core/bitstreams/b215c446-2370-481f-8876-7580640b26b0/content> (accessed on 9 March 2024).
21. Zeder, M.A.; Zeder, A. Domestication and early agriculture in the Mediterranean Basin: Origins, diffusion, and impact. *Proc. Natl. Acad. Sci. USA* **2008**, *105*, 11597–11604. [\[CrossRef\]](#) [\[PubMed\]](#)
22. Harlan, J.R. *Crops and Man*; American Society of Agronomy and Crop Science Society of America: Madison, WI, USA, 1975.
23. Doebley, J.F.; Gaut, B.S.; Smith, B.D. The molecular genetics of crop domestication. *Cell* **2006**, *127*, 1309–1321. [\[CrossRef\]](#) [\[PubMed\]](#)
24. Vincent, H.; Amri, A.; Castañeda-Álvarez, N.P.; Dempewolf, H.; Dullo, E.; Guarino, L.; Hole, D.; Mba, C.; Toledo, A.; Maxted, N. Modeling of crop wild relative species identifies areas globally for in situ conservation. *Commun. Biol.* **2019**, *2*, 136. [\[CrossRef\]](#) [\[PubMed\]](#)
25. Behre, K.-E. The history of rye cultivation in Europe. *Veg. Hist. Archaeobot.* **1992**, *1*, 141–156. [\[CrossRef\]](#)
26. Hancock, J.F. Evolution of European Agriculture. In *World Agriculture before and after 1492*; Hancock, J.F., Ed.; Springer International Publishing: Cham, Switzerland, 2022; pp. 19–29. [\[CrossRef\]](#)
27. Padulosi, S.; Heywood, V.; Hunter, D.; Jarvis, A. Underutilized species and climate change: Current status and outlook. In *Crop Adaptation to Climate Change*; Yadav, S.Y., Redden, R.J., Hatfield, J.L., Lotze-Campen, H., Hall, A.E., Eds.; Wiley: Hoboken, NJ, USA, 2011; pp. 507–521. [\[CrossRef\]](#)
28. ECPGR. Minor Cereals and Pseudocereals in Europe. Report of a Network Coordinating Group on Minor Crops. 1999. Available online: https://www.ecpgr.org/fileadmin/bioversity/publications/pdfs/607_Report_of_a_network_coordinating_group_on_minor_crops-1.pdf (accessed on 30 January 2024).
29. Wanner, H.; Pfister, C.; Neukom, R. The variable European Little Ice Age. *Quat. Sci. Rev.* **2022**, *287*, 107531. [\[CrossRef\]](#)

30. Degroot, D.; Anchukaitis, K.; Bauch, M.; Burnham, J.; Carnegy, F.; Cui, J.; de Luna, K.; Guzowski, P.; Hambrecht, G.; Huhtamaa, H.; et al. Towards a rigorous understanding of societal responses to climate change. *Nature* **2021**, *591*, 539–550. [\[CrossRef\]](#) [\[PubMed\]](#)
31. Zhang, M.Z.; Xu, J.P.; Callac, P.; Chen, M.Y.; Wu, Q.; Wach, M.; Mata, G.; Zhao, R.L. Insight into the evolutionary and domesticated history of the most widely cultivated mushroom *Agaricus bisporus* via mitogenome sequences of 361 global strains. *BMC Genom.* **2023**, *24*, 182. [\[CrossRef\]](#) [\[PubMed\]](#)
32. Shavit, E. The History of Desert Truffle Use. In *Desert Truffles*; Springer International Publishing AG: Berlin/Heidelberg, Germany, 2014; pp. 217–241. [\[CrossRef\]](#)
33. Morte, A.; Kagan-Zur, V.; Navarro-Ródenas, A.; Sitrit, Y. Cultivation of Desert Truffles—A Crop Suitable for Arid and Semi-Arid Zones. *Agronomy* **2021**, *11*, 1462. [\[CrossRef\]](#)
34. Morte, A.; Pérez-Gilabert, M.; Gutiérrez, A.; Arenas, F.; Marqués-Gálvez, J.E.; Bordallo, J.J.; Rodríguez, A.; Berná, L.M.; Lozano-Carrillo, C.; Navarro-Ródenas, A. Basic and applied research for desert truffle cultivation. In *Mycorrhiza—Eco-Physiology, Secondary Metabolites, Nanomaterials*, 4th ed.; Springer: Berlin/Heidelberg, Germany, 2017; pp. 23–42. [\[CrossRef\]](#)
35. Fischer, A.R.H.; Beers, P.J.; Van Latesteijn, H.; Andeweg, K.; Jacobsen, E.; Mommaas, H.; Van Trijp, H.C.M.; Veldkamp, A. Transforum system innovation towards sustainable food. A review. *Agron. Sustain. Dev.* **2012**, *32*, 595–608. [\[CrossRef\]](#)
36. Kakabouki, I.; Tataridas, A.; Mavroeidis, A.; Kousta, A.; Roussis, I.; Katsenios, N.; Efthimiadou, A.; Papastylianou, P. Introduction of alternative crops in the Mediterranean to satisfy EU Green Deal goals. A review. *Agron. Sustain. Dev.* **2021**, *41*, 71. [\[CrossRef\]](#)
37. Gitz, V.; Meybeck, A.; Lipper, L.; Young, C.D.; Braatz, S. *Climate Change and Food Security: Risks and Responses*; Food and Agriculture Organization of the United Nations (FAO) Report; FAO: Rome, Italy, 2016; Volume 110.
38. Lin, P.H.; Chao, Y.Y. Different Drought-Tolerant Mechanisms in Quinoa (*Chenopodium quinoa* Willd.) and Djulis (*Chenopodium formosanum* Koidz.) Based on Physiological Analysis. *Plants* **2021**, *10*, 2279. [\[CrossRef\]](#)
39. Assefa, Y.; Staggenborg, S.A.; Prasad, V.P.V. Grain Sorghum Water Requirement and Responses to Drought Stress: A Review. *Crop Manag.* **2010**, *9*, 1–11. [\[CrossRef\]](#)
40. Gill, A.R.; Loveys, B.R.; Cavagnaro, T.R.; Burton, R.A. The potential of industrial hemp (*Cannabis sativa* L.) as an emerging drought resistant fibre crop. *Plant Soil* **2023**, *493*, 7–16. [\[CrossRef\]](#)
41. Morte, A.; Navarro-Ródenas, A.; Nicolás, E. Physiological parameters of desert truffle mycorrhizal *Helianthemum almeriense* plants cultivated in orchards under water deficit conditions. *Symbiosis* **2010**, *52*, 133–139. [\[CrossRef\]](#)
42. Henzell, R.G.; Jordan, D.R. Grain Sorghum. In *Cereals*; Carena, M.J., Ed.; Springer: New York, NY, USA, 2009; pp. 183–197. [\[CrossRef\]](#)
43. Goel, K.; Kundu, P.; Gahlaut, V.; Sharma, P.; Kumar, A.; Thakur, S.; Verma, V.; Bhargava, B.; Chandora, R.; Zinta, G. Functional divergence of Heat Shock Factors (Hsfs) during heat stress and recovery at the tissue and developmental scales in C4 grain amaranth (*Amaranthus hypochondriacus*). *Front. Plant Sci.* **2023**, *14*, 1151057. [\[CrossRef\]](#)
44. Iqbal, S.; Basra, S.M.A.; Saddiq, M.S.; Yang, A.; Akhtar, S.S.; Jacobsen, S.-E. The Extraordinary Salt Tolerance of Quinoa. In *Emerging Research in Alternative Crops*; Hirich, A., Choukr-Allah, R., Ragab, R., Eds.; Springer: Cham, Switzerland, 2020; pp. 125–143. [\[CrossRef\]](#)
45. Mansour, M.M.F.; Emam, M.M.; Salama, K.H.A.; Morsy, A.A. Sorghum under saline conditions: Responses, tolerance mechanisms, and management strategies. *Planta* **2021**, *254*, 24. [\[CrossRef\]](#) [\[PubMed\]](#)
46. Kamenya, S.N.; Mikwa, E.O.; Song, B.; Odeny, D.A. Genetics and breeding for climate change in Orphan crops. *Theor. Appl. Genet.* **2021**, *134*, 1787. [\[CrossRef\]](#) [\[PubMed\]](#)
47. Geisen, S.; Krishnaswamy, K.; Myers, R. Physical and Structural Characterization of Underutilized Climate-Resilient Seed Grains: Millets, Sorghum, and Amaranth. *Front. Sustain. Food Syst.* **2021**, *5*, 599656. [\[CrossRef\]](#)
48. Pathan, S.; Ndunguru, G.; Clark, K.; Ayele, A.G. Yield and nutritional responses of quinoa (*Chenopodium quinoa* Willd.) genotypes to irrigated, rainfed, and drought-stress environments. *Front. Sustain. Food Syst.* **2023**, *7*, 1242187. [\[CrossRef\]](#)
49. Knodel, N. A Framework for Agricultural Decarbonisation: Environmental Assessment from Seed to Soil of a Cradle-To-Cradle Farm System with Industrial Hemp and Pyrogenic Carbon Capture & Storage. *SSRN Electron. J.* **2022**. [\[CrossRef\]](#)
50. Selwal, N.; Bedi, M.; Hamid, S.; Pujari, M. Buckwheat (*Fagopyrum esculentum*) Response and Tolerance to Abiotic Stress. In *Omics Approach to Manage Abiotic Stress in Cereals*; Springer: Singapore, 2022; pp. 575–597. [\[CrossRef\]](#)
51. Zamaratskaia, G.; Gerhardt, K.; Knicky, M.; Wendin, K. Buckwheat: An underutilized crop with attractive sensory qualities and health benefits. *Crit. Rev. Food Sci. Nutr.* **2023**, 1–16. [\[CrossRef\]](#)
52. Horn, L.; Shakela, N.; Mutorwa, M.K.; Naomab, E.; Kwaambwa, H.M. *Moringa oleifera* as a sustainable climate-smart solution to nutrition, disease prevention, and water treatment challenges: A review. *J. Agric. Food Res.* **2022**, *10*, 100397. [\[CrossRef\]](#)
53. Acevedo, M.; Pixley, K.; Zinyengere, N.; Meng, S.; Tufan, H.; Cichy, K.; Bizikova, L.; Isaacs, K.; Ghezzi-Kopel, K.; Porciello, J. A scoping review of adoption of climate-resilient crops by small-scale producers in low- and middle-income countries. *Nat. Plants* **2020**, *6*, 1231–1241. [\[CrossRef\]](#)
54. Angeli, V.; Silva, P.M.; Massuela, D.C.; Khan, M.W.; Hamar, A.; Khajehei, F.; Graeff-Hönninger, S.; Piatti, C. Quinoa (*Chenopodium quinoa* Willd.): An Overview of the Potentials of the “Golden Grain” and Socio-Economic and Environmental Aspects of Its Cultivation and Marketization. *Foods* **2020**, *9*, 216. [\[CrossRef\]](#)

55. Ahmadi, S.H.; Solgi, S.; Sepaskhah, A.R. Quinoa: A super or pseudo-super crop? Evidences from evapotranspiration, root growth, crop coefficients, and water productivity in a hot and semi-arid area under three planting densities. *Agric. Water Manag.* **2019**, *225*, 105784. [\[CrossRef\]](#)
56. Herman, S.; Marco, G.; Cecilia, B.; Alfonso, V.; Luis, M.; Cristián, V.; Sebastián, P.; Sebastián, A. Effect of water availability on growth, water use efficiency and omega 3 (ALA) content in two phenotypes of chia (*Salvia hispanica* L.) established in the arid Mediterranean zone of Chile. *Agric. Water Manag.* **2016**, *173*, 67–75. [\[CrossRef\]](#)
57. Rastogi, A.; Shukla, S. Amaranth: A new millennium crop of nutraceutical values. *Crit. Rev. Food Sci. Nutr.* **2013**, *53*, 109–125. [\[CrossRef\]](#) [\[PubMed\]](#)
58. Owaid, M.N. Bioecology and Uses of Desert Truffles (*Pezizales*) in the Middle East. *Walailak J. Sci. Technol. (WJST)* **2017**, *15*, 179–188. [\[CrossRef\]](#)
59. Hordofa, T.; Dirirsa, G.; Wondimu, T.; Tesfaye, A. Crop Water Requirement and Crop Coefficient of Tef (*Eragrostis tef*) in Central Rift Valley of Ethiopia. *J. Nat. Sci. Res.* **2020**, *11*, 34–39. [\[CrossRef\]](#)
60. Godino, M.; Arias, C.; Izquierdo, M.I. *Moringa oleifera*: Potential areas of cultivation on the Iberian Peninsula. *Acta Hort.* **2017**, *1158*, 405–412. [\[CrossRef\]](#)
61. Iqbal, S.; Basra, S.M.A.; Afzal, I.; Wahid, A. Exploring potential of well adapted quinoa lines for salt tolerance. *Int. J. Agric. Biol.* **2017**, *19*, 933–940. [\[CrossRef\]](#)
62. Dixit, N. Salinity Induced Antioxidant Defense in Roots of Industrial Hemp (IH: *Cannabis sativa* L.) for Fiber during Seed Germination. *Antioxidants* **2022**, *11*, 244. [\[CrossRef\]](#) [\[PubMed\]](#)
63. Estrada, Y.; Fernández-Ojeda, A.; Morales, B.; Egea-Fernández, J.M.; Flores, F.B.; Bolarín, M.C.; Egea, I. Unraveling the Strategies Used by the Underexploited Amaranth Species to Confront Salt Stress: Similarities and Differences with Quinoa Species. *Front. Plant Sci.* **2021**, *12*, 604481. [\[CrossRef\]](#) [\[PubMed\]](#)
64. Joshi, B.K. Buckwheat (*Fagopyrum esculentum* Moench and *F. tataricum* Gaertn.). In *Neglected and Underutilized Crops: Future Smart Food*; Elsevier: Amsterdam, The Netherlands, 2023; pp. 151–200. [\[CrossRef\]](#)
65. Afzal, I.; Haq, M.Z.U.; Ahmed, S.; Hirich, A.; Bazile, D. Challenges and Perspectives for Integrating Quinoa into the Agri-Food System. *Plants* **2023**, *12*, 3361. [\[CrossRef\]](#)
66. Christiansen, J.L.; Jacobsen, S.E.; Jørgensen, S.T. Photoperiodic effect on flowering and seed development in quinoa (*Chenopodium quinoa* Willd.). *Acta Agric. Scand. B Soil Plant Sci.* **2010**, *60*, 539–544. [\[CrossRef\]](#)
67. Rodríguez-Abello, D.C.; Navarro-Alberto, J.A.; Ramírez-Avilés, L.; Zamora-Bustillos, R. The effect of sowing time on the growth of chia (*Salvia hispanica* L.): What do nonlinear mixed models tell us about it? *PLoS ONE* **2018**, *13*, e0206582. [\[CrossRef\]](#) [\[PubMed\]](#)
68. Matías, J.; Rodríguez, M.J.; Cruz, V.; Calvo, P.; Reguera, M. Heat stress lowers yields, alters nutrient uptake and changes seed quality in quinoa grown under Mediterranean field conditions. *J. Agron. Crop Sci.* **2021**, *207*, jac.12495. [\[CrossRef\]](#)
69. Ben-Zeev, S.; Kirby, N.; Diehn, S.; Shtein, I.; Elbaum, R.; Saranga, Y. Unraveling the central role of root morphology and anatomy in lodging of tef (*Eragrostis tef*). *Plants People Planet* **2023**. early access. [\[CrossRef\]](#)
70. Todaro, V.; D'Oria, M.; Secci, D.; Zanini, A.; Tanda, M.G. Climate Change over the Mediterranean Region: Local Temperature and Precipitation Variations at Five Pilot Sites. *Water* **2022**, *14*, 2499. [\[CrossRef\]](#)
71. Wrigley, C.W.; Corke, H.; Walker, C.E. *Encyclopedia of Grain Science*; Elsevier: Oxford, UK, 2004; pp. 1–10. [\[CrossRef\]](#)
72. Taylor, J.R.N. Sorghum and Millets: Taxonomy, History, Distribution, and Production. In *Sorghum and Millets: Chemistry, Technology, and Nutritional Attributes*; Woodhead Publishing: Duxford, UK, 2019; pp. 1–21. [\[CrossRef\]](#)
73. Lutz, W.; Kc, S. Dimensions of global population projections: What do we know about future population trends and structures? *Philos. Trans. R. Soc. B Biol. Sci.* **2010**, *365*, 2779. [\[CrossRef\]](#) [\[PubMed\]](#)
74. Tian, Z.; Wang, J.W.; Li, J.; Han, B. Designing future crops: Challenges and strategies for sustainable agriculture. *Plant J.* **2021**, *105*, 1165–1178. [\[CrossRef\]](#) [\[PubMed\]](#)
75. Dugan, F.M.; Lupien, S.L.; Hu, J. Fungal plant pathogens associated with emerging crops in North America: A challenge for plant health professionals. *Plant Health Prog.* **2017**, *18*, 221–229. [\[CrossRef\]](#)
76. Tyl, C.; DeHaan, L.; Frels, K.; Bajgain, P.; Marks, M.D.; Anderson, J.A. Emerging Crops with Enhanced Ecosystem Services: Progress in Breeding and Processing for Food Use. *Cereal Foods World* **2020**, *65*, 1–8. [\[CrossRef\]](#)
77. Pais, D.F.; Marques, A.C.; Fuinhas, J.A. How to Promote Healthier and More Sustainable Food Choices: The Case of Portugal. *Sustainability* **2023**, *15*, 3868. [\[CrossRef\]](#)
78. Jordan, N.R.; Dorn, K.; Runck, B.; Ewing, P.; Williams, A.; Anderson, K.A.; Felice, L.; Haralson, K.; Goplen, J.; Altendorf, K.; et al. Sustainable commercialization of new crops for the agricultural bioeconomy. *Elementa* **2016**, *2016*, 81. [\[CrossRef\]](#)
79. Anuradha Kumari, M.; Zinta, G.; Chauhan, R.; Kumar, A.; Singh, S.; Singh, S. Genetic resources and breeding approaches for improvement of amaranth (*Amaranthus* spp.) and quinoa (*Chenopodium quinoa*). *Front. Nutr.* **2023**, *10*, 1129723. [\[CrossRef\]](#) [\[PubMed\]](#)
80. Calderón-González, Á.; Matías, J.; Cruz, V.; Molinero-Ruiz, L.; Fondevilla, S. Identification and Characterization of Sources of Resistance to *Peronospora variabilis* in Quinoa. *Agronomy* **2023**, *13*, 284. [\[CrossRef\]](#)
81. Kučka, M.; Ražná, K.; Čerteková, S.; Chňápek, M.; Mikolášová, L.; Gálová, Z.; Balážová, Z. Buckwheat—A Genomic, Transcriptomic and Proteomic View. *J. Microbiol. Biotechnol. Food Sci.* **2023**, *13*, e10059. [\[CrossRef\]](#)
82. Hassani, M.; Piechota, T.; Atamian, H.S. Prediction of Cultivation Areas for the Commercial and an Early Flowering Wild Accession of *Salvia hispanica* L. in the United States. *Agronomy* **2022**, *12*, 1651. [\[CrossRef\]](#)

83. Flajšman, M.; Ačko, D.K. Industrial hemp breeding and genetics. In *Industrial Hemp*; Academic Press: Cambridge, MA, USA, 2022; pp. 37–57.
84. Craine, E.B.; Davies, A.; Packer, D.; Miller, N.D.; Schmöckel, S.M.; Spalding, E.P.; Tester, M. A comprehensive characterization of agronomic and end-use quality phenotypes across a quinoa world core collection. *Front. Plant Sci.* **2023**, *14*, 1101547. [[CrossRef](#)]
85. Zurita-Silva, A.; Fuentes, F.; Zamora, P.; Jacobsen, S.E.; Schwember, A.R. Breeding quinoa (*Chenopodium quinoa* Willd.): Potential and perspectives. *Mol. Breed.* **2014**, *34*, 13–30. [[CrossRef](#)]
86. Jarvis, D.E.; Hoy, Y.S.; Lightfoot, D.J.; Schmöckel, S.M.; Li, B.; Borm, T.J.A.; Ohyanagi, K.; Mineta, K.; Michell, C.T.; Saber, N. The genome of *Chenopodium quinoa*. *Nature* **2017**, *542*, 307–312. [[CrossRef](#)]
87. Li, L.; Nie, M.; Lu, J.; He, C.; Wu, Q. Genome-wide identification, expression analysis and gene duplication analysis of Dof transcription factors between quinoa and its ancestral diploid sub-genome species reveal key CDF homologs involved in photoperiodic flowering regulation. *S. Afr. J. Bot.* **2023**, *162*, 545–558. [[CrossRef](#)]
88. Patiranage, D.S.R.; Rey, E.; Emrani, N.; Wellman, G.; Schmid, K.; Schmöckel, S.M.; Tester, M. Genome-wide association study in quinoa reveals selection pattern typical for crops with a short breeding history. *eLife* **2022**, *11*, e66873. [[CrossRef](#)] [[PubMed](#)]
89. Sandell, F.L.; Holzweber, T.; Street, N.R.; Dohm, J.C.; Himmelbauer, H. Genomic basis of seed colour in quinoa inferred from variant patterns using extreme gradient boosting. *Plant Biotechnol. J.* **2024**, *22*, 1312–1324. [[CrossRef](#)]
90. Bochicchio, R.; Philips, T.; Lovelli, S.; Labella, R.; Galgano, F.; Di Marisco, A.; Perniola, M.; Amato, M. Innovative Crop Productions for Healthy Food: The Case of Chia (*Salvia hispanica* L.). In *The Sustainability of Agro-Food and Natural Resource Systems in the Mediterranean Basin*; Springer: Berlin/Heidelberg, Germany, 2015; pp. 29–45. [[CrossRef](#)]
91. Cahill, J.P.; Provance, M.C. Genetics of qualitative traits in domesticated chia (*Salvia hispanica* L.). *J. Hered.* **2002**, *93*, 52–55. [[CrossRef](#)]
92. Zare, T.; Paril, J.F.; Barnett, E.M.; Kaur, P.; Appels, R.; Ebert, B.; Roessner, U.; Fournier-Level, A. Comparative genomics points to tandem duplications of SAD gene clusters as drivers of increased α -linolenic (ω -3) content in *S. hispanica* seeds. *Plant Genome* **2024**, *17*, e20430. [[CrossRef](#)] [[PubMed](#)]
93. Gupta, P.; Geniza, M.; Elser, J.; Al-Bader, N.; Baschieri, R.; Phillips, J.L.; Haq, E.; Preece, J.; Naithani, S.; Jaiswal, P. Reference genome of the nutrition-rich orphan crop chia (*Salvia hispanica*) and its implications for future breeding. *Front. Plant Sci.* **2023**, *14*, 1272966. [[CrossRef](#)] [[PubMed](#)]
94. Ponnamp, N.; Kumari, M.; Ganesan, S.; Barik, S.; Pattanaik, M.; Adamala, A.K.; Rao, V.K.; Rupa, T.R.; Varalakshmi, B.; Acharya, G.C. Breeding leafy amaranth (*Amaranthus* spp.) for white rust resistance. *S. Afr. J. Bot.* **2023**, *163*, 794–804. [[CrossRef](#)]
95. Dharajiya, D.T.; Trivedi, G.N.; Thakkar, N.J.; Pachchigar, K.P.; Teli, B.; Tiwari, K.K.; Blair, M.W. Genomics-Assisted Design of Biotic Stress Resistant Vegetable Amaranths. In *Genomic Designing for Biotic Stress Resistant Vegetable Crops*; Springer: Heidelberg, Germany, 2022; pp. 261–300. [[CrossRef](#)]
96. Lightfoot, D.J.; Jarvis, D.E.; Ramaraj, T.; Lee, R.; Jellen, E.N.; Maughan, P.J. Single-molecule sequencing and Hi-C-based proximity-guided assembly of amaranth (*Amaranthus hypochondriacus*) chromosomes provide insights into genome evolution. *BMC Biol.* **2017**, *15*, 74. [[CrossRef](#)] [[PubMed](#)]
97. Jamalluddin, N. Genetic Diversity Analysis and Trait Phenotyping for Drought Tolerance in Amaranth (*Amaranthus* spp.) Germplasm. Ph.D. Thesis, University of Nottingham Malaysia, Selangor, Malaysia, 2020.
98. Chen, J.; Liu, Y.; Liu, M.; Guo, W.; Wang, Y.; He, Q.; Chen, W.; Liao, Y.; Zhang, W.; Gao, Y.; et al. Pangenome analysis reveals genomic variations associated with domestication traits in broomcorn millet Check for updates. *Nat. Genet.* **2023**, *55*, 2243–2254. [[CrossRef](#)] [[PubMed](#)]
99. Luthar, Z.; Fabjan, P.; Mlinarič, K. Biotechnological methods for buckwheat breeding. *Plants* **2021**, *10*, 1547. [[CrossRef](#)] [[PubMed](#)]
100. Zhang, K.; He, M.; Fan, Y.; Zhao, H.; Gao, B.; Yang, K.; Li, F.; Tang, Y.; Gao, Q.; Lin, T.; et al. Resequencing of global Tartary buckwheat accessions reveals multiple domestication events and key loci associated with agronomic traits. *Genome Biol.* **2021**, *22*, 23. [[CrossRef](#)] [[PubMed](#)]
101. Zhang, L.; Li, X.; Ma, B.; Gao, Q.; Du, H.; Han, Y.; Li, Y.; Cao, Y.; Qi, M.; Zhu, Y.; et al. The Tartary Buckwheat Genome Provides Insights into Rutin Biosynthesis and Abiotic Stress Tolerance. *Mol. Plant* **2017**, *10*, 1224–1237. [[CrossRef](#)] [[PubMed](#)]
102. Zhou, M.; Sun, Z.; Ding, M.; Logacheva, M.D.; Kreft, I.; Wang, D.; Yan, M.; Shao, J.; Tang, Y.; Wu, Y.; et al. FtSAD2 and FtJAZ1 regulate activity of the FtMYB11 transcription repressor of the phenylpropanoid pathway in *Fagopyrum tataricum*. *New Phytol.* **2017**, *216*, 814–828. [[CrossRef](#)]
103. Shi, T.X.; Li, R.Y.; Zheng, R.; Chen, Q.F.; Li, H.Y.; Huang, J.; Zhu, L.W.; Liang, G.C. Mapping QTLs for 1000-grain weight and genes controlling hull type using SNP marker in Tartary buckwheat (*Fagopyrum tataricum*). *BMC Genom.* **2021**, *22*, 142. [[CrossRef](#)]
104. Zhao, H.; Wang, L.; Jia, Y.; Zhao, J.; Li, C.; Chen, H.; Wu, H.; Wu, Q. Accumulation of the bitter substance quercetin mediated by the overexpression of a novel seed-specific gene FtRDE2 in Tartary buckwheat. *Plant Physiol. Biochem.* **2024**, *207*, 108402. [[CrossRef](#)] [[PubMed](#)]
105. Naik, S.; Sudan, J.; Urwat, U.; Pakhtoon, M.M.; Bhat, B.; Sharma, V.; Sofi, P.A.; Shikari, A.B.; Bhat, B.A.; Sofi, N.R.; et al. Genome-wide SNP discovery and genotyping delineates potential QTLs underlying major yield-attributing traits in buckwheat. *Plant Genome* **2024**, *17*, e20427. [[CrossRef](#)] [[PubMed](#)]
106. Lai, D.; Zhang, K.; He, Y.; Fan, Y.; Li, W.; Shi, Y.; Gao, Y.; Huang, X.; He, J.; Zhao, H.; et al. Multi-omics identification of a key glycosyl hydrolase gene FtGH1 involved in rutin hydrolysis in Tartary buckwheat (*Fagopyrum tataricum*). *Plant Biotechnol. J.* **2023**, *22*, 1206–1223. [[CrossRef](#)]

107. Santra, D.K.; Khound, R.; Das, S. Proso millet (*Panicum miliaceum* L.) breeding: Progress, challenges and opportunities. *Adv. Plant Breed. Strateg. Cereals* **2019**, *5*, 223–257. [\[CrossRef\]](#)
108. Nagaraja, T.E.; Nandini, C.; Bhat, S.; Parveen, S.G. Artificial hybridization techniques in small millets—A review. *Front. Plant Sci.* **2023**, *14*, 1112117. [\[CrossRef\]](#)
109. Fukunaga, K.; Kawase, M. Crop Evolution of Foxtail Millet. *Plants* **2024**, *13*, 218. [\[CrossRef\]](#) [\[PubMed\]](#)
110. He, Q.; Wang, C.; He, Q.; Zhang, J.; Liang, H.; Lu, Z.; Kie, K.; Tang, S.; Zhou, Y.; Liu, B.; et al. A complete reference genome assembly for foxtail millet and Setaria-db, a comprehensive database for Setaria. *Mol. Plant* **2024**, *17*, 219–222. [\[CrossRef\]](#)
111. He, Q.; Tang, S.; Zhi, H.; Chen, J.; Zhang, J.; Liang, H.; Xie, K.; Tang, S.; Zhou, Y.; Liu, B.; et al. A graph-based genome and pan-genome variation of the model plant Setaria. *Nat. Genet.* **2023**, *55*, 1232–1242. [\[CrossRef\]](#) [\[PubMed\]](#)
112. Balamurugan, A.; Mallikarjuna, M.G.; Bansal, S.; Nayaka, S.C.; Rajashekara, H.; Chellapilla, T.S.; Prakash, G. Genome-wide identification and characterization of NBLRR genes in finger millet (*Eleusine coracana* L.) and their expression in response to *Magnaporthe grisea* infection. *BMC Plant Biol.* **2024**, *24*, 75. [\[CrossRef\]](#)
113. Habiyaemye, C.; Barth, V.; Highet, K.; Coffey, T.; Murphy, K.M. Phenotypic Responses of Twenty Diverse Proso Millet (*Panicum miliaceum* L.) Accessions to Irrigation. *Sustainability* **2017**, *9*, 389. [\[CrossRef\]](#)
114. Khound, R.; Sun, G.; Mural, R.V.; Schnable, J.C.; Santra, D.K. SNP discovery in proso millet (*Panicum miliaceum* L.) using low-pass genome sequencing. *Plant Direct* **2022**, *6*, e447. [\[CrossRef\]](#) [\[PubMed\]](#)
115. Magris, G.; Foria, S.; Ciani, S.; Santra, D.K.; Polenghi, O.; Cerne, V.; Morgante, M.; Di Gaspero, G. Targeted sequencing of the *Panicum miliaceum* gene space and genotyping of variant sites from population genetics studies, combined in a single assay, as a tool for broomcorn millet assisted breeding. *Euphytica* **2023**, *219*, 102. [\[CrossRef\]](#)
116. Boukail, S.; Macharia, M.; Miculan, M.; Masoni, A.; Calamai, A.; Palchetti, E.; Dell’Acqua, M. Genome wide association study of agronomic and seed traits in a world collection of proso millet (*Panicum miliaceum* L.). *BMC Plant Biol.* **2021**, *21*, 330. [\[CrossRef\]](#) [\[PubMed\]](#)
117. Mushtaq, N.U.; Alghamdi, K.M.; Saleem, S.; Shajar, F.; Tahir, I.; Bahieldin, A.; Rehman, R.U.; Hakeem, K.R. Selenate and selenite transporters in proso millet: Genome extensive detection and expression studies under salt stress and selenium. *Front. Plant Sci.* **2022**, *13*, 1060154. [\[CrossRef\]](#) [\[PubMed\]](#)
118. Wang, H.; Zhang, H.; Liu, J.; Ma, Q.; Wu, E.; Gao, J.; Yang, Q.; Feng, B. Transcriptome analysis reveals the mechanism of nitrogen fertilizers in starch synthesis and quality in waxy and non-waxy proso millet. *Carbohydr. Polym.* **2024**, *323*, 121372. [\[CrossRef\]](#)
119. Beyene, G.; Chauhan, R.D.; Villmer, J.; Husic, N.; Wang, N.; Gebre, E.; Girma, D.; Chanyalew, S.; Assefa, K.; Tabor, G.; et al. CRISPR/Cas9-mediated tetra-allelic mutation of the ‘Green Revolution’ SEMIDWARF-1 (SD-1) gene confers lodging resistance in tef (*Eragrostis tef*). *Plant Biotechnol. J.* **2022**, *20*, 1716. [\[CrossRef\]](#)
120. Woldeyohannes, A.B.; Desta, E.A.; Fadda, C.; Pè, M.E.; Dell’Acqua, M. Value of teff (*Eragrostis tef*) genetic resources to support breeding for conventional and smallholder farming: A review. *CABI Agric. Biosci.* **2022**, *3*, 27. [\[CrossRef\]](#)
121. VanBuren, R.; Man Wai, C.; Wang, X.; Pardo, J.; Yocca, A.E.; Wang, H.; Chaluvadi, S.R.; Han, G.; Bryant, D.; Edger, P.P.; et al. Exceptional subgenome stability and functional divergence in the allotetraploid Ethiopian cereal teff. *Nat. Commun.* **2020**, *11*, 884. [\[CrossRef\]](#) [\[PubMed\]](#)
122. Chanyalew, S.; Singh, H.; Tefera, H.; Sorrells, M. Molecular genetic map and QTL analysis of agronomic traits based on a *Eragrostis tef* × *E. pilosa* recombinant inbred population. *J. Genet. Breed.* **2005**, *59*, 53–66.
123. Yu, J.K.; Graznak, E.; Breseghello, F.; Tefera, H.; Sorrells, M.E. QTL mapping of agronomic traits in tef [*Eragrostis tef* (Zucc) Trotter]. *BMC Plant Biol.* **2007**, *7*, 30. [\[CrossRef\]](#)
124. Ligaba-Osena, A.; Salehin, M.; Numan, M.; Wang, X.; Choi, S.C.; Jima, D.; Bobay, L.M.; Guo, W. Genome-wide transcriptome analysis of the orphan crop tef (*Eragrostis tef* (Zucc.) Trotter) under long-term low calcium stress. *Sci. Rep.* **2022**, *12*, 19552. [\[CrossRef\]](#) [\[PubMed\]](#)
125. Haddadi, B.S.; Fang, R.; Girija, A.; Kattupalli, D.; Widdowson, E.; Beckmann, M.; Yadav, R.; Mur, L.A.J. Metabolomics targets tissue-specific responses in alleviating the negative effects of salinity in tef (*Eragrostis tef*) during germination. *Planta* **2023**, *258*, 67. [\[CrossRef\]](#)
126. Girija, A.; Han, J.; Corke, F.; Brook, J.; Doonan, J.; Yadav, R.; Jifar, H.; Mur, L.A.J. Elucidating drought responsive networks in tef (*Eragrostis tef*) using phenomic and metabolomic approaches. *Physiol. Plant* **2022**, *174*, e13597. [\[CrossRef\]](#)
127. Berenji, J.; Sikora, V.; Fournier, G.; Beherec, O. Genetics and selection of hemp. In *Hemp: Industrial Production and Uses*; CABI: London, UK, 2013; pp. 48–71. [\[CrossRef\]](#)
128. Barcaccia, G.; Palumbo, F.; Scariolo, F.; Vannozzi, A.; Borin, M.; Bona, S. Potentials and Challenges of Genomics for Breeding Cannabis Cultivars. *Front. Plant Sci.* **2020**, *11*, 573299. [\[CrossRef\]](#) [\[PubMed\]](#)
129. Weiblen, G.D.; Wenger, J.P.; Craft, K.J.; ElSohly, M.A.; Mehmedic, Z.; Treiber, E.L.; Marks, M.D. Gene duplication and divergence affecting drug content in *Cannabis sativa*. *New Phytol.* **2015**, *208*, 1241–1250. [\[CrossRef\]](#) [\[PubMed\]](#)
130. Laverty, K.U.; Stout, J.M.; Sullivan, M.J.; Shah, H.; Gill, N.; Holbrook, L.; Deikus, G.; Sebra, R.; Hughes, T.R.; Page, J.E.; et al. A physical and genetic map of *Cannabis sativa* identifies extensive rearrangements at the THC/CBD acid synthase loci. *Genome Res.* **2019**, *29*, 146–156. [\[CrossRef\]](#)
131. Soorni, A.; Fatahi, R.; Haak, D.C.; Salami, S.A.; Bombarely, A. Assessment of Genetic Diversity and Population Structure in Iranian Cannabis Germplasm. *Sci. Rep.* **2017**, *7*, 15668. [\[CrossRef\]](#) [\[PubMed\]](#)

132. Stack, G.M.; Cala, A.R.; Quade, M.A.; Toth, J.A.; Monserrate, L.A.; Wilkerson, D.G.; Carlson, C.H.; Mamerto, A.; Michael, T.P.; Crawford, S.; et al. Genetic mapping, identification, and characterization of a candidate susceptibility gene for powdery mildew in *Cannabis sativa* L. *Mol. Plant-Microbe Interact.* **2023**, *37*, 51–61. [\[CrossRef\]](#)
133. Wei, H.; Yang, Z.; Niyitanga, S.; Tao, A.; Xu, J.; Fang, P.; Lin, L.; Zhang, L.; Qi, J.; Ming, R.; et al. The reference genome of seed hemp (*Cannabis sativa*) provides new insights into fatty acid and vitamin E synthesis. *Plant Commun.* **2024**, *5*, 100718. [\[CrossRef\]](#)
134. Alavilli, H.; Poli, Y.; Verma, K.S.; Kumar, V.; Gupta, S.; Chaudhary, V.; Jyoti, A.; Sahi, S.V.; Kothari, S.L. Miracle Tree *Moringa oleifera*: Status of the Genetic Diversity, Breeding, In Vitro Propagation, and a Cogent Source of Commercial Functional Food and Non-Food Products. *Plants* **2022**, *11*, 3132. [\[CrossRef\]](#)
135. Shyamli, P.S.; Pradhan, S.; Panda, M.; Parida, A. De novo Whole-Genome Assembly of *Moringa oleifera* Helps Identify Genes Regulating Drought Stress Tolerance. *Front. Plant Sci.* **2021**, *12*, 766999. [\[CrossRef\]](#) [\[PubMed\]](#)
136. Shankar, R.; Mandal, S.; Rao, K.; Kalaivanan, D. Moringa Breeding: International Scenario. In Proceedings of the International Seminar on Exotic and Underutilized Horticultural Crops: Priorities & Emerging Trends, Bengaluru, India, 17–19 October 2023.
137. Pasha, S.N.; Shafi, K.M.; Joshi, A.G.; Meenakshi, I.; Harini, K.; Mahita, J.; Sajeevan, R.S.; Karpe, S.D.; Ghosh, P.; Nitish, S.; et al. The transcriptome enables the identification of candidate genes behind medicinal value of Drumstick tree (*Moringa oleifera*). *Genomics* **2020**, *112*, 621–628. [\[CrossRef\]](#) [\[PubMed\]](#)
138. Wang, L.; Zou, Q.; Wang, J.; Zhang, J.; Liu, Z.; Chen, X. Proteomic Profiles Reveal the Function of Different Vegetative Tissues of *Moringa oleifera*. *Protein J.* **2016**, *35*, 440–447. [\[CrossRef\]](#) [\[PubMed\]](#)
139. Burchi, F.; Fanzo, J.; Frison, E. The role of food and nutrition system approaches in tackling hidden hunger. *Int. J. Environ. Res. Public Health* **2011**, *8*, 358–373. [\[CrossRef\]](#) [\[PubMed\]](#)
140. Titcomb, T.J.; Tanumihardjo, S.A. Global Concerns with B Vitamin Statuses: Biofortification, Fortification, Hidden Hunger, Interactions, and Toxicity. *Compr. Rev. Food Sci. Food Saf.* **2019**, *18*, 1968–1984. [\[CrossRef\]](#) [\[PubMed\]](#)
141. El Bilali, H. Research on agro-food sustainability transitions: Where are food security and nutrition? *Food Secur.* **2019**, *11*, 559–577. [\[CrossRef\]](#)
142. Tadele, Z. Orphan crops: Their importance and the urgency of improvement. *Planta* **2019**, *250*, 677–694. [\[CrossRef\]](#)
143. Assogbadjo, A.E.; Chadare, F.J.; Manda, L.; Sinsin, B. A 20-Year Journey Through an Orphan African Baobab (*Adansonia digitata* L.) Towards Improved Food and Nutrition Security in Africa. *Front. Sustain. Food Syst.* **2021**, *5*, 675382. [\[CrossRef\]](#)
144. Huda, M.N.; Lu, S.; Jahan, T.; Ding, M.; Jha, R.; Zhang, K.; Zhang, W.; Georgiev, M.I.; Park, S.U.; Zhou, M. Treasure from garden: Bioactive compounds of buckwheat. *Food Chem.* **2021**, *335*, 127653. [\[CrossRef\]](#) [\[PubMed\]](#)
145. Janssen, F.; Pauly, A.; Rombouts, I.; Jansens, K.J.A.; Deleu, L.J.; Delcour, J.A. Proteins of Amaranth (*Amaranthus* spp.), Buckwheat (*Fagopyrum* spp.), and Quinoa (*Chenopodium* spp.): A Food Science and Technology Perspective. *Compr. Rev. Food Sci. Food Saf.* **2017**, *16*, 39–58. [\[CrossRef\]](#)
146. Palombini, S.V.; Claus, T.; Maruyama, S.A.; Gohara, A.K.; Souza, A.H.P.; de Souza, N.E.; Visentainer, J.V.; Gomes, S.T.M.; Matsushita, M. Evaluation of nutritional compounds in new amaranth and quinoa cultivars. *Food Sci. Technol.* **2013**, *33*, 339–344. [\[CrossRef\]](#)
147. Tang, Y.; Li, X.; Chen, P.X.; Zhang, B.; Liu, R.; Hernandez, M.; Draves, J.; Marccone, M.F.; Tsao, R. Assessing the fatty acid, carotenoid, and tocopherol compositions of amaranth and quinoa seeds grown in Ontario and their overall contribution to nutritional quality. *J. Agric. Food Chem.* **2016**, *64*, 1103–1110. [\[CrossRef\]](#) [\[PubMed\]](#)
148. Vega-Gálvez, A.; Miranda, M.; Vergara, J.; Uribe, E.; Puente, L.; Martínez, E.A. Nutrition facts and functional potential of quinoa (*Chenopodium quinoa* Willd.), an ancient Andean grain: A review. *J. Sci. Food Agric.* **2010**, *90*, 2541–2547. [\[CrossRef\]](#)
149. Jin, J.; Ohanenye, I.C.; Udenigwe, C.C. Buckwheat proteins: Functionality, safety, bioactivity, and prospects as alternative plant-based proteins in the food industry. *Crit. Rev. Food Sci. Nutr.* **2022**, *62*, 1752–1764. [\[CrossRef\]](#) [\[PubMed\]](#)
150. Agarwal, A.; Rizwana Tripathi, A.D.; Kumar, T.; Sharma, K.P.; Patel, S.K.S. Nutritional and Functional New Perspectives and Potential Health Benefits of Quinoa and Chia Seeds. *Antioxidants* **2023**, *12*, 1413. [\[CrossRef\]](#) [\[PubMed\]](#)
151. Da Silva, B.P.; Anunciação, P.C.; da Matyelka, J.C.S.; Della Lucia, C.M.; Martino, H.S.D.; Pinheiro-Sant’Ana, H.M. Chemical composition of Brazilian chia seeds grown in different places. *Food Chem.* **2017**, *221*, 1709–1716. [\[CrossRef\]](#) [\[PubMed\]](#)
152. Awol, S.M.; Kuyu, C.G.; Bereka, T.Y. Physicochemical stability, microbial growth, and sensory quality of teff flour as affected by packaging materials during storage. *LWT* **2023**, *189*, 115488. [\[CrossRef\]](#)
153. Awika, J.M. Sorghum: Its Unique Nutritional and Health-Promoting Attributes. In *Gluten-Free Ancient Grains: Cereals, Pseudocereals, and Legumes: Sustainable, Nutritious, and Health-Promoting Foods for the 21st Century*; Elsevier: Amsterdam, The Netherlands, 2017; pp. 21–54. [\[CrossRef\]](#)
154. Gharsallah, K.; Rezig, L.; Rajoka, M.S.R.; Mehwish, H.M.; Ali, M.A.; Chew, S.C. *Moringa oleifera*: Processing, phytochemical composition, and industrial applications. *S. Afr. J. Bot.* **2023**, *160*, 180–193. [\[CrossRef\]](#)
155. Khalifa, S.A.M.; Farag, M.A.; Yosri, N.; Sabir, J.S.M.; Saeed, A.; Al-Mousawi, S.M.; Taha, W.; Musharraf, S.G.; Patel, S.; El-Seedi, H.R.; et al. Truffles: From Islamic culture to chemistry, pharmacology, and food trends in recent times. *Trends Food Sci. Technol.* **2019**, *91*, 193–218. [\[CrossRef\]](#)
156. Olmos, E.; Roman-Garcia, I.; Reguera, M.; Mestanza, C.; Fernandez-Garcia, N. An update on the nutritional profiles of quinoa (*Chenopodium quinoa* Willd.), amaranth (*Amaranthus* spp.), and chia (*Salvia hispanica* L.), three key species with the potential to contribute to food security worldwide. *JSEA Rep.* **2022**, *2*, 591–602. [\[CrossRef\]](#)

157. Navruz-Varli, S.; Sanlier, N. Nutritional and health benefits of quinoa (*Chenopodium quinoa* Willd.). *J. Cereal Sci.* **2016**, *69*, 371–376. [CrossRef]
158. Villacrés, E.; Quelal, M.; Galarza, S.; Iza, D.; Silva, E. Nutritional Value and Bioactive Compounds of Leaves and Grains from Quinoa (*Chenopodium quinoa* Willd.). *Plants* **2022**, *11*, 213. [CrossRef] [PubMed]
159. Soriano-García, M.; Aguirre-Díaz, I.S.; Soriano-García, M.; Aguirre-Díaz, I.S. Nutritional Functional Value and Therapeutic Utilization of Amaranth. In *Nutritional Value of Amaranth*; IntechOpen: London, UK, 2019. [CrossRef]
160. Repo-Carrasco, R.; Espinoza, C.; Jacobsen, S.E. Nutritional value and use of the andean crops quinoa (*Chenopodium quinoa*) and kañiwa (*Chenopodium pallidicaule*). *Food Rev. Int.* **2003**, *19*, 179–189. [CrossRef]
161. Dogra, D. Common Buckwheat: Nutritional Profiling of Grains. *Asian J. Dairy Food Res.* **2019**, *38*, 333–340. [CrossRef]
162. Sofi, S.A.; Ahmed, N.; Farooq, A.; Rafiq, S.; Zargar, S.M.; Kamran, F.; Dar, T.A.; Mir, S.A.; Dar, B.N.; Khaneghah, A.M. Nutritional and bioactive characteristics of buckwheat, and its potential for developing gluten-free products: An updated overview. *Food Sci. Nutr.* **2023**, *11*, 2256. [CrossRef] [PubMed]
163. Vera-Cespedes, N.; Muñoz, L.A.; Rincón, M.Á.; Haros, C.M. Physico-Chemical and Nutritional Properties of Chia Seeds from Latin American Countries. *Foods* **2023**, *12*, 3013. [CrossRef]
164. Ravindran, G. Seed protein of millets: Amino acid composition, proteinase inhibitors and in-vitro protein digestibility. *Food Chem.* **1992**, *44*, 13–17. [CrossRef]
165. Teff: Nutrient Composition and Health Benefits. Ethiopia Strategy Support Program Working Paper 67. Nutritional Composition of Teff Grain. Available online: https://ebrary.ifpri.org/digital/api/collection/p15738coll2/id/128374/download+of+Teff+Grain,&rlz=1C1GCEU_esES1095ES1095&oq=Teff:nutrient+composition+and+health+benefits.+Ethiopia+Strategy+Support+Program+Working+Paper+67,+Nutritional+Composition+of+Teff+Grain,&gs_lcrp=EgZjaHJvbWUyBggAEEUYOTIGCAEQRRg60gEHMjQ5ajBqNKgCALACAA&sourceid=chrome&ie=UTF-8 (accessed on 22 February 2024).
166. Moyo, B.; Masika, P.J.; Hugo, A.; Muchenje, V. Nutritional characterization of Moringa (*Moringa oleifera* Lam.) leaves. *Afr. J. Biotechnol.* **2011**, *10*, 12925–12933. [CrossRef]
167. Oyeyinka, A.T.; Oyeyinka, S.A. *Moringa oleifera* as a food fortificant: Recent trends and prospects. *J. Saudi Soc. Agric. Sci.* **2018**, *17*, 127–136. [CrossRef]
168. Zhao, J.; Wang, D.; Wang, W.; Li, Y.; Sun, X. Nutritional and chemical composition of industrial hemp seeds. In *Industrial Hemp: Food and Nutraceutical Applications*; Academic Press: Cambridge, MA, USA, 2022; pp. 73–93. [CrossRef]
169. Gadallah, M.G.E.; Ashoush, I.S.; Gadallah, M.G.E.; Ashoush, I.S. Value Addition on Nutritional and Sensory Properties of Biscuit Using Desert Truffle (*Terfezia clavervyi*) Powder. *Food Nutr. Sci.* **2016**, *7*, 1171–1181. [CrossRef]
170. Bobkov, S. Biochemical and Technological Properties of Buckwheat Grains. In *Molecular Breeding and Nutritional Aspects of Buckwheat*; Elsevier: Amsterdam, The Netherlands, 2016; pp. 423–440. [CrossRef]
171. Rodríguez Gómez, M.J.; Maestro-Gaitán, I.; Calvo Magro, P.; Cruz Sobrado, V.; Reguera Blázquez, M.; Matías Prieto, J. Unique nutritional features that distinguish *Amaranthus cruentus* L. and *Chenopodium quinoa* Willd seeds. *Food Res. Int.* **2023**, *164*, 112160. [CrossRef] [PubMed]
172. Graziano, S.; Agrimonti, C.; Marmiroli, N.; Gulli, M. Utilisation and limitations of pseudocereals (quinoa, amaranth, and buckwheat) in food production: A review. *Trends Food Sci. Technol.* **2022**, *125*, 154–165. [CrossRef]
173. USDA—United States Department of Agriculture. *National Nutrient Database for Standard Reference Release*; USDA: Washington, DC, USA, 2015; p. 28.
174. Mattila, P.H.; Pihlava, J.M.; Hellström, J.; Nurmi, M.; Eurola, M.; Mäkinen, S.; Jalava, T.; Pihlanto, A. Contents of phytochemicals and antinutritional factors in commercial protein-rich plant products. *Food Qual. Saf.* **2018**, *2*, 213–219. [CrossRef]
175. Dziadek, K.; Kopeć, A.; Pastucha, E.; Piątkowska, E.; Leszczyńska, T.; Pisulewska, E.; Witkiewicz, R.; Francik, R. Basic chemical composition and bioactive compounds content in selected cultivars of buckwheat whole seeds, dehulled seeds and hulls. *J. Cereal Sci.* **2016**, *69*, 1–8. [CrossRef]
176. Khan, A.; Khan, N.A.; Bean, S.R.; Chen, J.; Xin, Z.; Jiao, Y. Variations in Total Protein and Amino Acids in the Sequenced Sorghum Mutant Library. *Plants* **2023**, *12*, 1662. [CrossRef] [PubMed]
177. Quan, Z.; Zhang, L.; Chang, W.; Ding, X.; Qian, J.; Tang, J. Determination and Analysis of Composition, Structure, and Properties of Teff Protein Fractions. *Foods* **2023**, *12*, 3965. [CrossRef] [PubMed]
178. Tejedor-Calvo, E.; Amara, K.; Reis, F.S.; Barros, L.; Martins, A.; Calhelha, R.C.; Venturini, M.E.; Blanco, D.; Redondo, D.; Marco, P.; et al. Chemical composition and evaluation of antioxidant, antimicrobial and antiproliferative activities of Tuber and Terfezia truffles. *Food Res. Int.* **2021**, *140*, 110071. [CrossRef] [PubMed]
179. Rodríguez Gómez, M.J.; Matías Prieto, J.; Cruz Sobrado, V.; Calvo Magro, P. Nutritional characterization of six quinoa (*Chenopodium quinoa* Willd) varieties cultivated in Southern Europe. *J. Food Compos. Anal.* **2021**, *99*, 103876. [CrossRef]
180. Abugoch James, L.E. Quinoa (*Chenopodium quinoa* Willd.): Composition, chemistry, nutritional, and functional properties. *Adv. Food Nutr. Res.* **2009**, *58*, 1–31. [CrossRef]
181. Pellegrini, M.; Lucas-Gonzales, R.; Ricci, A.; Fontecha, J.; Fernández-López, J.; Pérez-Álvarez, J.A.; Viuda-Martos, M. Chemical, fatty acid, polyphenolic profile, techno-functional and antioxidant properties of flours obtained from quinoa (*Chenopodium quinoa* Willd.) seeds. *Ind. Crops Prod.* **2018**, *111*, 38–46. [CrossRef]
182. Simopoulos, A.P. The importance of the omega-6/omega-3 fatty acid ratio in cardiovascular disease and other chronic diseases. *Exp. Biol. Med.* **2008**, *233*, 674–688. [CrossRef] [PubMed]

183. Tang, Y.; Li, X.; Chen, P.X.; Zhang, B.; Hernandez, M.; Zhang, H.; Marcone, M.F.; Liu, R.; Tsao, R. Characterisation of fatty acid, carotenoid, tocopherol/tocotrienol compositions and antioxidant activities in seeds of three *Chenopodium quinoa* Willd. genotypes. *Food Chem.* **2015**, *174*, 502–508. [\[CrossRef\]](#) [\[PubMed\]](#)
184. Simopoulos, A.P. The importance of the ratio of omega-6/omega-3 essential fatty acids. *Biomed. Pharmacother.* **2002**, *56*, 365–379. [\[CrossRef\]](#)
185. Ruan, J.; Zhou, Y.; Yan, J.; Zhou, M.; Woo, S.H.; Weng, W.; Cheng, J.; Zhang, K. Tartary Buckwheat: An Under-utilized Edible and Medicinal Herb for Food and Nutritional Security. *Food Rev. Int.* **2022**, *38*, 440–454. [\[CrossRef\]](#)
186. Gulpinar, A.R.; Erdogan Orhan, I.; Kan, A.; Senol, F.S.; Celik, S.A.; Kartal, M. Estimation of in vitro neuroprotective properties and quantification of rutin and fatty acids in buckwheat (*Fagopyrum esculentum* Moench) cultivated in Turkey. *Food Res. Int.* **2012**, *46*, 536–543. [\[CrossRef\]](#)
187. Shen, Y.; Zheng, L.; Jin, J.; Li, X.; Fu, J.; Wang, M.; Guan, Y.; Song, X. Phytochemical and biological characteristics of Mexican chia seed oil. *Molecules* **2018**, *23*, 3219. [\[CrossRef\]](#) [\[PubMed\]](#)
188. Kulczyński, B.; Kobus-Cisowska, J.; Taczanowski, M.; Kmiecik, D.; Gramza-Michałowska, A. The chemical composition and nutritional value of chia seeds—Current state of knowledge. *Nutrients* **2019**, *11*, 1242. [\[CrossRef\]](#)
189. Mehmood, S.; Orhan, I.; Ahsan, Z.; Aslan, S.; Gulfrac, M. Fatty acid composition of seed oil of different *Sorghum bicolor* varieties. *Food Chem.* **2008**, *109*, 855–859. [\[CrossRef\]](#) [\[PubMed\]](#)
190. Desta, K.T.; Choi, Y.M.; Shin, M.J.; Yoon, H.; Wang, X.; Lee, Y.; Yi, J.; Jeon, Y.A.; Lee, S. Comprehensive evaluation of nutritional components, bioactive metabolites, and antioxidant activities in diverse sorghum (*Sorghum bicolor* (L.) Moench) landraces. *Food Res. Int.* **2023**, *173*, 113390. [\[CrossRef\]](#)
191. Liu, K.S. Comparison of Lipid Content and Fatty Acid Composition and Their Distribution within Seeds of 5 Small Grain Species. *J. Food Sci.* **2011**, *76*, C334–C342. [\[CrossRef\]](#)
192. Zhang, W.; Xu, J.; Bennetzen, J.L.; Messing, J. Teff, an orphan cereal in the chloridoideae, provides insights into the evolution of storage proteins in grasses. *Genome Biol. Evol.* **2016**, *8*, 1712–1721. [\[CrossRef\]](#) [\[PubMed\]](#)
193. Yisak, H.; Yaya, E.E.; Chandravanshi, B.S.; Redi-Abshiro, M. GC-MS profiling of fatty acids and nutritional properties of the white and brown teff [*Eragrostis tef* (Zuccagni) Trotter] varieties cultivated in different parts of Ethiopia. *J. Food Compos. Anal.* **2022**, *107*, 104405. [\[CrossRef\]](#)
194. Ruttarattanamongkol, K.; Siebenhandl-Ehn, S.; Schreiner, M.; Petrasch, A.M. Pilot-scale supercritical carbon dioxide extraction, physico-chemical properties and profile characterization of *Moringa oleifera* seed oil in comparison with conventional extraction methods. *Ind. Crops Prod.* **2014**, *58*, 68–77. [\[CrossRef\]](#)
195. Lee, H.; Lee, H.; Nam, K.; Zahra, Z.; Zahra, Z.; Farooqi, M.Q.U. Potentials of truffles in nutritional and medicinal applications: A review. *Fungal Biol. Biotechnol.* **2020**, *7*, 9. [\[CrossRef\]](#)
196. Akyüz, M.; Kirbağ, S. Nutritive Value of Desert Truffles Species of Genera *Terfezia* and *Picoa* (Ascomycetes) from Arid and Semiarid Regions of Eastern Turkey. *Int. J. Med. Mushrooms* **2018**, *20*, 1097–1106. [\[CrossRef\]](#)
197. Ghazali, H.M.; Mohammed, A.S. *Moringa (Moringa oleifera) Seed Oil: Composition, Nutritional Aspects, and Health Attributes*. In *Nuts and Seeds in Health and Disease Prevention*; Elsevier: Amsterdam, The Netherlands, 2011; pp. 787–793. [\[CrossRef\]](#)
198. Zouboulis, C.C.; Hossini, A.M.; Hou, X.; Wang, C.; Weylandt, K.H.; Pietzner, A. Effects of *Moringa oleifera* Seed Oil on Cultured Human Sebocytes In Vitro and Comparison with Other Oil Types. *Int. J. Mol. Sci.* **2023**, *24*, 332. [\[CrossRef\]](#)
199. Ahmadou, F.; Bourais, I.; Aqil, Y.; Shariati, M.A.; Hlebová, M.; Martsynkovsky, S.; Nagdalian, A.; El Hajjaji, S. Physiochemical Characteristics and Fatty Acids Composition of *Moringa Oleifera* Oil of Far North Cameroon. *J. Microbiol. Biotechnol. Food Sci.* **2023**, *13*, e10250. [\[CrossRef\]](#)
200. Pereira, E.; Encina-Zelada, C.; Barros, L.; Gonzales-Barron, U.; Cadavez, V.; Ferreira, I.C. Chemical and nutritional characterization of *Chenopodium quinoa* Willd (quinoa) grains: A good alternative to nutritious food. *Food Chem.* **2019**, *280*, 110–114. [\[CrossRef\]](#)
201. Hathcock, J.N. *Vitamin and Mineral Safety*, 2nd ed.; Council for Responsible Nutrition (CRN): Washington, DC, USA, 2004.
202. Latham, M.C.; FAO. Nutrición humana en el mundo en desarrollo. In *De las Naciones Unidas para la Agricultura y la Alimentación*; FAO: Rome, Italy, 2002.
203. Schmidt, D.; Verruma-Bernardi, M.R.; Forti, V.A.; Borges, M.T.M.R. Quinoa and Amaranth as Functional Foods: A Review. *Food Rev. Int.* **2023**, *39*, 2277–2296. [\[CrossRef\]](#)
204. Ramírez-Ojeda, A.M.; Moreno-Rojas, R.; Cámara-Martos, F. Mineral and trace element content in legumes (lentils, chickpeas and beans): Bioaccessibility and probabilistic assessment of the dietary intake. *J. Food Compos. Anal.* **2018**, *73*, 17–28. [\[CrossRef\]](#)
205. Zhu, F. Chemical composition and health effects of Tartary buckwheat. *Food Chem.* **2016**, *203*, 231–245. [\[CrossRef\]](#)
206. Zhao, M.; Gou, J.; Zhang, K.; Ruan, J. Principal Components and Cluster Analysis of Trace Elements in Buckwheat Flour. *Foods* **2023**, *12*, 225. [\[CrossRef\]](#) [\[PubMed\]](#)
207. Huang, X.Y.; Zeller, F.J.; Huang, K.F.; Shi, T.X.; Chen, Q.F. Variation of major minerals and trace elements in seeds of tartary buckwheat (*Fagopyrum tataricum* Gaertn.). *Genet. Resour. Crop Evol.* **2014**, *61*, 567–577. [\[CrossRef\]](#)
208. Ullah, R.; Nadeem, M.; Khalique, A.; Imran, M.; Mehmood, S.; Javid, A.; Hussain, J. Nutritional and therapeutic perspectives of Chia (*Salvia hispanica* L.): A review. *J. Food Sci. Technol.* **2016**, *53*, 1750–1758. [\[CrossRef\]](#) [\[PubMed\]](#)
209. Muñoz, L.A.; Cobos, A.; Diaz, O.; Aguilera, J.M. Chia Seed (*Salvia hispanica*): An Ancient Grain and a New Functional Food. *Food Rev. Int.* **2013**, *29*, 394–408. [\[CrossRef\]](#)

210. Adebo, J.A.; Kesa, H. Evaluation of nutritional and functional properties of anatomical parts of two sorghum (*Sorghum bicolor*) varieties. *Heliyon* **2023**, *9*, e17296. [\[CrossRef\]](#) [\[PubMed\]](#)
211. Espitia-Hernández, P.; González, M.L.C.; Ascacio-Valdés, J.A.; Dávila-Medina, D.; Flores-Naveda, A.; Silva, T.; Chacón, X.R.; Sepúlveda, L. Sorghum (*Sorghum bicolor* L.) as a potential source of bioactive substances and their biological properties. *Crit. Rev. Food Sci. Nutr.* **2022**, *62*, 2269–2280. [\[CrossRef\]](#)
212. Akansha, K.S.; Chauhan, E.S. Nutritional composition, physical characteristics and health benefits of Teff grain for human consumption: A critical review. *Pharma Innov. J.* **2018**, *7*, 03–07.
213. Reta, C.; Asmellash, T.; Atlabachew, M.; Mehari, B. Multielement analysis coupled with chemometrics modelling for geographical origin classification of teff [*Eragrostis tef* (Zuccagni) Trotter] grains from Amhara region. *BMC Chem.* **2023**, *17*, 50. [\[CrossRef\]](#)
214. Abebe, A.; Chandravanshi, B.S. Levels of essential and non-essential metals in the raw seeds and processed food (roasted seeds and bread) of maize/corn (*Zea mays* L.) cultivated in selected areas of Ethiopia. *Bull. Chem. Soc. Ethiop.* **2017**, *31*, 185–199. [\[CrossRef\]](#)
215. Assefa, K.; Cannarozzi, G.; Girma, D.; Kamies, R.; Chanyalew, S.; Plaza-Wüthrich, S.; Biosch, R.; Rindisbacher, A.; Rafudeen, S.; Tadele, Z. Genetic diversity in tef [*Eragrostis tef* (Zucc.) Trotter]. *Front. Plant Sci.* **2015**, *6*, 177. [\[CrossRef\]](#) [\[PubMed\]](#)
216. Ntshambiwa, K.T.; Seifu, E.; Mokhawa, G. Nutritional composition, bioactive components and antioxidant activity of *Moringa stenopetala* and *Moringa oleifera* leaves grown in Gaborone, Botswana. *Food Prod. Process. Nutr.* **2023**, *5*, 7. [\[CrossRef\]](#)
217. Agamou, A.; Armel, J.; Edith, F.N.; Moses, M.C. Influence of compounds contents and particle size on some functional properties of moringa oleifera leaves powders. *Asian Food Sci. J.* **2021**, *20*, 60–71.
218. Zhu, F. Dietary fiber polysaccharides of amaranth, buckwheat and quinoa grains: A review of chemical structure, biological functions and food uses. *Carbohydr. Polym.* **2020**, *248*, 116819. [\[CrossRef\]](#) [\[PubMed\]](#)
219. Dziedzic, K.; Kurek, S.; Mildner-Szkudlarz, S.; Kreft, I.; Walkowiak, J. Fatty acids profile, sterols, tocopherol and squalene content in *Fagopyrum tataricum* seed milling fractions. *J. Cereal Sci.* **2020**, *96*, 103118. [\[CrossRef\]](#)
220. Awulachew, M.T. *Investigating the Effects of Teff-Sorghum-Fenugreek Flour Blending Ratios on Quality Attributes of Injera*; Addis Ababa University: Addis Ababa, Ethiopia, 2022.
221. Sultana, S. Nutritional and functional properties of *Moringa oleifera*. *Metabol. Open* **2020**, *8*, 100061. [\[CrossRef\]](#)
222. Zniber, I.; Boukcim, H.; Khabar, L.; Ducouso, M.; Henkrar, F.; El Mouttaqi, A.; Hirich, A. Characterization of Desert Truffles in the Great Moroccan Sahara: A Review. *Environ. Sci. Proc.* **2022**, *16*, 55. [\[CrossRef\]](#)
223. Venskutonis, P.R.; Kraujalis, P. Nutritional Components of Amaranth Seeds and Vegetables: A Review on Composition, Properties, and Uses. *Compr. Rev. Food Sci. Food Saf.* **2013**, *12*, 381–412. [\[CrossRef\]](#)
224. Hrnič, M.K.; Ivanovski, M.; Cör, D.; Knez, Ž. Chia Seeds (*Salvia hispanica* L.): An overview-phytochemical profile, isolation methods, and application. *Molecules* **2020**, *25*, 11. [\[CrossRef\]](#)
225. Sidorova, Y.S.; Shipelin, V.A.; Petrov, N.A.; Zorin, S.N.; Mazo, V.K. Anxiolytic and Antioxidant Effect of Phytoecdysteroids and Polyphenols from *Chenopodium quinoa* on an In Vivo Restraint Stress Model. *Molecules* **2022**, *27*, 9003. [\[CrossRef\]](#)
226. Imran, M.; Salehi, B.; Sharifi-Rad, J.; Aslam Gondal, T.; Saeed, F.; Imran, A.; Shahbaz, M.; Tsouh Fokou, P.V.; Umair Arshad, M.; Khan, H.; et al. Kaempferol: A key emphasis to its anticancer potential. *Molecules* **2019**, *24*, 2277. [\[CrossRef\]](#) [\[PubMed\]](#)
227. Li, F.; Zhang, X.; Zheng, S.; Lu, K.; Zhao, G.; Ming, J. The composition, antioxidant and antiproliferative capacities of phenolic compounds extracted from tartary buckwheat bran [*Fagopyrum tartaricum* (L.) Gaerth]. *J. Funct. Foods* **2016**, *22*, 145–155. [\[CrossRef\]](#)
228. Pellegrini, M.; Lucas-Gonzalez, R.; Sayas-Barberá, E.; Fernández-López, J.; Pérez-Álvarez, J.A.; Viuda-Martos, M. Bioaccessibility of Phenolic Compounds and Antioxidant Capacity of Chia (*Salvia hispanica* L.) Seeds. *Plant Foods Hum. Nutr.* **2018**, *73*, 47–53. [\[CrossRef\]](#) [\[PubMed\]](#)
229. Apea-Bah, F.B.; Li, X.; Beta, T. Phenolic composition and antioxidant properties of cooked rice dyed with sorghum-leaf bio-colorants. *Foods* **2021**, *10*, 2058. [\[CrossRef\]](#) [\[PubMed\]](#)
230. Tanwar, R.; Panghal, A.; Chaudhary, G.; Kumari, A.; Chhikara, N. Nutritional, phytochemical and functional potential of sorghum: A review. *Food Chem. Adv.* **2023**, *3*, 100501. [\[CrossRef\]](#)
231. Ravisankar, S.; Abegaz, K.; Awika, J.M. Structural profile of soluble and bound phenolic compounds in teff (*Eragrostis tef*) reveals abundance of distinctly different flavones in white and brown varieties. *Food Chem.* **2018**, *263*, 265–274. [\[CrossRef\]](#) [\[PubMed\]](#)
232. Gil-Ramírez, A.; Pavo-Caballero, C.; Baeza, E.; Baenas, N.; Garcia-Viguera, C.; Marín, F.R.; Soler-Rivas, C. Mushrooms do not contain flavonoids. *J. Funct. Foods* **2016**, *25*, 1–13. [\[CrossRef\]](#)
233. Gil, J.V.; Esteban-Muñoz, A.; Fernández-Espinar, M.T. Changes in the polyphenolic profile and antioxidant activity of wheat bread after incorporating quinoa flour. *Antioxidants* **2022**, *11*, 33. [\[CrossRef\]](#)
234. Lis, B.; Jedrejek, D.; Rywaniak, J.; Soluch, A.; Stochmal, A.; Olas, B. Flavonoid Preparations from *Taraxacum officinale* L. Fruits—A Phytochemical, Antioxidant and Hemostasis Studies. *Molecules* **2020**, *25*, 5402. [\[CrossRef\]](#)
235. Rani, N.Z.A.; Husain, K.; Kumolosasi, E. *Moringa* genus: A review of phytochemistry and pharmacology. *Front. Pharmacol.* **2018**, *9*, 108. [\[CrossRef\]](#)
236. Pakade, V.; Cukrowska, E.; Chimuka, L. Comparison of antioxidant activity of *Moringa oleifera* and selected vegetables in South Africa. *S. Afr. J. Sci.* **2013**, *109*, 1–5. [\[CrossRef\]](#)
237. Ryan, E.; Galvin, K.; O'Connor, T.P.; Maguire, A.R.; O'Brien, N.M. Phytosterol, squalene, tocopherol content and fatty acid profile of selected seeds, grains, and legumes. *Plant Foods Hum. Nutr.* **2007**, *62*, 85–91. [\[CrossRef\]](#) [\[PubMed\]](#)

238. El-Gendy, A.N.G.; Pistelli, L.; Omer, E.A.; Elsayed, S.I.M.; Elshamy, A.I.; Hendawy, S.F. Growth, yield productivity, and oil composition of two Amaranthus species grown under two different environmental conditions in Egypt. *Crop Sci.* **2023**, *63*, 1472–1480. [\[CrossRef\]](#)
239. Gupta, R.; Sharma, A.K.; Dobhal, M.P.; Sharma, M.C.; Gupta, R.S. Antidiabetic and antioxidant potential of β -sitosterol in streptozotocin-induced experimental hyperglycemia. *J. Diabetes* **2011**, *3*, 29–37. [\[CrossRef\]](#)
240. Dziedzic, K.; Szwengiel, A.; Górecka, D.; Rudzińska, M.; Korczak, J.; Walkowiak, J. The effect of processing on the phytosterol content in buckwheat groats and by-products. *J. Cereal Sci.* **2016**, *69*, 25–31. [\[CrossRef\]](#)
241. Motyka, S.; Skala, E.; Ekiert, H.; Szopa, A. Health-promoting approaches of the use of chia seeds. *J. Funct. Foods* **2023**, *103*, 105480. [\[CrossRef\]](#)
242. El-Alfy, T.S.; Ezzat, S.M.; Sleem, A.A. Chemical and biological study of the seeds of *Eragrostis tef* (Zucc.) Trotter. *Nat. Prod. Res.* **2012**, *26*, 619–629. [\[CrossRef\]](#) [\[PubMed\]](#)
243. Weete, J.D.; Kulifaj, M.; Montant, C. Distribution of sterols in fungi. II. Brassicasterol in Tuber and Terfezia species. *Can. J. Microbiol.* **2011**, *31*, 1127–1130. [\[CrossRef\]](#)
244. Escribano, J.; Cabanes, J.; Atienzar, M.J.; Tremolada, M.I.; Pando, L.R.G.; Carmona, F.G.; Herrero, F.G. Characterization of betalains, saponins and antioxidant power in differently colored quinoa (*Chenopodium quinoa*) varieties. *Food Chem.* **2017**, *234*, 285–294. [\[CrossRef\]](#)
245. Gengatharan, A.; Dykes, G.A.; Choo, W.S. Betalains: Natural plant pigments with potential application in functional foods. *LWT* **2015**, *64*, 645–649. [\[CrossRef\]](#)
246. Gandía-Herrero, F.; García-Carmona, F. Biosynthesis of betalains: Yellow and violet plant pigments. *Trends Plant Sci.* **2013**, *18*, 334–343. [\[CrossRef\]](#) [\[PubMed\]](#)
247. Esatbeyoglu, T.; Wagner, A.E.; Schini-Kerth, V.B.; Rimbach, G. Betanin—A food colorant with biological activity. *Mol. Nutr. Food Res.* **2015**, *59*, 36–47. [\[CrossRef\]](#) [\[PubMed\]](#)
248. Hussain, M.I.; Farooq, M.; Syed, Q.A.; Ishaq, A.; Al-Ghamdi, A.A.; Hatamleh, A.A. Botany, nutritional value, phytochemical composition and biological activities of quinoa. *Plants* **2021**, *10*, 2258. [\[CrossRef\]](#) [\[PubMed\]](#)
249. Tang, Y.; Tsao, R. Phytochemicals in quinoa and amaranth grains and their antioxidant, anti-inflammatory, and potential health beneficial effects: A review. *Mol. Nutr. Food Res.* **2017**, *61*, 1600767. [\[CrossRef\]](#) [\[PubMed\]](#)
250. Repo-Carrasco-Valencia, R.; Hellström, J.K.; Pihlava, J.M.; Mattila, P.H. Flavonoids and other phenolic compounds in Andean indigenous grains: Quinoa (*Chenopodium quinoa*), kañiwa (*Chenopodium pallidicaule*) and kiwicha (*Amaranthus caudatus*). *Food Chem.* **2010**, *120*, 128–133. [\[CrossRef\]](#)
251. Haros, M.; Carlsson, N.G.; Almgren, A.; Larsson-Alminger, M.; Sandberg, A.S.; Andlid, T. Phytate degradation by human gut isolated *Bifidobacterium pseudocatenulatum* ATCC27919 and its probiotic potential. *Int. J. Food Microbiol.* **2009**, *135*, 7–14. [\[CrossRef\]](#)
252. Hurrell, R.F.; Reddy, M.B.; Juillerat, M.A.; Cook, J.D. Degradation of phytic acid in cereal porridges improves iron absorption by human subjects. *Am. J. Clin. Nutr.* **2003**, *77*, 1213–1219. [\[CrossRef\]](#)
253. Lopez, H.W.; Krespine, V.; Guy, G.; Messenger, A.; Demigne, C.; Remesy, C. Prolonged fermentation of whole wheat sourdough reduces phytate level and increases soluble magnesium. *J. Agric. Food Chem.* **2001**, *49*, 2657–2662. [\[CrossRef\]](#)
254. Vashishth, A.; Ram, S.; Beniwal, V. Cereal phytases and their importance in improvement of micronutrients bioavailability. *3 Biotech* **2017**, *7*, 42. [\[CrossRef\]](#)
255. García-Mantrana, I.; Monedero, V.; Haros, M. Application of phytases from bifidobacteria in the development of cereal-based products with amaranth. *Eur. Food Res. Technol.* **2014**, *238*, 853–862. [\[CrossRef\]](#)
256. Ruales, J.; Nair, B.M. Saponins, phytic acid, tannins and protease inhibitors in quinoa (*Chenopodium quinoa*, Willd) seeds. *Food Chem.* **1993**, *48*, 137–143. [\[CrossRef\]](#)
257. Gee, J.M.; Price, K.R.; Ridout, C.L.; Wortley, G.M.; Hurrell, R.F.; Johnson, I.T. Saponins of quinoa (*Chenopodium quinoa*): Effects of processing on their abundance in quinoa products and their biological effects on intestinal mucosal tissue. *JFA* **1993**, *63*, 201–209. [\[CrossRef\]](#)
258. Haros, C.M.; Schönlechner, R. *Pseudocereals: Chemistry and Technology*; John Wiley & Sons: Hoboken, NJ, USA, 2017.
259. Kårlund, A.; Paukkonen, I.; Gómez-Gallego, C.; Kolehmainen, M. Intestinal Exposure to Food-Derived Protease Inhibitors: Digestion Physiology- and Gut Health-Related Effects. *Healthcare* **2021**, *9*, 2. [\[CrossRef\]](#) [\[PubMed\]](#)
260. Vorster, J.; van der Westhuizen, W.; du Plessis, G.; Marais, D.; Sparvoli, F.; Cominelli, E.; Camilli, E.; Ferrari, M.; Le Donne, C.; Marconi, S.; et al. In order to lower the antinutritional activity of serine protease inhibitors, we need to understand their role in seed development. *Front. Plant Sci.* **2023**, *14*, 1252223. [\[CrossRef\]](#)
261. Gélinas, B.; Seguin, P. Oxalate in grain amaranth. *J. Agric. Food Chem.* **2007**, *55*, 4789–4794. [\[CrossRef\]](#) [\[PubMed\]](#)
262. Siener, R.; Hönow, R.; Seidler, A.; Voss, S.; Hesse, A. Oxalate contents of species of the Polygonaceae, Amaranthaceae and Chenopodiaceae families. *Food Chem.* **2006**, *98*, 220–224. [\[CrossRef\]](#)
263. Chai, W.; Liebman, M. Effect of Different Cooking Methods on Vegetable Oxalate Content. *J. Agric. Food Chem.* **2005**, *53*, 3027–3030. [\[CrossRef\]](#) [\[PubMed\]](#)
264. Miyake, K.; Tanaka, T.; McNeil, P.L. Lectin-based food poisoning: A new mechanism of protein toxicity. *PLoS ONE* **2007**, *2*. [\[CrossRef\]](#) [\[PubMed\]](#)
265. Hamid, L.L.; Al-Meani, S.A.L. Extraction and Purification of a Lectins from Iraqi Truffle (*Terfezia* sp.). *Egypt J. Chem.* **2021**, *64*, 2983–2987. [\[CrossRef\]](#)

266. Pompeu, D.G.; Mattioli, M.A.; Ribeiro, R.; Gonçalves, D.B.; Magalhães, J.; Marangoni, S.; Da Silva, J.A.; Granjeiro, P.A. Purification, partial characterization and antimicrobial activity of Lectin from *Chenopodium quinoa* seeds. *Food Sci. Technol.* **2015**, *35*, 696–703. [\[CrossRef\]](#)
267. Valadez-Vega, C.; Lugo-Magaña, O.; Figueroa-Hernández, C.; Bautista, M.; Betanzos-Cabrera, G.; Bernardino-Nicanor, A.; González-Amaro, R.M.; Alonso-Villegas, R.; Morales-González, J.A.; González-Cruz, L. Effects of Germination and Popping on the Anti-Nutritional Compounds and the Digestibility of *Amaranthus hypochondriacus* Seeds. *Foods* **2022**, *11*, 2075. [\[CrossRef\]](#) [\[PubMed\]](#)
268. Clements, W.T.; Lee, S.R.; Bloomer, R.J. Nitrate Ingestion: A Review of the Health and Physical Performance Effects. *Nutrients* **2014**, *6*, 5224–5264. [\[CrossRef\]](#) [\[PubMed\]](#)
269. Srujana, M.N.S.; Kumari, B.A.; Maheswari, K.U.; Devi, K.B.S.; Suneetha, W.J. Antioxidant and Anti Nutritional Composition of Germinated Quinoa (*Chenopodium quinoa* Willd). *Curr. J. Appl. Sci. Technol.* **2020**, *38*, 1–10. [\[CrossRef\]](#)
270. Oleszek, W.; Junkuszew, M.; Stochmal, A. Determination and Toxicity of Saponins from *Amaranthus cruentus* Seeds. *J. Agric. Food Chem.* **1999**, *47*, 3685–3687. [\[CrossRef\]](#) [\[PubMed\]](#)
271. Sriraj, P.; Toomsan, B.; Butnan, S. Effects of Neem Seed Extract on Nitrate and Oxalate Contents in Amaranth Fertilized with Mineral Fertilizer and Cricket Frass. *Horticulturae* **2022**, *8*, 898. [\[CrossRef\]](#)
272. Kumari, S.; Bhinder, S.; Singh, B.; Kaur, A. Physicochemical properties, non-nutrients and phenolic composition of germinated freeze-dried flours of foxtail millet, proso millet and common buckwheat. *J. Food Compos. Anal.* **2023**, *115*, 105043. [\[CrossRef\]](#)
273. Dunaevsky, Y.E.; Pavlukova, E.B.; Belozersky, M.A. Isolation and properties of anionic protease inhibitors from buckwheat seeds. *Biochem. Mol. Biol. Int.* **1996**, *40*, 199–208. [\[CrossRef\]](#) [\[PubMed\]](#)
274. Tsybina, T.A.; Dunaevsky, Y.E.; Musolyamov, A.K.; Egorov, T.A.; Belozersky, M.A. Cationic inhibitors of serine proteinases from buckwheat seeds. *Biochemistry* **2001**, *66*, 941–947. [\[CrossRef\]](#)
275. Dey, S.; Saxena, A.; Kumar, Y.; Maity, T.; Tarafdar, A. Understanding the Antinutritional Factors and Bioactive Compounds of Kodo Millet (*Paspalum scrobiculatum*) and Little Millet (*Panicum sumatrense*). *J. Food Qual.* **2022**, *2022*, 1578448. [\[CrossRef\]](#)
276. Udeh, H.O.; Duodu, K.G.; Jideani, A.I.O. Effect of malting period on physicochemical properties, minerals, and phytic acid of finger millet (*Eleusine coracana*) flour varieties. *Food Sci. Nutr.* **2018**, *6*, 1858–1869. [\[CrossRef\]](#)
277. Amalraj, A.; Pius, A. Influence of Oxalate, Phytate, Tannin, Dietary Fiber, and Cooking on Calcium Bioavailability of Commonly Consumed Cereals and Millets in India. *Cereal Chem.* **2015**, *92*, 389–394. [\[CrossRef\]](#)
278. Baye, K. Teff: Nutrient composition and health benefits. *Int. Food Policy Res. Inst.* **2014**, *67*, 1–17.
279. Karaçoban, İ.; Bilgiçli, N.; Yaver, E. Impact of Fermentation, Autoclaving and Phytase Treatment on the Antioxidant Properties and Quality of Teff Cookies. *Food Technol. Biotechnol.* **2023**, *61*, 328. [\[CrossRef\]](#)
280. Kataria, A.; Sharma, S.; Dar, B.N. Changes in phenolic compounds, antioxidant potential and antinutritional factors of Teff (*Eragrostis tef*) during different thermal processing methods. *Int. J. Food Sci. Technol.* **2021**, *57*, 6893–6902. [\[CrossRef\]](#)
281. Tiruneh, R. Minerals and Oxalate content of feed and water in relation with ruminant urolithiasis in Adea district, central Ethiopia. *Rev. Med. Vet.* **2004**, *155*, 272–277.
282. Kataria, A.; Sharma, S.; Singh, A.; Singh, B. Effect of hydrothermal and thermal processing on the antioxidative, antinutritional and functional characteristics of *Salvia hispanica*. *J. Food Meas. Charact.* **2022**, *16*, 332–343. [\[CrossRef\]](#)
283. Rayan, A.M. EHE Effects of dehulling, soaking, and cooking on the nutritional quality of *Moringa oleifera* seeds. *J. Agroaliment. Process. Technol.* **2016**, *22*, 156–165.
284. Saa, R.W.; Fombang Nig, E.; Radha, C.; Ndjantou, E.B.; Njintang Yanou, N. Effect of soaking, germination, and roasting on the proximate composition, antinutrient content, and some physicochemical properties of defatted *Moringa oleifera* seed flour. *J. Food Process Preserv.* **2022**, *46*, e16329. [\[CrossRef\]](#)
285. Aloo, S.O.; Kwame, F.O.; Oh, D.H. Identification of possible bioactive compounds and a comparative study on in vitro biological properties of whole hemp seed and stem. *Food Biosci.* **2023**, *51*, 102329. [\[CrossRef\]](#)
286. Alonso-Esteban, J.I.; Torija-Isasa, M.E.; de Sánchez-Mata, M.C. Mineral elements and related antinutrients, in whole and hulled hemp (*Cannabis sativa* L.) seeds. *J. Food Compos. Anal.* **2022**, *109*, 104516. [\[CrossRef\]](#)
287. Zhang, M.; Li, T.; Guo, G.; Liu, Z.; Hao, N. Production of Nattokinase from Hemp Seed Meal by Solid-State Fermentation and Improvement of Its Nutritional Quality. *Fermentation* **2023**, *9*, 469. [\[CrossRef\]](#)
288. Bouyahya Idrissi, O. Analysis of nutrient and antinutrient content of the truffle (*Tirmania pinoyi*) from Morocco. *Int. Food Res. J.* **2018**, *25*, 174–178.
289. FAO; IFAD; UNICEF; WFP; WHO. *The State of Food Security and Nutrition in the World 2023*; FAO: Rome, Italy, 2023. [\[CrossRef\]](#)
290. FAO. *The State of Food and Agriculture 2023—Revealing the True Cost of Food to Transform Agrifood Systems*; FAO: Rome, Italy, 2023. [\[CrossRef\]](#)
291. Hendrawan, V.S.A.; Komori, D.; Kim, W. Possible factors determining global-scale patterns of crop yield sensitivity to drought. *PLoS ONE* **2023**, *18*, e0281287. [\[CrossRef\]](#) [\[PubMed\]](#)
292. Chen, H.; Xu, B.; Wang, Y.; Li, W.; He, D.; Zhang, Y.; Zhang, X.; Xing, X. Emerging natural hemp seed proteins and their functions for nutraceutical applications. *Food Sci. Hum. Wellness* **2023**, *12*, 929–941. [\[CrossRef\]](#)
293. López, D.N.; Galante, M.; Robson, M.; Boeris, V.; Spelzini, D. Amaranth, quinoa and chia protein isolates: Physicochemical and structural properties. *Int. J. Biol. Macromol.* **2018**, *109*, 152–159. [\[CrossRef\]](#) [\[PubMed\]](#)

294. Ashaolu, T.J.; Le, T.D.; Suttikhana, I. Stability and bioactivity of peptides in food matrices based on processing conditions. *Food Res. Int.* **2023**, *168*, 112786. [[CrossRef](#)] [[PubMed](#)]
295. Chen, Z.; Yang, Q.; Yang, Y.; Zhong, H. The effects of high-pressure treatment on the structure, physicochemical properties and digestive property of starch—A review. *Int. J. Biol. Macromol.* **2023**, *244*, 125376. [[CrossRef](#)] [[PubMed](#)]
296. Garcia-Vaquero, M.; Tiwari, B.K. Innovative extraction technologies for high-value compounds. In *Pulse Foods: Processing, Quality and Nutraceutical Applications*; Academic Press: Cambridge, MA, USA, 2020. [[CrossRef](#)]
297. Taco, V.; Savarino, P.; Benali, S.; Villacrés, E.; Raquez, J.M.; Gerbaux, P.; Duez, P.; Nachtergaeel, A. Deep eutectic solvents for the extraction and stabilization of Ecuadorian quinoa (*Chenopodium quinoa* Willd.) saponins. *J. Clean. Prod.* **2022**, *363*, 132609. [[CrossRef](#)]
298. Ma, X.; Wang, Z.; Zheng, C.; Liu, C. A comprehensive review of bioactive compounds and processing technology of sesame seed. *Oil Crop Sci.* **2022**, *7*, 88–94. [[CrossRef](#)]
299. Abdelshafy, A.M.; Rashwan, A.K.; Osman, A.I. Potential food applications and biological activities of fermented quinoa: A review. *Trends Food Sci. Technol.* **2024**, *144*, 104339. [[CrossRef](#)]
300. Thakur, P.; Kumar, K.; Ahmed, N.; Chauhan, D.; Rizvi, Q.U.E.H.; Jan, S.; Singh, T.P.; Dhaliwal, H.S. Effect of soaking and germination treatments on nutritional, anti-nutritional, and bioactive properties of amaranth (*Amaranthus hypochondriacus* L.), quinoa (*Chenopodium quinoa* L.), and buckwheat (*Fagopyrum esculentum* L.). *Curr. Res. Food Sci.* **2021**, *4*, 917–925. [[CrossRef](#)] [[PubMed](#)]
301. D’Almeida, C.T.S.; Mameri, H.; Menezes, N.S.; Carvalho, C.W.P.; Queiroz, V.A.V.; Cameron, L.C.; Morel, M.-H.; Takeiti, C.Y.; Ferreira, M.S.L. Effect of extrusion and turmeric addition on phenolic compounds and kafirin properties in tannin and tannin-free sorghum. *Food Res. Int.* **2021**, *149*, 110663. [[CrossRef](#)] [[PubMed](#)]
302. Sharma, B.; Gujral, H.S. Influence of nutritional and antinutritional components on dough rheology and in vitro protein & starch digestibility of minor millets. *Food Chem.* **2019**, *299*, 125115. [[CrossRef](#)] [[PubMed](#)]
303. Cai, Y.-Q.; Gao, H.; Song, L.-M.; Tao, F.-Y.; Ji, X.-Y.; Yu, Y.; Cao, Y.; Tang, S.; Xue, P. Optimization of green deep eutectic solvent (DES) extraction of *Chenopodium quinoa* Willd. husks saponins by response surface methodology and their antioxidant activities. *RSC Adv.* **2023**, *13*, 29408–29418. [[CrossRef](#)]
304. Chen, Y.; Yuan, W.; Xu, Q.; Reddy, M.B. Neuroprotection of phytic acid in Parkinson’s and Alzheimer’s disease. *J. Funct. Foods* **2023**, *110*, 105856. [[CrossRef](#)]
305. Paes, L.T.; D’Almeida, C.T.d.S.; do Carmo, M.A.V.; da Silva Cruz, L.; Bubula de Souza, A.; Viana, L.M.; Gonçalves Maltarollo, V.; Martino, H.S.D.; Domingues de Almeida Lima, G.; Larraz Ferreira, M.S.; et al. Phenolic-rich extracts from toasted white and tannin sorghum flours have distinct profiles influencing their antioxidant, antiproliferative, anti-adhesive, anti-invasive, and antimalarial activities. *Food Res. Int.* **2024**, *176*, 113739. [[CrossRef](#)] [[PubMed](#)]
306. McDonnell, E. La despensa nacional: Quinoa and the Spatial Contradictions of Peru’s Gastronomic Revolution. *Lat. Am. Res. Rev.* **2023**, *59*, 84–104. [[CrossRef](#)]
307. Córdoba-Cerón, D.M.; Carranza-Saavedra, D.; Roa-Acosta, D.F.; Hoyos-Concha, J.L.; Solanilla-Duque, J.F. Physical and culinary analysis of long gluten-free extruded pasta based on high protein quinoa flour. *Front. Sustain. Food Syst.* **2022**, *6*, 1017324. [[CrossRef](#)]
308. Badia-Olmos, C.; Sánchez-García, J.; Laguna, L.; Zúñiga, E.; Mónica Haros, C.; Maria Andrés, A.; Tarrega, A. Flours from fermented lentil and quinoa grains as ingredients with new techno-functional properties. *Food Res. Int.* **2024**, *177*, 113915. [[CrossRef](#)]
309. Kierulf, A.; Whaley, J.; Liu, W.; Enayati, M.; Tan, C.; Perez-Herrera, M.; You, Z.; Abbaspourrad, A. Protein content of amaranth and quinoa starch plays a key role in their ability as Pickering emulsifiers. *Food Chem.* **2020**, *315*, 126246. [[CrossRef](#)]
310. Yalcin, S.; Atik, İ.; Atik, A. Effects of chia flour as a fat substitute on the physicochemical, nutritional and sensory properties of biscuits. *Int. J. Food Sci. Technol.* **2023**, *58*, 3760–3768. [[CrossRef](#)]
311. Momchilova, M.M.; Gradinarska-Ivanova Yordanov, D.G.; Zsivanovits, G.I. Microstructure and technological properties of cooked meat sausages prepared with emulsions of vegetable oils as substitutes for animal fat. *Food Res.* **2023**, *7*, 22–29. [[CrossRef](#)] [[PubMed](#)]
312. Fang, B.; Chang, L.; Ohm, J.B.; Chen, B.; Rao, J. Structural, functional properties, and volatile profile of hemp protein isolate as affected by extraction method: Alkaline extraction-isoelectric precipitation vs. salt extraction. *Food Chem.* **2023**, *405*, 135001. [[CrossRef](#)] [[PubMed](#)]
313. Zahari, I.; Ferawati, F.; Helstad, A.; Ahlström, C.; Östbring, K.; Rayner, M.; Purhagen, J.K. Development of High-Moisture Meat Analogues with Hemp and Soy Protein Using Extrusion Cooking. *Foods* **2020**, *9*, 772. [[CrossRef](#)] [[PubMed](#)]
314. George, T.T.; Oyenih, A.B.; Rautenbach, F.; Obilana, A.O. Characterization of Moringa oleifera Leaf Powder Extract Encapsulated in Maltodextrin and/or Gum Arabic Coatings. *Foods* **2021**, *10*, 3044. [[CrossRef](#)] [[PubMed](#)]
315. Ferreira, I.; Dias, T.; Mouazen, A.M.; Cruz, C. Using Science and Technology to Unveil the Hidden Delicacy *Terfezia arenaria*, a Desert Truffle. *Foods* **2023**, *12*, 3527. [[CrossRef](#)] [[PubMed](#)]
316. Agregán, R.; Guzel, N.; Guzel, M.; Bangar, S.P.; Zengin, G.; Kumar, M.; Lorenzo, J.M. The Effects of Processing Technologies on Nutritional and Anti-nutritional Properties of Pseudocereals and Minor Cereal. *Food Bioproc. Technol.* **2023**, *16*, 961–986. [[CrossRef](#)]
317. Bruno, M.C. Quinoa: Origins and Development. In *Encyclopedia of Global Archaeology*; Smith, C., Ed.; Springer: New York, NY, USA, 2014; pp. 6215–6220. [[CrossRef](#)]

318. Tanwar, B.; Goyal, A.; Irshaan, S.; Kumar, V.; Sihag, M.K.; Patel, A.; Kaur, I. *Whole Grains and Their Bioactives*; John Wiley & Sons: Hoboken, NJ, USA, 2019.
319. Zettel, V.; Hitzmann, B. Applications of chia (*Salvia hispanica* L.) in food products. *Trends Food Sci. Technol.* **2018**, *80*, 43–50. [\[CrossRef\]](#)
320. Levent, H. Effect of partial substitution of gluten-free flour mixtures with chia (*Salvia hispanica* L.) flour on quality of gluten-free noodles. *J. Food Sci. Technol.* **2017**, *54*, 1971–1978. [\[CrossRef\]](#)
321. Zettel, V.; Hitzmann, B. Chia (*Salvia hispanica* L.) as fat replacer in sweet pan breads. *Int. J. Food Sci. Technol.* **2016**, *51*, 1425–1432. [\[CrossRef\]](#)
322. Coelho, M.S.; de las Salas-Mellado, M.M. Effects of substituting chia (*Salvia hispanica* L.) flour or seeds for wheat flour on the quality of the bread. *LWT Food Sci. Technol.* **2015**, *60*, 729–736. [\[CrossRef\]](#)
323. Felisberto, M.H.F.; Wahanik, A.L.; Gomes-Ruffi, C.R.; Clerici, M.T.P.S.; Chang, Y.K.; Steel, C.J. Use of chia (*Salvia hispanica* L.) mucilage gel to reduce fat in pound cakes. *LWT Food Sci. Technol.* **2015**, *63*, 1049–1055. [\[CrossRef\]](#)
324. Senna, C.; Soares, L.; Egea, M.B.; Fernandes, S.S. The Techno-Functionality of Chia Seed and Its Fractions as Ingredients for Meat Analogs. *Molecules* **2024**, *29*, 440. [\[CrossRef\]](#)
325. Fernández-López, J.; Viuda-Martos, M.; Sayas-Barberá, M.E.; de Vera, C.N.-R.; Lucas-González, R.; Roldán-Verdú, A.; Botella-Martínez, C.; Pérez-Alvarez, J.A. Chia, quinoa, and their coproducts as potential antioxidants for the meat industry. *Plants* **2020**, *9*, 359. [\[CrossRef\]](#)
326. Nehra, M.; Gahlawat, S.K. *Chia and Quinoa: Superfoods for Health*; CRC Press: Boca Raton, FL, USA, 2022. [\[CrossRef\]](#)
327. Arendt, E.K.; Zannini, E. 13—Amaranth. In *Cereal Grains for the Food and Beverage Industries*; Arendt, E.K., Zannini, E., Eds.; Woodhead Publishing: Sawston, UK, 2013; pp. 439–473. [\[CrossRef\]](#)
328. Bvenura, C.; Kambizi, L. Future grain crops. In *Future Foods: Global Trends, Opportunities, and Sustainability Challenges*; Academic Press: Cambridge, MA, USA, 2022; pp. 81–105. [\[CrossRef\]](#)
329. Ma, Z.F.; Sun, Q.; Yap, P.S.; Khoo, H.E. Moringa: Phytopharmacological Properties and Its Potential as a Functional Food Ingredient. *Sustain. Food Sci. Compr. Approach* **2023**, *1–4*, 102–106. [\[CrossRef\]](#)
330. Wong, S.E.; Illingworth, K.A.; Siow, L.F. Moringa Proteins: Nutrition, Functionality, and Applications. In *Sustainable Protein Sources*; Academic Press: Cambridge, MA, USA, 2024; pp. 493–513. [\[CrossRef\]](#)
331. D’Auria, G.; Nitride, C.; Ferranti, P. *Moringa oleifera* Lam. Proteins: Properties and Food Applications. *Sustain. Food Sci. Compr. Approach* **2023**, *1–4*, 89–101. [\[CrossRef\]](#)
332. Vergara-Jimenez, M.; Almatrafi, M.M.; Fernandez, M.L. Bioactive Components in *Moringa oleifera* Leaves Protect against Chronic Disease. *Antioxidants* **2017**, *6*, 91. [\[CrossRef\]](#)
333. Milla, P.G.; Peñalver, R.; Nieto, G. Health Benefits of Uses and Applications of *Moringa oleifera* in Bakery Products. *Plants* **2021**, *10*, 318. [\[CrossRef\]](#)
334. Ibrahim, I.; Eldi, E.; Amir, A.; Bunyamin, B. Moderation of Good Corporate Governance: Earnings Management and Firm Value Against Company Performance. *Atestasi J. Ilm. Akunt.* **2022**, *5*, 496–510. [\[CrossRef\]](#)
335. Mohamed, F.A.E.-F.; Salama, H.H.; El-Sayed, S.M.; El-Sayed, H.S.; Zahran, H.A.-H. *Utilization of Natural Antimicrobial and Antioxidant of Moringa oleifera Leaves Extract in Manufacture of Cream Cheese*; CABI: Wallingford, UK, 2019.
336. Mashau, M.E.; Munandi, M.; Ramashia, S.E. Exploring the influence of *Moringa oleifera* leaves extract on the nutritional properties and shelf life of mutton patties during refrigerated storage. *CyTA J. Food* **2021**, *19*, 389–398. [\[CrossRef\]](#)
337. Cardines, P.H.F.; Baptista, A.T.A.; Gomes, R.G.; Bergamasco, R.; Vieira, A.M.S. *Moringa oleifera* seed extracts as promising natural thickening agents for food industry: Study of the thickening action in yogurt production. *LWT* **2018**, *97*, 39–44. [\[CrossRef\]](#)
338. Ogunsina, B.S.; Radha, C.; Indrani, D. Quality characteristics of bread and cookies enriched with debittered *Moringa oleifera* seed flour. *Int. J. Food Sci. Nutr.* **2011**, *62*, 185–194. [\[CrossRef\]](#)
339. Coello, K.E.; Peñas, E.; Martinez-Villaluenga, C.; Cartea, M.E.; Velasco, P.; Frias, J. Pasta products enriched with moringa sprout powder as nutritive dense foods with bioactive potential. *Food Chem.* **2021**, *360*, 130032. [\[CrossRef\]](#)
340. Aoki, H.; Nakatsuka-Mori, T.; Ueno, Y.; Nabeshima, Y.; Oyama, H. Analysis of functional ingredients of tempe-like fermented *Moringa oleifera* seeds (*Moringa* tempe) prepared with *Rhizopus* species. *J. Biosci. Bioeng.* **2023**, *135*, 306–312. [\[CrossRef\]](#)
341. Bankole, M.; Bodjrenou, S.; Honfo, F.; Codo, G.; Bodecker, J.; Termote, C.; Chadare, F. Hounkpatin W.A. Valorization of *Vigna radiata* (L.) Wilczek. and *Moringa oleifera* to improve food recipes of 6–23-month-old children in northern Benin. *J. Agric. Food Res.* **2023**, *13*, 100639. [\[CrossRef\]](#)
342. Affonfere, M.; Madode, Y.E.; Chadare, F.J.; Azokpota, P.; Hounhouigan, D.J. A dual food-to-food fortification with moringa (*Moringa oleifera* Lam.) leaf powder and baobab (*Adansonia digitata* L.) fruit pulp increases micronutrients solubility in sorghum porridge. *Sci. Afr.* **2022**, *16*, e01264. [\[CrossRef\]](#)
343. Anberbir, S.M.; Satheesh, N.; Abera, A.A.; Kassa, M.G.; Tenagashaw, M.W.; Asres, D.T.; Tiruneh, A.T.; Habtu, T.A.; Sadik, J.A.; Wudineh, T.A.; et al. Evaluation of nutritional composition, functional and pasting properties of pearl millet, teff, and buckwheat grain composite flour. *Appl. Food Res.* **2024**, *4*, 100390. [\[CrossRef\]](#)
344. Kumari, S.; Singh, B.; Kaur, A. Influence of malted buckwheat, foxtail and proso millet flour incorporation on the physicochemical, protein digestibility and antioxidant properties of gluten-free rice cookies. *Food Chem. Adv.* **2023**, *3*, 100557. [\[CrossRef\]](#)
345. Nalbandian, E.; Karkle, E.; Ganjyal, G.M. Physicochemical properties of buckwheat flours and their influence on focaccia bread quality. *Cereal Chem.* **2024**, *101*, 220–230. [\[CrossRef\]](#)

346. Auyeskhani, U.; Azgambetov, A.; Sadykov, T.; Dairabayeva, D.; Talamona, D.; Chan, M.Y. Reducing meat consumption in Central Asia through 3D printing of plant-based protein-enhanced alternatives—A mini review. *Front. Nutr.* **2024**, *10*, 1308836. [\[CrossRef\]](#)
347. Passi, P.; Kaur, K.; Kaur, J.; Kaul, S. Quality characterization, bread making properties and storage stability of foxtail millet incorporated bread ready-mix. *J. Stored Prod. Res.* **2023**, *104*, 102208. [\[CrossRef\]](#)
348. Divyashree, S.; Ramu, R.; Sreenivasa, M.Y. Evaluation of new candidate probiotic lactobacillus strains isolated from a traditional fermented food—Multigrain-millet dosa batter. *Food Biosci.* **2024**, *57*, 103450. [\[CrossRef\]](#)
349. Rani, M.; Dabur, R.S.; Bishnoi, S.; Jairath, G. Influence of storage on physico-chemical physiognomies of fermented whey cereal (pearl millet and moth bean) beverage. *J. Food Sci. Technol.* **2024**, *61*, 117–128. [\[CrossRef\]](#)
350. Naumenko, O.; Hetman, I.; Chyzyh, V.; Gunko, S.; Bal-Prylypko, L.; Bilko, M.; Tsentylo, L.; Lialyk, A.; Ivanytska, A.; Liashenko, S. Improving the Quality of Wheat Bread by Enriching Teff Flour. *East.-Eur. J. Enterp. Technol.* **2023**, *3*, 33–41. [\[CrossRef\]](#)
351. Hasani, Z.; Hosseini, H.; Shojaei-Aliabadi, S.; Moslehishad, M. Effects of Replacing Breadcrumbs and Soy Protein with Teff (*Eragrostis tef*) Flour on Physicochemical and Sensory Characteristics of Typical Hamburgers. *Iran. J. Nutr. Sci. Food Technol.* **2023**, *18*, 55–63.
352. Kebede Ali, M.; Abera, S.; Neme Tolesa, G. Optimization of extrusion cooking process parameters to develop teff (*Eragrostis* (Zucc) Trotter)-based products: Physical properties, functional properties, and sensory quality. *Cogent Food Agric.* **2023**, *9*, 2279705. [\[CrossRef\]](#)
353. Burton, R.A.; Andres, M.; Cole, M.; Cowley, J.M.; Augustin, M.A. Industrial hemp seed: From the field to value-added food ingredients. *J. Cannabis Res.* **2022**, *4*, 45. [\[CrossRef\]](#)
354. Korus, J.; Witzak, M.; Ziobro, R.; Juszczak, L. Hemp (*Cannabis sativa* subsp. *sativa*) flour and protein preparation as natural nutrients and structure forming agents in starch based gluten-free bread. *LWT* **2017**, *84*, 143–150. [\[CrossRef\]](#)
355. Ertaş, N.; Aslan, M. Antioxidant and physicochemical properties of cookies containing raw and roasted hemp flour. *Acta Sci. Pol. Technol. Aliment.* **2020**, *19*, 177–184. [\[CrossRef\]](#)
356. Nissen, L.; di Carlo, E.; Gianotti, A. Prebiotic potential of hemp blended drinks fermented by probiotics. *Food Res. Int.* **2020**, *131*, 109029. [\[CrossRef\]](#)
357. Norajit, K.; Gu, B.-J.; Ryu, G.-H. Effects of the addition of hemp powder on the physicochemical properties and energy bar qualities of extruded rice. *Food Chem.* **2011**, *129*, 1919–1925. [\[CrossRef\]](#)
358. Benucci, G.M.N.; Bonito, G.; Falini, L.B.; Bencivenga, M.; Donnini, D. Truffles, timber, food, and fuel: Sustainable approaches for multi-cropping truffles and economically important plants. In *Edible Ectomycorrhizal Mushrooms: Current Knowledge and Future Prospects*; Springer: Berlin/Heidelberg, Germany, 2012; pp. 265–280.
359. Liu, H.; Lv, M.; Peng, Q.; Shan, F.; Wang, M. Physicochemical and textural properties of tartary buckwheat starch after heat-moisture treatment at different moisture levels. *Starch Stärke* **2015**, *67*, 276–284. [\[CrossRef\]](#)
360. Xiao, Y.; Liu, H.; Wei, T.; Shen, J.; Wang, M. Differences in physicochemical properties and in vitro digestibility between tartary buckwheat flour and starch modified by heat-moisture treatment. *LWT* **2017**, *86*, 285–292. [\[CrossRef\]](#)
361. Almeida, R.L.J.; Santos, N.C.; Feitoza, J.V.F.; da Silva, G.M.; de Muniz, C.E.S.; da Eduardo, R.S.; de Ribeiro, V.H.A.; de Silva, V.M.A.; de Mota, M.M.A. Effect of heat-moisture treatment on the thermal, structural and morphological properties of Quinoa starch. *Carbohydr. Polym. Technol. Appl.* **2022**, *3*, 100192. [\[CrossRef\]](#)
362. Dong, J.; Huang, L.; Chen, W.; Zhu, Y.; Dun, B.; Shen, R. Effect of Heat-Moisture Treatments on Digestibility and Physicochemical Property of Whole Quinoa Flour. *Foods* **2021**, *10*, 3042. [\[CrossRef\]](#)
363. Vicente, A.; Villanueva, M.; Caballero, P.A.; Muñoz, J.M.; Ronda, F. Buckwheat grains treated with microwave radiation: Impact on the techno-functional, thermal, structural, and rheological properties of flour. *Food Hydrocoll.* **2023**, *137*, 108328. [\[CrossRef\]](#)
364. Vicente, A.; Villanueva, M.; Caballero, P.A.; Lazaridou, A.; Biliaderis, C.G.; Ronda, F. Flours from microwave-treated buckwheat grains improve the physical properties and nutritional quality of gluten-free bread. *Food Hydrocoll.* **2024**, *149*, 109644. [\[CrossRef\]](#)
365. Vicente, A.; Villanueva, M.; Caballero, P.A.; Muñoz, J.M.; Ronda, F. Microwave Modification of Quinoa Grains at Constant and Varying Water Content Modulates Changes in Structural and Physico-Chemical Properties of the Resulting Flours. *Foods* **2023**, *12*, 1421. [\[CrossRef\]](#)
366. Liu, H.; Guo, X.; Li, W.; Wang, X.; Lv, M.; Peng, Q.; Wang, M. Changes in physicochemical properties and in vitro digestibility of common buckwheat starch by heat-moisture treatment and annealing. *Carbohydr. Polym.* **2015**, *132*, 237–244. [\[CrossRef\]](#)
367. Siwatch, M.; Yadav, R.B.; Yadav, B.S. Annealing and heat-moisture treatment of amaranth starch: Effect on structural, pasting, and rheological properties. *J. Food Meas. Charact.* **2022**, *16*, 2323–2334. [\[CrossRef\]](#)
368. Náthia-Neves, G.; Villanueva, M.; Ronda, F. Impact of lipids on the functional, rheological, pasting and thermal properties of ultrasound-processed canary seed flours. *Food Hydrocoll.* **2024**, *150*, 109727. [\[CrossRef\]](#)
369. Ahmed, J.; Thomas, L.; Arfat, Y.A.; Joseph, A. Rheological, structural and functional properties of high-pressure treated quinoa starch in dispersions. *Carbohydr. Polym.* **2018**, *197*, 649–657. [\[CrossRef\]](#)
370. Liu, H.; Wang, L.; Cao, R.; Fan, H.; Wang, M. In vitro digestibility and changes in physicochemical and structural properties of common buckwheat starch affected by high hydrostatic pressure. *Carbohydr. Polym.* **2016**, *144*, 1–8. [\[CrossRef\]](#) [\[PubMed\]](#)

371. Li, G.; Zhu, F. Effect of high pressure on rheological and thermal properties of quinoa and maize starches. *Food Chem.* **2018**, *241*, 380–386. [[CrossRef](#)] [[PubMed](#)]
372. Vallons, K.J.R.; Ryan, L.A.M.; Arendt, E.K. Promoting structure formation by high pressure in gluten-free flours. *LWT Food Sci. Technol.* **2011**, *44*, 1672–1680. [[CrossRef](#)]
373. Gutiérrez, Á.L.; Rico, D.; Ronda, F.; Martín-Diana, A.B.; Caballero, P.A. Development of a gluten-free whole grain flour by combining soaking and high hydrostatic pressure treatments for enhancing functional, nutritional and bioactive properties. *J. Cereal Sci.* **2022**, *105*, 103458. [[CrossRef](#)]
374. Newman, T.V.; San-Juan-Rodriguez, A.; Parekh, N.; Swart, E.C.S.; Klein-Fedyshin, M.; Shrank, W.H.; Hernández, I. Impact of community pharmacist-led interventions in chronic disease management on clinical, utilization, and economic outcomes: An umbrella review. *Res. Social. Adm. Pharm.* **2020**, *16*, 1155–1165. [[CrossRef](#)]
375. Jan, N.; Hussain, S.Z.; Naseer, B.; Bhat, T.A. Amaranth and quinoa as potential nutraceuticals: A review of anti-nutritional factors, health benefits and their applications in food, medicinal and cosmetic sectors. *Food Chem. X* **2023**, *18*, 100687. [[CrossRef](#)]
376. Cox, A.L.; Eigenmann, P.A.; Sicherer, S.H. Clinical Relevance of Cross-Reactivity in Food Allergy. *J. Allergy Clin. Immunol. Pract.* **2021**, *9*, 82–99. [[CrossRef](#)]
377. Wang, T.; Zhang, X.; Zhou, N.; Shen, Y.; Li, B.; Chen, B.E.; Li, X. Association Between Omega-3 Fatty Acid Intake and Dyslipidemia: A Continuous Dose-Response Meta-Analysis of Randomized Controlled Trials. *J. Am. Heart Assoc.* **2023**, *12*, e029512. [[CrossRef](#)]
378. Shahidi, F.; Ambigaipalan, P. Omega-3 Polyunsaturated Fatty Acids and Their Health Benefits. *Annu. Rev. Food Sci. Technol.* **2018**, *9*, 345–381. [[CrossRef](#)]
379. Su, X.; Lu, G.; Ye, L.; Shi, R.; Zhu, M.; Yu, X.; Li, Z.; Jia, X.; Feng, L. *Moringa oleifera* Lam.: A comprehensive review on active components, health benefits and application. *RSC Adv.* **2023**, *13*, 24353–24384. [[CrossRef](#)]
380. Al Juhaimi, F.; Ghafoor, K.; Ahmed, I.A.M.; Babiker, E.E.; Özcan, M.M. Comparative study of mineral and oxidative status of *Sonchus oleraceus*, *Moringa oleifera* and *Moringa peregrina* leaves. *J. Food Meas. Charact.* **2017**, *11*, 1745–1751. [[CrossRef](#)]
381. Do, B.H.; Hoang, N.S.; Nguyen, T.P.T.; Ho, N.Q.C.; Le, T.L.; Doan, C.C. Phenolic Extraction of *Moringa oleifera* Leaves Induces Caspase-Dependent and Caspase-Independent Apoptosis through the Generation of Reactive Oxygen Species and the Activation of Intrinsic Mitochondrial Pathway in Human Melanoma Cells. *Nutr. Cancer* **2021**, *73*, 869–888. [[CrossRef](#)] [[PubMed](#)]
382. Kerdsoomboon, K.; Chumsawat, W.; Auesukaree, C. Effects of *Moringa oleifera* leaf extracts and its bioactive compound gallic acid on reducing toxicities of heavy metals and metalloids in *Saccharomyces cerevisiae*. *Chemosphere* **2021**, *270*, 128659. [[CrossRef](#)] [[PubMed](#)]
383. Wang, F.; Bao, Y.; Zhang, C.; Zhan, L.; Khan, W.; Siddiqua, S.; Ahmad, S.; Capanoglu, E.; Skalicka-Woźniak, K.; Zou, L. Bioactive components and anti-diabetic properties of *Moringa oleifera* Lam. *Crit. Rev. Food Sci. Nutr.* **2022**, *62*, 3873–3897. [[CrossRef](#)] [[PubMed](#)]
384. Wang, Y.; Ouyang, Q.; Chang, X.; Yang, M.; He, J.; Tian, Y.; Sheng, J. Anti-photoaging effects of flexible nanoliposomes encapsulated *Moringa oleifera* Lam. isothiocyanate in UVB-induced cell damage in HaCaT cells. *Drug Deliv.* **2022**, *29*, 871–881. [[CrossRef](#)] [[PubMed](#)]
385. Singhal, A.K.; Jarald, E.E.; Showkat, A.; Daud, A. In vitro evaluation of *Moringa oleifera* gum for colon-specific drug delivery. *Int. J. Pharm. Investig.* **2012**, *2*, 48–51. [[CrossRef](#)]
386. Leonard, W.; Zhang, P.; Ying, D.; Fang, Z. Hempseed in food industry: Nutritional value, health benefits, and industrial applications. *Compr. Rev. Food Sci. Food Saf.* **2020**, *19*, 282–308. [[CrossRef](#)] [[PubMed](#)]
387. Abedi, E.; Sahari, M.A. Long-chain polyunsaturated fatty acid sources and evaluation of their nutritional and functional properties. *Food Sci. Nutr.* **2014**, *2*, 443–463. [[CrossRef](#)]
388. Pojić, M.; Mišan, A.; Sakač, M.; Dapčević Hadnađev, T.; Šarić, B.; Milovanović, I.; Hadnađev, M. Characterization of byproducts originating from hemp oil processing. *J. Agric. Food Chem.* **2014**, *62*, 12436–12442. [[CrossRef](#)] [[PubMed](#)]
389. Schwartz, H.; Ollilainen, V.; Piironen, V.; Lampi, A.-M. Tocopherol, tocotrienol and plant sterol contents of vegetable oils and industrial fats. *J. Food Compos. Anal.* **2008**, *21*, 152–161. [[CrossRef](#)]
390. Pisanti, S.; Malfitano, A.M.; Ciaglia, E.; Lamberti, A.; Ranieri, R.; Cuomo, G.; Abate, M.; Faggiana, G.; Proto, M.C.; Fiore, D.; et al. Cannabidiol: State of the art and new challenges for therapeutic applications. *Pharmacol. Ther.* **2017**, *175*, 133–150. [[CrossRef](#)] [[PubMed](#)]
391. Li, X.Y.; Liu, Y.H.; Wang, B.; Chen, C.Y.; Zhang, H.M.; Kang, J.X. Identification of a sustainable two-plant diet that effectively prevents age-related metabolic syndrome and extends lifespan in aged mice. *J. Nutr. Biochem.* **2018**, *51*, 16–26. [[CrossRef](#)]
392. Frankowski, J.; Przybylska-Balcerek, A.; Stuper-Szablewska, K. Concentration of Pro-Health Compound of Sorghum Grain-Based Foods. *Foods* **2022**, *11*, 216. [[CrossRef](#)]
393. Amarakoon, D.; Lou, Z.; Lee, W.J.; Smolensky, D.; Lee, S.H. A mechanistic review: Potential chronic disease-preventive properties of sorghum. *J. Sci. Food Agric.* **2021**, *101*, 2641–2649. [[CrossRef](#)] [[PubMed](#)]
394. Berenji, J.; Dahlberg, J.A. Perspectives of Sorghum in Europe. *J. Agron. Crop Sci.* **2004**, *190*, 332–338. [[CrossRef](#)]
395. Luthar, Z.; Germ, M.; Likar, M.; Golob, A.; Vogel-Mikuš, K.; Pongrac, P.; Kušar, A.; Pravst, I.; Kreft, I. Breeding Buckwheat for Increased Levels of Rutin, Quercetin and Other Bioactive Compounds with Potential Antiviral Effects. *Plants* **2020**, *9*, 1638. [[CrossRef](#)] [[PubMed](#)]

396. Zielinska, D.; Turemko, M.; Kwiatkowski, J.; Zielinski, H. Evaluation of flavonoid contents and antioxidant capacity of the aerial parts of common and tartary buckwheat plants. *Molecules* **2012**, *17*, 9668–9682. [\[CrossRef\]](#)
397. Kalinova, J.P.; Vrchotova, N.; Triska, J. Contribution to the study of rutin stability in the achenes of Tartary buckwheat (*Fagopyrum tataricum*). *Food Chem.* **2018**, *258*, 314–320. [\[CrossRef\]](#)
398. Klepacka, J.; Najda, A. Effect of commercial processing on polyphenols and antioxidant activity of buckwheat seeds. *Int. J. Food Sci. Technol.* **2020**. [\[CrossRef\]](#)
399. Llanaj, E.; Ahanchi, N.S.; Dizdari, H.; Taneri, P.E.; Niehot, C.D.; Wehrli, F.; Khatami, F.; Raeisi-Dehkordi, H.; Kastrati, L.; Bano, A.; et al. Buckwheat and Cardiometabolic Health: A Systematic Review and Meta-Analysis. *J. Pers. Med.* **2022**, *12*, 1940. [\[CrossRef\]](#) [\[PubMed\]](#)
400. Gebru, Y.A.; Sbhatu, D.B.; Kim, K.-P. Nutritional Composition and Health Benefits of Teff (*Eragrostis tef* (Zucc.) Trotter). *J. Food Qual.* **2020**, *2020*, 9595086. [\[CrossRef\]](#)
401. Barretto, R.; Buenavista, R.M.; Rivera, J.L.; Wang, S.; Prasad, P.V.V.; Siliveru, K. Teff (*Eragrostis tef*) processing, utilization and future opportunities: A review. *Int. J. Food Sci. Technol.* **2021**, *56*, 3125–3137. [\[CrossRef\]](#)
402. Santos-Sánchez, G.; Álvarez-López, A.I.; Ponce-España, E.; Carrillo-Vico, A.; Bollati, C.; Bartolomei, M.; Lammi, C.; Chamorro, I.C. Hempseed (*Cannabis sativa*) protein hydrolysates: A valuable source of bioactive peptides with pleiotropic health-promoting effects. *Trends Food Sci. Technol.* **2022**, *127*, 303–318. [\[CrossRef\]](#)
403. Cruz-Chamorro, I.; Álvarez-Sánchez, N.; Santos-Sánchez, G.; Pedroche, J.; Fernández-Pachón, M.-S.; Millán, F.; Millán-Linares, M.C.; Lardone, P.J.; Bejarano, I.; Guerrero, J.M.; et al. Immunomodulatory and Antioxidant Properties of Wheat Gluten Protein Hydrolysates in Human Peripheral Blood Mononuclear Cells. *Nutrients* **2020**, *12*, 1673. [\[CrossRef\]](#) [\[PubMed\]](#)
404. Cruz-Chamorro, I.; Álvarez-Sánchez, N.; Álvarez-Ríos, A.I.; Santos-Sánchez, G.; Pedroche, J.; Millán, F.; Carrera Sánchez, C.; Fernández-Pachón, M.S.; Millán-Linares, M.C.; Martínez-López, A.; et al. Safety and Efficacy of a Beverage Containing Lupine Protein Hydrolysates on the Immune, Oxidative and Lipid Status in Healthy Subjects: An Intervention Study (the Lupine-1 Trial). *Mol. Nutr. Food Res.* **2021**, *65*, e2100139. [\[CrossRef\]](#) [\[PubMed\]](#)
405. Cruz-Chamorro, I.; Santos-Sánchez, G.; Álvarez-López, A.I.; Pedroche, J.; Lardone, P.J.; Arnoldi, A.; Carrillo-Vico, A. Pleiotropic biological effects of *Lupinus* spp. protein hydrolysates. *Trends Food Sci. Technol.* **2023**, *133*, 244–266. [\[CrossRef\]](#)
406. Gonzalez, C.G. Climate Change, Food Security, and Agrobiodiversity: Toward a Just, Resilient, and Sustainable Food System. *Fordham Environ. Law Rev.* **2010**, *22*, 493–522.
407. Singh, R.K.; Sreenivasulu, N.; Prasad, M. Potential of underutilized crops to introduce the nutritional diversity and achieve zero hunger. *Funct. Integr. Genom.* **2022**, *22*, 1459–1465. [\[CrossRef\]](#)
408. Ruiz, K.B.; Biondi, S.; Martínez, E.A.; Orsini, F.; Antognoni, F.; Jacobsen, S.E. Quinoa—A Model Crop for Understanding Salt-tolerance Mechanisms in Halophytes. *Plant Biosyst.* **2016**, *150*, 357–371. [\[CrossRef\]](#)
409. Hinojosa, L.; González, J.A.; Barrios-Masias, F.H.; Fuentes, F.; Murphy, K.M. Quinoa abiotic stress responses: A review. *Plants* **2018**, *7*, 106. [\[CrossRef\]](#)
410. Ruiz, K.B.; Biondi, S.; Osés, R.; Acuña-Rodríguez, I.S.; Antognoni, F.; Martínez-Mosqueira, E.A.; Coulibaly, A.; Canahua-Murillo, A.; Pinto, M.; Zurita-Silva, A.; et al. Quinoa biodiversity and sustainability for food security under climate change. A review. *Agron. Sustain. Dev.* **2014**, *34*, 349–359. [\[CrossRef\]](#)
411. Adolf, V.I.; Jacobsen, S.E.; Shabala, S. Salt tolerance mechanisms in quinoa (*Chenopodium quinoa* Willd.). *Environ. Exp. Bot.* **2013**, *92*, 43–54. [\[CrossRef\]](#)
412. Hussain, M.I.; Farooq, M.; Muscolo, A.; Rehman, A. Crop diversification and saline water irrigation as potential strategies to save freshwater resources and reclamation of marginal soils—A review. *Environ. Sci. Pollut. Res.* **2020**, *27*, 28695–28729. [\[CrossRef\]](#) [\[PubMed\]](#)
413. Harisha, C.B.; Narayanpur, V.B.; Rane, J.; Ganiger, V.M.; Prasanna, S.M.; Vishwanath, Y.C.; Reddi, S.G.; Halli, H.M.; Boraiah, K.M.; Basavaraj, P.S.; et al. Promising Bioregulators for Higher Water Productivity and Oil Quality of Chia under Deficit Irrigation in Semiarid Regions. *Plants* **2023**, *12*, 662. [\[CrossRef\]](#) [\[PubMed\]](#)
414. Zamani, Z.; Zeidali, E.; Alizadeh, H. Effect of drought stress and nitrogen chemical fertilizer on root properties and yield in three quinoa cultivars (*Chenopodium quinoa* Willd.). *Crop Sci. Res. Arid Reg.* **2024**, *5*, 487–500. [\[CrossRef\]](#)
415. Shabala, S.; Hariadi, Y.; Jacobsen, S.E. Genotypic difference in salinity tolerance in quinoa is determined by differential control of xylem Na⁺ loading and stomatal density. *J. Plant Physiol.* **2013**, *170*, 906–914. [\[CrossRef\]](#)
416. Sarkar, D.; Kar, S.K.; Chattopadhyay, A.; Rakshit, A.; Tripathi, V.K.; Dubey, P.K.; Abhilash, P.C. Low input sustainable agriculture: A viable climate-smart option for boosting food production in a warming world. *Ecol. Indic.* **2020**, *115*, 106412. [\[CrossRef\]](#)
417. Haseeb, M.; Iqbal, S.; Hafeez, M.B.; Baloch, H.; Zahra, N.; Mumtaz, M.; Ahmad, G.; Fatima, E.M.; Raza, S.; Raza, A.; et al. Characterizing of heavy metal accumulation, translocation and yield response traits of *Chenopodium quinoa*. *J. Agric. Food Res.* **2023**, *14*, 100741. [\[CrossRef\]](#)
418. Haller, H.; Pronoza, L.; Dyer, M.; Ahlgren, M.; Bergqvist, L.; Flores-Carmenate, G.; Jonsson, A. Phytoremediation of Heavy-Metal-Contaminated Soils: Capacity of Amaranth Plants to Extract Cadmium from Nutrient-Poor, Acidic Substrates. *Challenges* **2023**, *14*, 28. [\[CrossRef\]](#)
419. Nazeer, S.; Yaman Firincioglu, S. Amaranth in Animal Nutrition. *J. Agric. Food Environ. Anim. Sci.* **2022**, *3*, 195–211.

420. Aderibigbe, O.R.; Ezekiel, O.O.; Owolade, S.O.; Korese, J.K.; Sturm, B.; Hensel, O. Exploring the potentials of underutilized grain amaranth (*Amaranthus* spp.) along the value chain for food and nutrition security: A review. *Crit. Rev. Food Sci. Nutr.* **2022**, *62*, 656–669. [[CrossRef](#)] [[PubMed](#)]
421. Ngugi, C.C.; Oyoo-Okoth, E.; Manyala, J.O.; Fitzsimmons, K.; Kimotho, A. Characterization of the nutritional quality of amaranth leaf protein concentrates and suitability of fish meal replacement in Nile tilapia feeds. *Aquac. Rep.* **2017**, *5*, 62–69. [[CrossRef](#)]

Disclaimer/Publisher’s Note: The statements, opinions and data contained in all publications are solely those of the individual author(s) and contributor(s) and not of MDPI and/or the editor(s). MDPI and/or the editor(s) disclaim responsibility for any injury to people or property resulting from any ideas, methods, instructions or products referred to in the content.